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Hatzfeld

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(54) **PLANTS HAVING ENHANCED
YIELD-RELATED TRAITS AND A METHOD
FOR MAKING THE SAME**

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(58) **Field of Classification Search**
None
See application file for complete search history.

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(57) **ABSTRACT**

The present invention relates generally to the field of molecular biology and concerns a method for enhancing various economically important yield-related traits in plants. More specifically, the present invention concerns a method for enhancing yield-related traits in plants by modulating expression in a plant of a nucleic acid encoding an ASPAT (Aspartate AminoTransferase) polypeptide. The present invention also concerns plants having modulated expression of a nucleic acid encoding an ASPAT polypeptide, which plants have enhanced yield-related traits relative to control plants. The invention also provides hitherto unknown ASPAT-encoding nucleic acids and constructs comprising the same, useful in performing the methods of the invention. Furthermore, the present invention relates generally to the field of molecular biology and concerns a method for increasing various plant yield-related traits by increasing expression in a plant of a nucleic acid sequence encoding a MYB91 like transcription factor (MYB91) polypeptide. The present invention also concerns plants having increased expression of a nucleic acid sequence encoding an MYB91 polypeptide, which plants have increased yield-related traits relative to control plants. The invention additionally relates to nucleic acid sequences, nucleic acid constructs, vectors and plants containing said nucleic acid sequences. Even furthermore, the present invention relates generally to the field of molecular biology and concerns a method for improving various plant growth characteristics by modulating expression in a plant of a nucleic acid encoding a GASA (Gibberellic Acid-Stimulated *Arabidopsis*). The present invention also concerns plants having modulated expression of a nucleic acid encoding a GASA, which plants have improved growth characteristics relative to corresponding wild type plants or other control plants. The invention also provides constructs useful in the methods of the invention. Yet furthermore, the present invention relates generally to the field of molecular biology and concerns a method for enhancing various economically important yield-related traits in plants. More specifically, the present invention concerns a method for enhancing yield-related traits in plants by modulating expression in a plant of a nucleic acid encoding an AUX/IAA (auxin/indoleacetic acid) polypeptide. The present invention also concerns plants having modulated expression of a nucleic acid encoding IAA polypeptide, which plants have enhanced yield-related traits relative to control plants. The invention also provides constructs comprising AUX/IAA-encoding nucleic acids, useful in performing the methods of the invention.

28 Claims, 40 Drawing Sheets

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		1		50
100	(1)	-----MASTMLSLASTTPSASLSMQEILKGKARLGSGSVSTLFNK		
102	(1)	-----MASTMISLASATPSASLSVQETLKGMRLGSSSVSTLFNK		
110	(1)	-----		
76	(1)	-----MASSFLSAASHAVSPSCSLSTTHKGKPM LGG---NTLRFH		
112	(1)	-----MATAAAFSVSSPAASAVAARSKVFGG---VNQARTR		
114	(1)	-----MALAMMIRNAASKRGMT P-----		
118	(1)	-----MAIRNSLTGQFLRR-----		
170	(1)	-MAASTSSISRLGFRHHQPLGTNPGSHSQPSG--SVSFLSGSHCFYFKPL		
172	(1)	MAATSTSTSYRLGFRHLHQQLAPCSGSHPQTSG--AVSFLSGSHNFSFKSL		
174	(1)	--MTAASSSSLLGSSRIGSGPTISGLHSDSLNPTSISFSSNLQGLSLRSS		
176	(1)	--MTAASSSSLLGSSRIGSRPTISGLHSDSLNPRSITFSSTLQGLSLRSS		
44	(1)	-----		
2	(1)	-----		
4	(1)	-----MPSANVRGAQPSADRRLSTLVRHLLPSSA---RTATT		
24	(1)	--MRPPVILKTTTSLLDSSSSSPPCDRRLNLTARHFLPQMA----S----		
6	(1)	-----		
14	(1)	-----MKTNDFSSSSSSSPSDRRIGALLRHLTA-----GTDAD		
8	(1)	-----MKTTHFSSSSS---SDRRIGALLRHLNS-----GSDSD		
50	(1)	-----MNPELTSPSSS---SDRRISVLARHLVG-----VEMDP		
54	(1)	-----MHTQQSPSPS---ADRRLSVLARHLEPSSV-----AVEGH		
62	(1)	-----		
Consensus	(1)		S	R L
		51		100
100	(41)	EKGNPFIAKKSFGRI SM TVAVNVSRFEGIAMAPPDPILGVSEAFRADTDV		
102	(41)	EKGNPSIKKKSFGRI SM TVAVNVSRFEGIAMAPPDPILGVSEAFRADIDV		
110	(1)	-----MAIAVNTSRFEGVTMAPPDPILGVSEAFRADNSE		
76	(38)	KGPNSFSSSRSRGRISMAVAVNVSRFEGIPMAPPDPILGVSEAFKADNSD		
112	(34)	TGCRVGITRKNFGRVMMALAVDVSRFEGVPMAPPDPILGVSEAFKADKSE		
114	(19)	-----ISGHFGGLR-SMSSWWKSVEPAPKDPILGVTEAFLADPSP		
118	(15)	-----SSVAGARLMSSSSSWFRSIEPAPKDPILGVTEAFLADQSP		
170	(48)	EATRQSQLSRVSVVVKAESRSEEMQVDISLSPRVTAVKPSKTVAITDQAT		
172	(49)	ETTRRSQLSRISVVVKAESRSEEMQLDISLSPRVNAVKPSKTVAITDQAT		
174	(49)	GAKRQ-LYSRGTG SVVIAQNMDRVEVDLSLSPRVNSVKPSKTVAITDQAT		
176	(49)	GSKRQ-LYSRGTG SVVIAQNMDRVEVDLSLSPRVNSVKPSKTVAITDQAT		
44	(1)	-----MAPSVFEHVQQAPEDPILGVT VAYNKDPSP		
2	(1)	-----MASSSVFAGLAQAPEDPILGVT VAYNKDPSP		
4	(35)	TSTSSSAADADSSLQAFPTMASSSVFAGLAQAPEDPILGVT VAYNKDPSP		
24	(41)	-----HDS-ISASPTSASDSVFNHLVRAPEDPILGVT VAYNKDPSP		
6	(1)	-----MDSVFSNVARAPEDPILGVT VAYNNDPSP		
14	(34)	RVSS-----VFASPTSGGAG-GSVFAHLVQAPEDAILGV TIAYNKDPSP		
8	(31)	NLSS-----LYASPTSGGTG-GSVFSHLVQAPEDPILGVT VAYNKDPSP		
50	(31)	QNDS-----ISAFPTSGSDS-NSVFSHVVRGPEDPILGVT VAYNKDPSP		
54	(33)	SNHS-----IVGAPTSGNDGKQSVF SHIVRAPEDPILGVT VAYNKDTSP		
62	(1)	-----MNSQHPDGSVFSNIVRAPEDPILGVT VAYNKDTSP		
Consensus	(51)		G	S F I AP DPILGVT AY D SP

FIGURE 1

		101		150
100	(91)	KKLNLGVGAYRTEELQPYVLDDVVKKAENLMLE-RGENKEYLP	IEGLAAFN	
102	(91)	KKLNLGVGAYRTEELQPYVLDDVVKKAENLMLE-RGENKEYLAI	EGGLAAFN	
110	(35)	MKLNLGVGAYRTEELQPYVLNVVKKAEENLMLE-RGENKEYLP	IEGLAAFN	
76	(88)	VKLNLGVGAYRTEELQPYVLNVVKKAEENLMLE-RGDNKEYLP	IEGLAAFN	
112	(84)	LKLNLGVGAYRTEELQPYVLNVVKKAEENLMLE-KGENKEYLP	IEGLAAFN	
114	(58)	EKVNNGVGAYRDDNGKPVVLECVREAEKRLAG--STFMEYLP	MGGS AKMV	
118	(55)	NKVNNGVGAYRDDHGKPVVLECVREAEERRVAG--SQFMEYLP	MGGS IKMI	
170	(98)	ALAQAGVPVIRLAAGEPDFDTPAVIAEAGINAI	REGHTRYTPNAGTQELR	
172	(99)	ALVQAGVPVIRLAAGEPDFDTPVIVIAEAGINAI	REGHTRYTPNAGTQELR	
174	(98)	ALVQAGVPVIRLAAGEPDFDTPAPIAEAGINAI	REGHTRYTPNAGTMELR	
176	(98)	ALVQAGVPVIRLAAGEPDFDTPAPIAEAGINAI	REGHTRYTPNAGTMELR	
44	(31)	LKVNNGVGAYRTEEGKPLVLNVVRRAEQQLVADR	SRNKEYQPITGISQFN	
2	(32)	VKVNNGVGAYRTEEGKPLVLNVVRRAEQMLINN	PSRVKEYLPITGLADFN	
4	(85)	VKVNNGVGAYRTEEGKPLVLNVVRRAEQMLINN	PSRVKEYLPITGLADFN	
24	(81)	VKLNLGVGAYRTEEGKPLVLNVVRRVEQQLIND	VS RNKEYIPIVGLADFN	
6	(30)	VKINLGVGAYRTEEGKPLVLDVVRKAEQQLVND	PSRVKEYIPIVGISDFN	
14	(77)	IKLNLGVGAYRTEEGKPLVLNVVRKAEQQLIND	RSRIKEYLPIVGLVEFN	
8	(74)	VKLNLGVGAYRTEEGKPLVLNVVRKAEQQLIND	RTRIKEYLPIVGLVEFN	
50	(74)	VKLNLGVGAYRTEEGKPLVLNVVRKAEQQLVND	RSRVKEYLPITGLAEFN	
54	(77)	MKLNLGVGAYRTEEGKPLVLNVVRQAEQQLVND	RSRIKEYLPITGLADFN	
62	(36)	IKLNLGVGAYRTEEGKPLVLNVVRRAEQQLVND	PSRVKEYLPIVGLAEFN	
Consensus	(101)	VKLNLGVGAYRTEEGKPLVLNVVRKAEQ LI RS	KEYLPI GLAEFN	
		151		200
100	(140)	KVTAELLFGADNPVIKQQRVATVQGLSGTGS	LRLAAALIER YFP-GA QVL	
102	(140)	KVTAELLFGADNQVIEQQRVATVQGLSGTGS	LRLAAALIER YFP-GA QVL	
110	(84)	KVTAELLFGAGNPVIEQQRVATVQGLSGTGS	LRLAAALIER YFP-GAKVL	
76	(137)	KATAELLGADNPAIKQQRVATVQGLSGTGS	LRLGAALIER YFP-GAKVL	
112	(133)	KATAELLGADNPVINQGLVATLQSLSGTGS	LRLAAAFIQR YFP-EAKVL	
114	(106)	DLTLKLAYGDNSEFIKDKRIA AVQTLSGTG	ACRLFA DFQKRFSP-GS QIY	
118	(103)	EESLKLAFGDNSEFIKDKRIA AVQALSGTG	ACRLFA AFQQR FHP-NTQIY	
170	(148)	VAICQKLKEENGISYKPD-----QILVSNG	AKQSIYQAILAVCSPGDEVI	
172	(149)	VAICHKLKEENGISYTPD-----QILVSNG	AKQSIYQAMLAVCSPGDEVI	
174	(148)	SAICHKLKEENGLSYTPD-----QIVVSNG	AKQSIYQAVLAVCSPGDEV L	
176	(148)	SAICHKLKEENGLSYTPD-----QIVVSNG	AKQSIYQAVLAVCSPGDEV L	
44	(81)	KLSAKLILGANS PAIAENRVATVQALSGTG	ALRVGA EFISRHYA-KPIIF	
2	(82)	KLSAKLIFGADSPA IQENRVATVQCLSGTG	SLRVGGEFLARHYH-ERTIY	
4	(135)	KLSAKLIFGADSPA IQENRVATVQCLSGTG	SLRVGGEFLARHYH-ERTIY	
24	(131)	KLSAKLIFGADSPA IQDNRVTTVQCLSGTG	SLRVGGEFLAKHYH-QRTIY	
6	(80)	KLSAKLILGADSPA ITESRVTTVQCLSGTG	SLRVGA EFLKTHYH-QSVIY	
14	(127)	KLSAKLILGADSPA IRENRVTTVECLSGTG	SLRVGGEFLARHYH-QKTIY	
8	(124)	KLSAKLILGADSPA IRENRI TTVECLSGTG	SLRVGGEFLAKHYH-QKTIY	
50	(124)	KLSAKLMFGANCPA IQENRVTTVQCLSGTG	SLRVGA EFLAKHHH-QRTIY	
54	(127)	KLSAKLILGADSPA IQENRVTTVQCLSGTG	SLRVGGEFLAQHYH-QRTIY	
62	(86)	KLSAKLIFGADSPA IQENRVATVQGLSGTG	SLRIGA EFLARHYH-QHTIY	
Consensus	(151)	KLSAKLI GADSPA I ENRVATVQ LSGTG	SLRVGA EFL RHY IY	

FIGURE 1 (continued)

	201	250
100	(189) ISSPTWGNHKNIFNDARVPWSEYRYYPKTVGLDFEGMISDIKAAPEGSF	
102	(189) ISSPTWGNHKNIFNDARVPWSEYRYYPKTVGLDFEGMISDIKAAPEGSF	
110	(133) ISSPTWGNHKNIFNDARVPWSEYRYYPKTVGLDFDGMISDIKAAPEGSF	
76	(186) ISAPTWGNHKNIFNDASVPWSEYRYYPKTVGLDFEGMIEDIKSAPEGSF	
112	(182) ISSPTWGNHKNIFNDARVPWSEYRYYPKTVGLDFEGMIADIEAAPEGSF	
114	(155) IPVPTWSNHHNIWKDAQVPQKTYHYYPETKGLDFSALMDDVKNAPEGSF	
118	(152) IPVPTWANHHNIWRDAGVPMKTFRYYPESRGLDFSGLMDDIKNAPDGSF	
170	(193) IPAPFWVSYPEMARLADATPVILPTSISENFLLLDPKQLESKLNK---SR	
172	(194) IPAPFWVSYPEMARLADATPVILPTSISENFLLLDPKLLESKLSAK---SR	
174	(193) IPAPYWVSYPEMARMADAMPVILPTSISEDFLLLDPKLLESKLTEK---SR	
176	(193) IPAPYWVSYPEMARMADATPVILPTSISEDFLLLDPKLLESKLTEK---SR	
44	(130) LPNPTWGNHNKIFPLGGVPQKPYRYYPKTRGLDYEGLLEDLKAAPDGAV	
2	(131) IPQPTWGNHPKVFTLAGLTVRSYRYYPATRGLDFQGLLEDLGSAPSGAI	
4	(184) IPQPTWGNHPKVFTLAGLTVRSYRYYPATRGLDFQGLLEDLGSAPSGAI	
24	(180) LPTPTWGNHPKVFNLAGLSVKTYRYYPATRGLDFQGLLEDLGSAPSGSI	
6	(129) IPKPTWGNHPKVFNLAGLSVEYFRYYDPATRGLDFKGLLEDLGAAPSGAI	
14	(176) IPQPTWGNHPKIFTLAGLSVKTYRYYPSTRGLNFGQGLLEDLSAAPQSGSI	
8	(173) ITQPTWGNHPKIFTLAGLTVKTYRYYPATRGLNFGQGLLEDLGAAAPSGSI	
50	(173) IPQPTWGNHPKIFTLAGLSVKTYRYYPATRGLNFGQGLVEDLNSAPSGAI	
54	(176) IPQPTWGNHTKIFALAGLSVKSYRYYPATRGLHFQGLLEDLGSAPSGAI	
62	(135) IPVPTWGNHPKIFTIAGLSVKTYRYYPETRGLDFKGLLEDLGAAPTGAI	
Consensus	(201) IP PTWGNHP IF LAGLS K YRYYP TRGLDF GLEDL AAP GS	
	251	300
100	(239) VLLHGCAHNPTGIDPTPEQWEKIADVIQEK-NHVPFFDVAYQGFASGSLD	
102	(239) VLLHGCAHNPTGIDPTPEQWEKIADVIQEK-NHIPFFDVAYQGFASGSLD	
110	(183) VLLHGCAHNPTGIDPTPEQWEKIADVIQEK-NHIPFFDVAYQGFASGSLD	
76	(236) ILLHGCAHNPTGIDPTPEQWEKIADLIEEK-NHIPFFDVAYQGFASGSLD	
112	(232) VLLHGCAHNPTGIDPTPEQWEKIADVIQEK-KHMPFFDVAYQGFASGSLD	
114	(205) FLLHACAHNPTGVDPTTEEQWREISQLFKAK-KHFAFFDMAYQGFASGDPA	
118	(202) FLLHACAHNPTGVDPSEEQWREISSQIKAK-GHFPFFDMAYQGFASGDPE	
170	(240) LLILCSPSNPTGSVYPKLLLEEIAKIVAKHPRLLVLSDIEYEHIIYAPAT	
172	(241) LLILCSPSNPTGSVYSKLLLEEIAKIVAKHPRLLVLSDIEYEHIIYAPAT	
174	(240) LLILCSPSNPTGSVYPRKLLLEEIAEIVARHPRLLVISDEIYEHIIYAPAT	
176	(240) LLILCSPSNPTGSVYPRKLLLEEIAEIVARHPRLLVISDEIYEHIIYAPAT	
44	(180) ILLHACAHNPTGVDPTTEEQWEGIRQVIRSK-HQLPFFDCAYQGFASGSLD	
2	(181) VLLHACAHNPTGVDPTLDQWEQIR-----	
4	(234) VLLHACAHNPTGVDPTLDQWEQIRQLMRSK-ALLPFFDSAYQGFASGSLD	
24	(230) VLLHACAHNPTGVDPTLEQWEQIRQLIRSK-ALLPFFDSAYQGFASGSLD	
6	(179) VLLHACAHNPTGVDPTSEQWEQIRQLMRSK-SLLPFFDSAYQGFASGSLD	
14	(226) VLLHACAHNPTGVDPTLEQWEQIRKLMRSK-GLMPFFDSAYQGFASGSLD	
8	(223) VLLHACAHNPTGVDPTIQQWEQIRKLMRSK-GLMPFFDSAYQGFASGSLD	
50	(223) VLLHACAHNPTGVDPTSQQWEQIRKLMRSK-GLMPFFDSAYQGFASGSLD	
54	(226) VLLHACAHNPTGVDPTKDQWEQIRRLMRSK-GLLPFFDSAYQGFASGSLD	
62	(185) VLLHACAHNPTGVDPTLEQWEQIRQLMRSK-GLLPFFDSAYQGFASGSLD	
Consensus	(251) VLLHACAHNPTGVDPT EQWE IA LIRSK LLPFFD AYQGFASGSLD	

FIGURE 1 (continued)

301 350

100 (288) ADASSVRLFAARGMELLIAQS-----YSKNLGLYAERIGAINVVCSS
102 (288) ADASSVRLFAARGMELLVAQS-----YSKNLGLYAERIGAINVVCSS
110 (232) ADASSVRLFAARGMELLVAQS-----YSKNLGLYAERIGAINVVCSS
76 (285) EDAASVRLFVARGIEVLVAQS-----YSKNLGLYAERIGAINVISSS
112 (281) EDAFSVRLFVKRGMEVFVAQS-----YSKNLGLYSERVGAINVVCSSA
114 (254) RDAKSIRIFLEDGHHIGISQS-----YAKNMGLYGQRVGCLSVLCED
118 (251) RDAKAIKIFLEDGHLIGLAQS-----YAKNMGLYGQRAAGSLSVLCED
170 (290) HTSFASLPGMWERTLTVNGFS-----KAFAMTGWRLGYLAGPKHF
172 (291) HTSFASLPGMWERTLTVNGFSKINIWKAGNLQAFAMTGWRLGYLAGPKHF
174 (290) HTSFASLPGMWDRTLTVNGFS-----KAFAMTGWRLGYLAGPKHF
176 (290) HTSFASLPGMWDRTLTVNGFS-----KAFAMTGWRLGYLAGPKHF
44 (229) KDAHAVRLFVADGGECFVAQS-----YAKNMGLYGERVGALSIVCTN
2 (205) -----
4 (283) QDAQSVRMFVADGGELLMAQS-----YAKNMGLYGERVGALSIVCGS
24 (279) ADAQPVRLFVADGGELLVAQS-----YAKNLGLYGERVGALSIVCKS
6 (228) TDAQSVRTFVADGGECCLIAQS-----YAKNMGLYGERVGALSIVCKS
14 (275) TDAKPIRMFVADGGELLVAQS-----YAKNMGLYGERVGALSIVCKA
8 (272) TDAKPIRMFVADGGECCLVAQS-----YAKNMGLYGERVGALSIVCKS
50 (272) ADAQPVRMFVADGGELLVAQS-----YAKNMGLYGERIGALSIVCKT
54 (275) TDAQSVRMFVADGGELVLAQS-----YAKNMGLYGERVGALSIVCRN
62 (234) ADAQSVRMFVADGGECCLAAQS-----YAKNMGLYGERVGALSIVCKA
Consensus (301) DA SVRLFVADG ELLVAQS YAKNMGLYGERVGALSIVC S

351 400

100 (330) ADAAARVKSQKRIARPMYSNPPVHGARIVANVVGDPILFNEWKEEMEL
102 (330) ADAAARVKSQKRIARPMYSNPPVHGARIVANVVGDPALFNEWKAEMEM
110 (274) ADAAARVKSQKRIARPMYSNPPVHGARIVANVVGDPALFNEWKEEMEL
76 (327) PESAARVKSQKRIARPMYSNPPVHGARIVADVGNPVLNEWKAEMEM
112 (323) PEVADRVKSQKRLARPMYSNPPVHGAKIVANVVGDPMTFGEWKQEMEL
114 (296) PKQAVAVKSQKQLARPMYSNPPVHGALIVSTILEDPELKSLLWKEVKVM
118 (293) EKQAVAVKSQKQLARPMYSNPPVHGALIVSTVLGDPDLKLLWKEVKVM
170 (330) VAACNKIQSQFTSGASSISQKAGVAALGLGYAGGEAVSTMVTAFRERRDF
172 (341) VAACNKIQSQFTSGASSISQKAGVAALGLGYAGGEAVSTMVKAFMERRDF
174 (330) VSACNKLQSQFTSGASSISQKAAVAALGLGYAGGEAVATMVKAFRERRDF
176 (330) ISACNKLQSQFTSGASSISQKAAVAALGLGYAGGEAVATMLKAFHERRDF
44 (271) AAVASRVDSQKLVIRPMYSSPPAHGAAIAATILADGRLFQEWTVELKGM
2 (205) -----
4 (325) ADVAVRVESQKLVIRPMYSNPPVHGASIVATILKDSAMFNEWTVELKGM
24 (321) ADVASRVESQKLVIRPMYSSPPVHGASIVAAILKDRNLFNDWTIELKAM
6 (270) ADVASKVESQVKKLVIRPMYSSPPVHGASIVATILKSSDMYNNWTIELKEM
14 (317) ADVAGRVSQKLVIRPMYSNPPVHGASIVAVILDRNLFNEWTVELKAM
8 (314) ADVAGRVSQKLVIRPMYSSPPVHGASIVAVILDRNLFNEWTVELKAM
50 (314) ADVAGRVSQKLVIRPMYSNPPVHGASIVAAILKDRDLYNEWTVELKAM
54 (317) ADVTSRVESQKLVIRPMYSNPPVHGASIVATILKDRNLYHEWTVELKAM
62 (276) ADVASRVESQKLVIRPMYSNPPVHGASIVATILKDSAMYNEWTVELKAM
Consensus (351) AD A RV SQLK VARPMYSNPPVHGA IVA IL D LFNEW ELK M

FIGURE 1 (continued)

		401		450
100	(380)	AGRIKNVRQKLFDSL	--AKDKSGKDWSFILKQIGMFSFTGLNKTQSEN	M
102	(380)	AGRIKNVRQKLFDSL	--AKDKSGKDWSFILKQIGMFSFTGLNKAQSDN	M
110	(324)	AGRIKNVRQKLFDSL	--AKDKCGKDWSFILKQIGMFSFTGLSKVQSEN	M
76	(377)	AGRIKNVRQQLYDSIT	--SKDKSGKDWSFILKQIGMFSFTGLNKNQSDN	M
112	(373)	AGRIKNVRQKLYDSL	--AKDKSGKDWSFILRQIGMFSYTGLNKAQSDN	M
114	(346)	ADRIIGMRTTTLRESLE	--KLGSP--SWEHVTQIGMFCYSGLTPEQVDRL	
118	(343)	ADRIIGMRTTTLRENLE	--KKGSTL--PWQHITNQIGMFCYSGLTPEQVDRM	
170	(380)	LIKSFGEEMEGVGLSEPLGAFYLFIDFSSYYGAEVEGFGKIDDSALCRYL		
172	(391)	LIRSFGEIDGVKISEPRGAFYLFIDTSSYYGTEAEGFGKIEDSDSLCRYL		
174	(380)	LVKSFGGEIDGVKISEPRGAFYLFIDLSSYYGVEVDGFGTINNSESLCRYL		
176	(380)	LVKSFGGEIDGVKISVPRGAFYLFIDLSSYYGVEVDGFGTVNNSESLCRYL		
44	(321)	ADRIISMRRQQLYDALQ	--ARGTPG--DWTHVLKQIGMFTFTGLNKSQVEFM	
2	(205)	-----		
4	(375)	ADRIISMRRQQLFDALK	--TRETPG--DWSHIKQIGMFTFTGLNSDQVAFM	
24	(371)	ADRIISMRRQQLFDALC	--SRGTPG--DWSHIKQIGMFTFTGLNAEQVSFM	
6	(320)	ADRIKSMRRQQLFEAIQ	--ARGTPG--DWSHIKQIGMFTFTGLNKEQVEFM	
14	(367)	ADRIISMRRQQLFEALR	--ARGTPG--DWTHIHKQIGMFTFTGLNPAQVSYM	
8	(364)	ADRIISMRRQQLFEALR	--TRGTPG--DWSHIKQIGMFTFTGLNPAQVSFM	
50	(364)	ADRIISMRRQQLFEALH	--ARGTPG--DWSHIVKQIGMFTFTGLNKSQVAFM	
54	(367)	ADRIISMRRQQLFDALR	--AKGTPG--DWSHIKQIGMFTFTGLNKEQVAFM	
62	(326)	ADRIISMRRQQLFDTLR	--DRGTPG--DWSHIKQIGMFTFTGLNTEQVAFM	
Consensus	(401)	ADRI MRQ LFDL	AR T G DWSHIKQIGMFTFTGLN QV FM	
		451		500
100	(428)	TNKWHVYMTDGRISLAGLSLAKCEYLADAIIDSYHNVS	-----	
102	(428)	TNKWHVYMTKDGRISLAGLSLAKCEYLADAIIDSYHNVS	-----	
110	(372)	TNKWHVYMTKDGRISLAGLSLAKCEYLADAIIDSFHCVS	-----	
76	(425)	TNKWHVYMTKDGRISLAGLSLAKCEYLADAIIDSYHNVS	-----	
112	(421)	TDKWHIYMTKDGRISLAGLSLAKCDYLADAIIDSFHNVN	-----	
114	(393)	TSEYHIYMTNRNGRISMAGVTTGNVGYLANAIHEVTKSS	-----	
118	(390)	TNEFHIYMTNRNGRISMAGLNTGNVGYVADAIHEVTKSF	-----	
170	(430)	LDQAQVALVPGVAFGDDSCIRISYAASLTTLQAAVERIKKALLPLKSAVP		
172	(441)	LDQAQVALVPGVAFGDDSCIRISYAASLTTLQEAVERIKKALLPLKSAVP		
174	(430)	LDKSQVALVPGDAFGDDTCIRISYAASLTTLQAAVERIKKALVTLRPPVP		
176	(430)	LDKSQVALVPGDAFGDDTCIRISYAASLTTLQAAVERIKKALVTLKPPVP		
44	(368)	TRQYHIYMTSDGRISMAGLSSKTVPHLADAIHAAVVGARQS	-----	
2	(205)	-----		
4	(422)	RQEYHIYMTSDGRISMAGLSGRTIPHLADAIHAAVTKLK	-----	
24	(418)	TKEFHIYMTSDGRISMAGLSSKTVPLLADAIHAAVTRVV	-----	
6	(367)	TKEFHIYMTSDGRISMAGLSSKTVPHLADAMHAAVTRLG	-----	
14	(414)	TKEYHIYMTSDGRISMAGLSSKTVPHLADAIHAVVTKAL	-----	
8	(411)	TKEYHIYMTSDGRISMAGLSSKTVPHLADAIHAVVTKAV	-----	
50	(411)	TKEYHIYMTSDGRISMAGLSSKTVPHLADAMHAAVKRVV	-----	
54	(414)	TKEYHIYMTSDGRISMAGLSSRTVPHLTDAIHAAVTRAR	-----	
62	(373)	TKEYHIYMTSDGRISMAGLSSRTVPHLADAIHAAVTRIP	-----	
Consensus	(451)	T YHIYMT DGRISMAGLS TV HLADAIHAAV RV		

FIGURE 1 (continued)

		501
100	(467)	-
102	(467)	-
110	(411)	-
76	(464)	-
112	(460)	-
114	(431)	-
118	(428)	-
170	(480)	V
172	(491)	V
174	(480)	V
176	(480)	V
44	(410)	-
2	(205)	-
4	(461)	-
24	(457)	-
6	(406)	-
14	(453)	-
8	(450)	-
50	(450)	-
54	(453)	-
62	(412)	-
Consensus	(501)	

FIGURE 1 (continued)

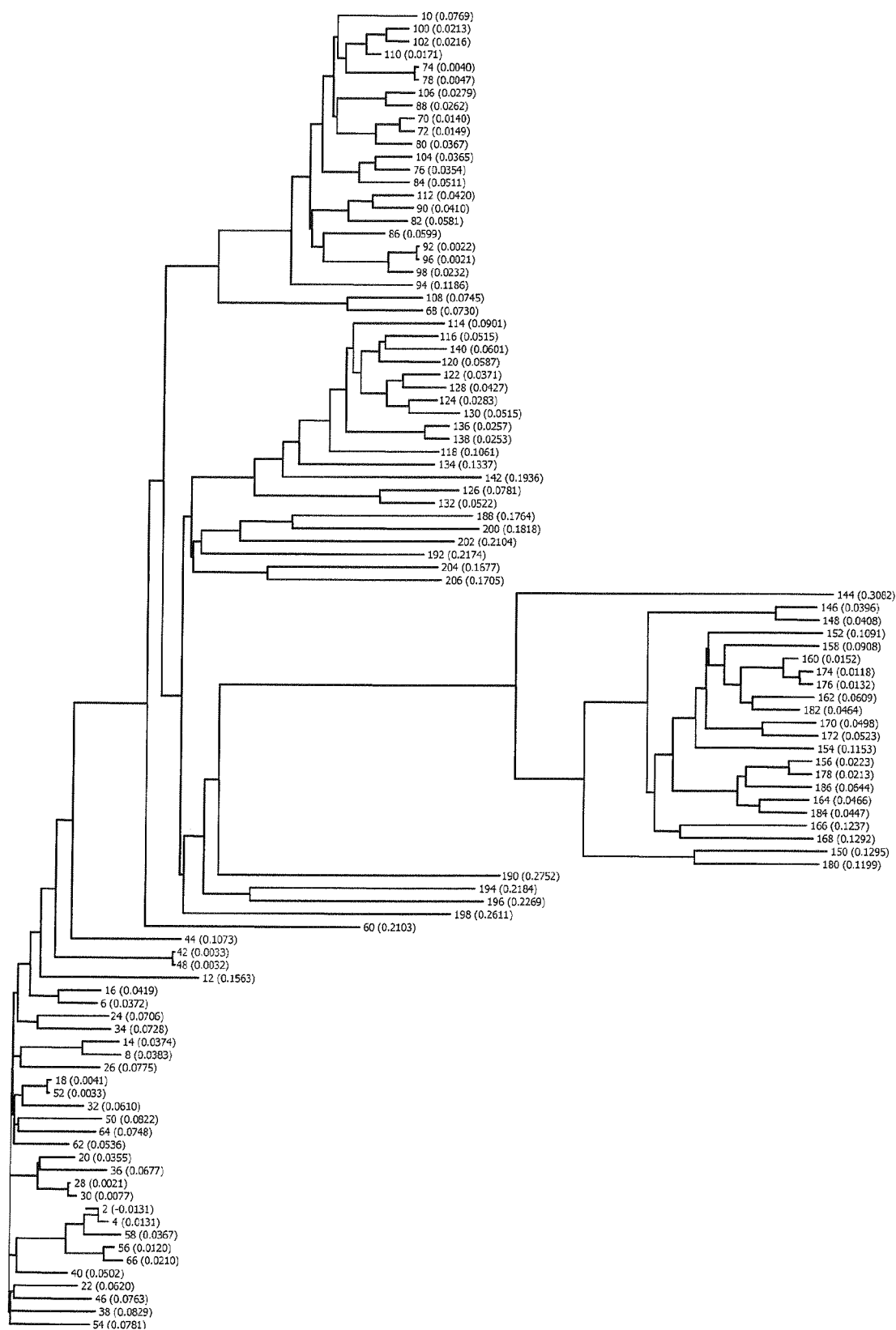


FIGURE 2

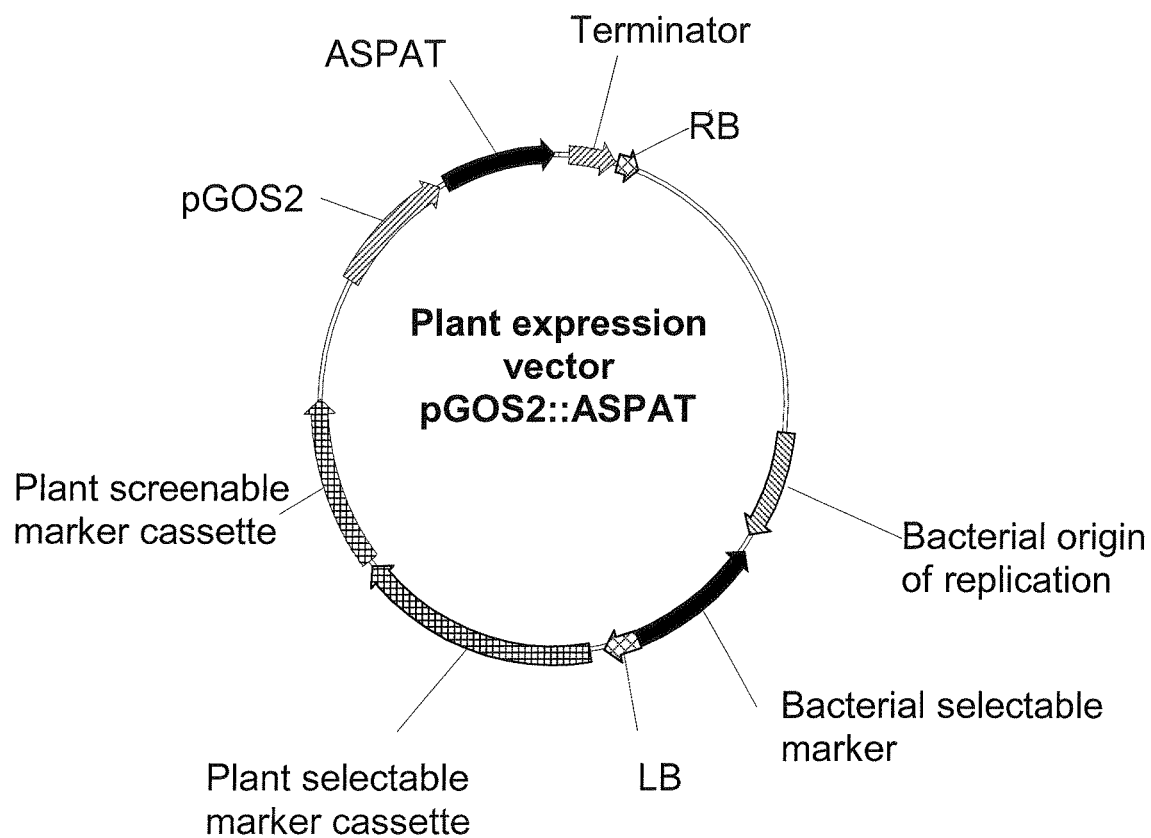


FIGURE 3

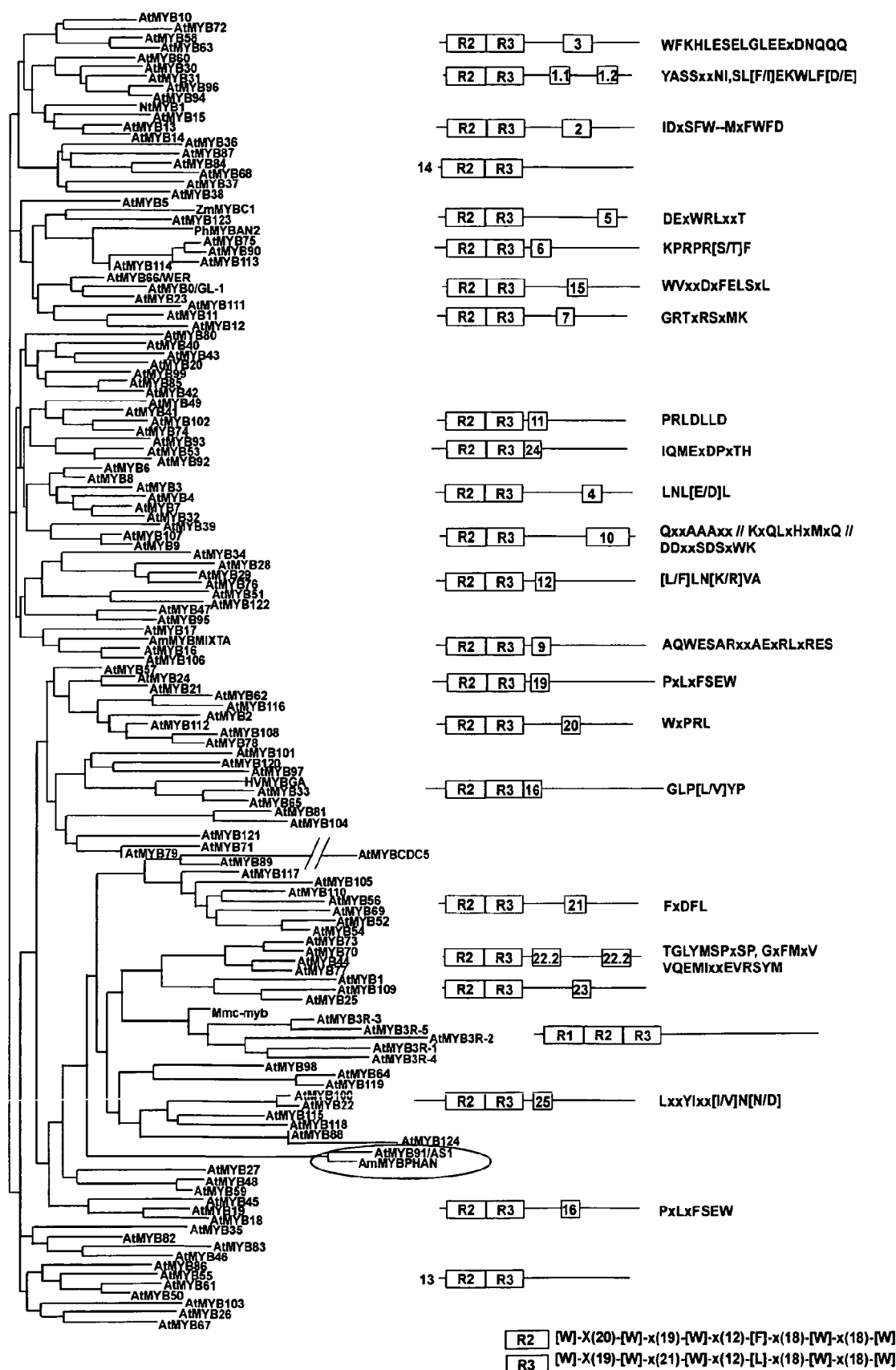


FIGURE 4

CLUSTAL W (1.81) multiple sequence alignment

```
Poptr_MYB91      -----MKERQWRRAEEDALLRAYVKQYGPREWNLVSQRMNTPLNRDAKSLERWKNYLKP
Medtr_MYB91__PHAN_ --MSDMKDRQRWRAEEDALLRAYVKQYGPREWNLVSQRMNTPLNRDAKSLERWKNYLKP
Pissa_MYB91      -MSLEMKDRQRWRAEEDALLRAYVKQYGPREWNLVSQRMNTPLNRDAKSLERWKNYLKP
Glyma_MYB91__PHANa_ -----MKDRQRWRAEEDALLRAYVKQYGPREWNLVSQRMNTPLNRDAKSLERWKNYLKP
Glyma_MYB91__PHANb_ -----MKERQWRRAEEDALLRSYVKQYGPREWNLVSQRMNTYPLNRDAKSLERWKNYLKP
Lotco_MYB91__PHANb_ -----MKERQRWSSEEDALLHAYVQYGPREWNLVSQRMNTPLNRDTKSLERWKNYLKP
Lotco_MYB91__PHANa_ -----MKERQRWTSEEDALLCAYVKQYGPREWHLVSQRMNTTLHRDAKSLERWKNYLKP
Eucgr_MYB91      -----MKERQWRRAEEDALLRAYVKQYGPREWHLVSQRMNTPLNRDAKSLERWKNYLKP
Maldo_MYB91      -----MKERQRWSAEEDALLRAYVKQYGPREWNLVSQRMNTPLDRDAKSLERWKNYLKP
Lyces_MYB91      -----MRERQWRRAEEDALLRAYVRQYGPKEWPLVSQRMNTPLNRDAKSLERWKNYLKP
Soltu_MYB91      -----MRERQWRRAEEDALLRAYVRQYGPKEWHLVSQRMNTPLNRDAKSLERWKNYLKP
Nicta_MYB91      -----MRERQWRSEEDALLRAYVKQYGPKEWHLVSQRMNTALNRDAKSLERWKNYLKP
Vitvi_MYB91      -----MKERQWRRAEEDALLRAYVRQYGPREWNLVSQRMNTPLDRDAKSLERWKNYLRP
Goshi_MYB91      -----MKERQWRRAEEDALLCAYVKQYGPREWNLVSHRMNTPLNRDAKSLERWNNYLKP
Aqufo_MYB91      -----MKERQWRRAEEDALLRAYVKQYGPREWNLVSQRMNTPLDRDAKSLERWKNYLKP
Escca_MYB91      -----MKERQWRRAEEDALLRAYVKQYGPREWNLVSQRMNTHLDRDAKSLERWKNYLKP
Arath_AS1_MYB91  -----MKERQRWSGEEDALLRAYVRQFGPREWHLVSERMNKPLNRDAKSLERWKNYLKP
Carhi_MYB91      -----MKERQRWSGEEDALLRAYVRQFGPREWHLVSERMNKPLNRDAKSLERWKNYLKP
Brana_MYB91      -----MKERQRWSGEEDALLRAYVRQFGPREWHLVSERMNKPLNRDAKSLERWKNYLKP
Antma_MYB91      -----MKERQWRPEEDALLRAYVKEYGPRDWHLVTQRMNKPLNRDAKSLERWKNYLKP
Orysa_MYB91      MQPPPMRERQRWRPEEDAILLAYVRQYGPRESLVSQRMNRPPLHRDAKSLERWKNYLRP
Zeama_MYB91__RS2_ -----MKERQWRPEEDAILLAYVRQYGPREWHLVSQRMNVALDRDAKSLERWKNYLRP
Moral_MYB91      -----MKERQRWQPEEDALLRAYVKQYGPREDWNLVYQRMGKPLHRDPKSLERWKNYLKP
*.:*** ***: * :*: :*: :* ** *. * .:*****:***:
XXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX
XXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX
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Poptr_MYB91      GIKKGSLTEEEQSLVIRLQAKHGNKWKKIAAEVPGRTAKRLGKWWVEVFKEKQQR-ELKEN
Medtr_MYB91__PHAN_ GIKKGSLTEEEQRLVISLQATHGNKWKKIAAQVPGRTAKRLGKWWVEVFKEKQQRRTKSGSI
Pissa_MYB91      GIKKGSLTEEEQHLVISLQATHGNKWKKIAAQVPGRTAKRLGKWWVEVFKEKQQRRTKG-I
Glyma_MYB91__PHANa_ GIKKGSLTEEEQRLVINLQATHGNKWKKIAAQVPGRTAKRLGKWWVEVFKEKQQRRTKG-N
Glyma_MYB91__PHANb_ GIKKGSLTEEEQRLVIHLQAKYGNKWKKIAAEVPGRTAKRLGKWWVEVFKEKQQRREKKE-I
Lotco_MYB91__PHANb_ GIKKGSLTKEEQRLVILLQANYGNKWKKIAAEVPGRTAKRLGKWWVEVFKEKQQRREKIE-I
Lotco_MYB91__PHANa_ GIKKGSLTEEEQRLVIRLQAKHGNKWKKIAAEVPGRTAKRLGKWWVEVFKEKQQRREKQE-I
Eucgr_MYB91      GIKKGSLSSEEQRLVIQLQAKHGNKWKKIAAEIPGRTAKRLGKWWVEVFKEKQQR-EQKEN
Maldo_MYB91      GIKKGSLTEEEQRLVICLQAKHGNKWKKIAAEVPGRTAKRLGKWWVEVFKEKQQR-EQKNK
Lyces_MYB91      GIKKGSLTEDEQRLVIQLQAKHGNKWKKIAAEVPGRTAKRLGKWWVEVFKEKQQR-EQKEN
Soltu_MYB91      GIKKGSLTEDEQRLVIQLQAKHGNKWKKIAAEVPGRTAKRLGKWWVEVFKEKQQR-EQKEN
Nicta_MYB91      GIKKGSLTQEEQRLVIHLQAKHGNKWKKIAAEVPGRTAKRLGKWWVEVFKEKQHR-EQKEN
Vitvi_MYB91      GIKKGSLTEEEQRLVIRLQAKHGNKWKKIAAEVPGRTAKRLGKWWVEVFKEKQQR-EQKEN
Goshi_MYB91      GIKKGSLTEEEQRLVIRLQAKHGNKWKKIAAEVPGRTAKRLGKWWVEVFKEKQQR-EHKEK
Aqufo_MYB91      GIKKGSLTEEEQRLVIRLQAKHGNKWKKIAAEVPGRTAKRLGKWWVEVFKEKQQR-EQKEN
Escca_MYB91      GIKKGSLTEEEQRLVIRLQAKHGNKWKKIAAEVPGRTAKRLGKWWVEVFKEKQQR-EQKET
Arath_AS1_MYB91  GIKKGSLTEEEQRLVIRLQEKHGNKWKKIAAEVPGRTAKRLGKWWVEVFKEKQQR-EEKES
Carhi_MYB91      GIKKGSLTEEEQRLVIRLQEKHGNKWKKIAAEVPGRTAKRLGKWWVEVFKEKQQR-EEKES
Brana_MYB91      GIKKGSLTEEEQRLVIRLQEKHGNKWKKIAAEVPGRTARRLGKWWVEVFKEKQQR-EEKES
Antma_MYB91      GIKKESLTQEEQILVINLQAKHGNKWKKIAAEVPGRTAKRLGKWWVEVFKEKQQR-EKDN
Orysa_MYB91      GIKKGSLTDDQRLVIRLQAKHGNKWKKIAAEVPGRTAKRLGKWWVEVFKEKQQRRLDRD
Zeama_MYB91__RS2_ GIKKGSLTEEEQRLVIRLQAKHGNKWKKIAAEVPGRTAKRLGKWWVEVFKEKQQRRLRDS-
Moral_MYB91      GLKKGSLTPEEQSLVISLQAKYGNKWKKIAAEVPGRTAKRLGKWWVEVFKEKQLKQLQLQK
*:** *: ** ** *: :*****:*****:*****:***: :
XXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX
XXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX
```

FIGURE 5

NKTVEPIDE-----GKYDRILETFAEKLVKERP---SPAFVMATS
 NRTVDPIND-----SKYEHILESFAEKLVKERP---SPSFVMAAS
 NKTVDPIN-----SKYEHILESFAEKLVKERP---SPSFVMAAS
 SCTIDPISD-----SKYEHILESFAEKLVKERPSTSTSTSFVMATS
 NRIADPINN-----SKYEHILESFAEKLVKERP---SPSFVMAAS
 NGIVSPISD-----TKYEHMLEGFAEKLVKHEHT---LPSFAMAAS
 SKSIGPVDD-----SKYDHILETFAEKLVKHEP---SPSYLMAAS
 -KGALPIDE-----GYKDHILENFAEKLVKERS---TPALLMATA
 -KITDPIVE-----GYDTHILETFAEKLVKER-----APTYLMATS
 NKVVDPVDE-----GKYDHILETFAEKIVKERS---VPGLLMATS
 NKVVDPVDE-----GKYDHILETFAEKIVKERS---VPGLLMATS
 NKVVDPVDE-----GKYDHILETFAEKIVKERS---VPGLLMATS
 NKVVDPIBE-----GKYDRILETFAEKLVKERP---APAFLMATS
 HKTVEPVEE-----GKYDRILETFAEKLVKQGH---SSAFPMAS
 NKAPEPIE-----GKYDSILETFAEKLVKCEP---NPPFLMATS
 SKTIDPIE-----GKYDQILETFAEKLVKERP---NPPLYMGTS
 NKRVEPIDE-----SKYDRILESFAEKLVKERSN-VVPAAAAAATV
 NKRVEPIDE-----SKYDRILESFAEKLVKERSNNIVVVPSPAGKV
 NKRVEPIDE-----SKYDRILESFAEKLVKERSSS---VPSAVMAS
 KKITEPIE-----GKYDRILETFAEKIVKERVVS--RIITPMPTS
 RRRLLPPLDGDGERGCAG-----GRYDWLLEDFADKLVNDHHR-----RMMA
 -RRPPEPSPDER-----GRYEWLLENFAEKLVGPERQQAAPASPILLMA
 KPSPQPDGNIPVAVAVAGGSSPADKAVQGPYDHILETFAEKIVYHQRP---NLNPAIPLP

XXXXXXXXXX

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NGTFLHPHPHPPPHPSTPAPTMLPPWLSNSNS-----TSTVRPPSPSVTLTSL
NSSYLHTDAQAP-----TPGLLP SWLSNSNN-----AAPVRPNSPSVTLTSL
NSSYLHTDAQAA-----TPGLLP SWLSNSNN-----TAPVRPNSPSVTLTSL
NSSFLHADAPAP-----APALLPSWLSNSNG-----TAPVRPPSPSVTLTSL
DGAFLLLTDTAP-----ASSLRPSWLSNSSS-----AAAIGPSSL SVKLSL
SNEAFLHTN-----SSAMLP SWLSNYDS-----TSTP-PSISVTLTSL
NGPFLHLTDTPAATP-----ASALLPPWLSNSNNP-----ATAGQPPSPSVTLTSL
NGGFIHTDSPALAP-----TLLPPWLSNSNG-----TPTLRPSPSVTLTSL
NGAYLHTETSSPAP-----TILPPWLSNSNV-----SPNVRPPSPSVTLTSL
NGGFLHADASTPTPQ-----TLLPPWLSNSSA-----PSTVRSSSPSVTLTSL
NGGFLHSDASTPTPQ-----NLLPPWLSNSTA-----PSTVRSSSPSVTLTSL
NGGFLHADAPASPQ-----TLLPPWLSNSTA-----TSTVRSPSPSVTLTSL
NGNFLHPDPPAP-----PPPTLLPPWLSNSNC-----TSTVRPPSPSVTLTSLC
NGGFLHTDPPSP-----APPTLLPPWLSNSSN-----ASVVTPPSPSVTLTSL
NGGFLHS-DPPAPP-----TMLPPWMASSNG-----TTVRPSSPSVTLTSL
NGGYLQNSAATVPPP-----TLLPPWLSSSSA-----PPTTSPSPSVTLTSL
VMANSNGGFLHSEQQVQP--PNPVI PPWLATSNNGN-----NVVARPPSVTLTSLSP
VMANSNGGFLQHSEQTQPQPPNVI PPWLATSNNGN-----NVVVRPPSVTLTSLSP
SNGGFQQAPPNNNNNNNNNNNNHVI PPWLATSNNGS-----NVVARPPSVTLTSLSP
NSGFLQNDPSPHSAQS-----VLPPWLASSMT-----TTIRQSPSVTLTSL
AP-----ILPPWMSS-----SPSSSSSPSVTLTSLA
AP-----VLPPWLSSNAGFAAAAAA VAHPPPRPSPSVTLTSLA
VPFPMNPDPVLSLGSVNSTPPALPPWMNLNVNVN-----ATTSSLSSCTTSSAT

```

FIGURE 5 (continued)

Poptr_MYB91 PSTVAAS-----PPIPWLQPERGPENTPLVLGNLPPHGIVPVCGESFLMSELVDC
Medtr_MYB91__PHAN_ PS-----TVAAPPPWMQPVRGPDN--APLVLGNVAPHGAVLSYGESMVMSELVDC
Pissa_MYB91 PS-----TVAAPPPWMQPVRGPDN--APLVLGNVAPHGAVLSYGENMVMSELIDC
Glyma_MYB91__PHANa_ PS-----TVAAPPPWMQPVRGPDNASPLVLGNVAPHGAVLAFGENMVMSELVEEC
Glyma_MYB91__PHANb_ SS-----TVATPPFSLWPPERGPDNAPFVLGNVSALHGAIPTLSDSMHMSQMVH
Lotco_MYB91__PHANb_ PS-----TVATPRG-----LENNAPFVLRNVTAHNGSVPSFSDHILMSELVGF
Lotco_MYB91__PHANa_ PS-----TVAGPPPWRG-----LENNALAMAN--TAPHGTVPAPFSDNMLVSELVDC
Eucgr_MYB91 P-----ATVPAS-----QPIPWLQADRGLDSGLSLTGLPNHGFLPTSGENILMSELAEAC
Maldo_MYB91 P-----TVAPS-----PPIPWLQQDRGSD-GSFVVGNLPHHGVPACGENLVISELVEEC
Lyces_MYB91 P-----STVP---PTPTPGIPWLQTDGRPDNAPLILSSFPHHSVAP-CGENPFITELAEAC
Soltu_MYB91 P-----STVP---PTPTPGIPWLQTDGRPDNASLILSSFPHGGVAPPCGENPFITELAEAC
Nicta_MYB91 P-----STVPP-TPTPTPGIPWLQTDGRPENAPLILSSFPHGGVAPPCGENPFVTELVEEC
Vitvi_MYB91 PSTVATS-----PTIPWLQPERGPDATPLVLGNLPPHGAVPTSGENLLISELVEEC
Goshi_MYB91 PSTVAAA-----PPIPWLQPER-MSETSPVLGNRVPHGSFPRS-ENLLISELMDC
Aqufo_MYB91 PS-----TVTPPPSIPWLQSDRGAENPSLGLG--SLSSHGSGSTGGDNHVMADLVEEC
Escca_MYB91 PS-----TIAPCTSMSWLQPDGRGNDNSNPSLVLGNFPPTHVPVPPSGGDRMLVPDLVEEC
Arath_AS1_MYB91 TVAAAAQPP-----IPWLQQQQPERAENGGGLVLGSMMPSCSGSS--ESVFLSELVEEC
Carhi_MYB91 TLAASTPPPPQ---IPWLQQQQ--QPERGENGLVLGSMMPSCSGSSSESVFLSELVEEC
Brana_MYB91 VAATPPQQQP---IPWLQQQQ---PEASPGGLVLGSMIPSCSGSN--ESVFMSELVEEC
Antma_MYB91 PS-----VVPPAPAIPLWHPDNTTHGPNLSLGLVVAPFMGENHIVPELLEEC
Orysa_MYB91 SAAVAPAPAAP---PPTWGG-----GGGGEVVVAELMEC
Zeama_MYB91__RS2_ SAAVAPGPPAP---APWMPDRAADAAPYGFPSQHGGAAPPGMVVDGQALAEAEAC
Moral_MYB91 PS-----PSVSLSLSPSEPVQQQTLEQEMNRFLPVQQMASIFQC
... :
Conserved Domain (CD) XXXXXX

Poptr_MYB91 CRELEEGHRAWAAHKKEAAWRLRRVELQLESESCRRREKMEIEIESKISLREEEKASLD
Medtr_MYB91__PHAN_ CKELEEVHHALAAHKKEAAWRLSRVELQLESEKASRRREKMEIEIAKIKALREEQAVLD
Pissa_MYB91 CKELEEGHHALAAHKKEAAWRLSRVELQLESEKASRRREKMEIEIAKIKALREEQAVLD
Glyma_MYB91__PHANa_ CKELDEVHHALAGHKKEAAWRLSRVELQLESEKAGRRREKMEIEIAKIKALREEQTAALD
Glyma_MYB91__PHANb_ CKELEEGHRAALATHKKEAAWRLSRVELQLESEKANRRREKIEEFKAKIKALQEEBKAALG
Lotco_MYB91__PHANb_ SKELEEGHRAALAAHKKEAWRLRLELQLESEKACRRRETVEEFKANIKALQEEQTAALN
Lotco_MYB91__PHANa_ CKELEEVHGALAAHKKEATWRLRRVELQLESEKANRRREKIEETEAKIKALREQQNAALE
Eucgr_MYB91 CKELEEGHRAWAAHKKEAAWRLKRLELQLESEKACRRREKMEIEIAKINTLRBEQKASLD
Maldo_MYB91 SRELEEMHRAWAAHKKEASWRLRRVELQLDSEKACRRREKMEIEIAKVKALREEQKAALD
Lyces_MYB91 CKDLDEGHRTWTAHKKEAAWRLRRVELQLESEKASKVREKMEIEIAKMKALREEQKATLD
Soltu_MYB91 CKDLDEGHRTWTAHKKEATWRLRRVELQLESEKASKVREKMEIEIAKMKALREEQKATLD
Nicta_MYB91 CKELDEGHRAWAAHKKEAAWRLRRVELQLESEKICKVREKMEIEIAKMKALREEQKATLD
Vitvi_MYB91 CRELEEGHRAWAAHKKEAAWRLRRVELQLESEKACRRREKMEIEIESVKALREEQKATLD
Goshi_MYB91 CRQLEDGRRRAWAHRKEAAWRLRRVELQLESEKASRKRKKMEIEIESKIALREEQKSTLD
Aqufo_MYB91 CRELEEGHRAWAHHKKEAAWRLKRVELQLESEKACRRRDKMEIEIESKIRALRDEQKVTLE
Escca_MYB91 CRELEESHRAALVAHKKEAAWRLKRVELQLESEKACRRREKMEIEIEMKVRALREEQKVTLD
Arath_AS1_MYB91 CRELEEGHRAWADHKKEAAWRLRLELQLESEKTCRQREKMEIEIAKMKALREEQKNAME
Carhi_MYB91 CRELEEGHRVWSEHKKEAAWRLRLELQLESEKTCRQREKMEIEIAKMKALREEQKIAME
Brana_MYB91 CRELEEGHRAWAEHKKEAAWRLRLELQLESEKTSRQREKTEEIEIAKMKALREEQKMAME
Antma_MYB91 CRELEEGQRAWAAHRKEAAWRLKRVELQLESEKACRRREKMEIEIAKMKALREEQKASLD
Orysa_MYB91 CREMEEGQRAWAAHRKEAAWRMKRVEMQLETERACRRREATEBEFEAKMRALREEQAAAVE
Zeama_MYB91__RS2_ CRELEEGCRAWAAHRRREAARWLKRVEQQLEMEREMRRREVWBEFEAKMRTMRLEQAAAAE
Moral_MYB91 CKELEEGRQSWLQHKKEATWRPSRLEQQLESEKSKRKEKMEIEDAKIRSLREEEMAFLS
..... :
Conserved Domain (CD) XXX

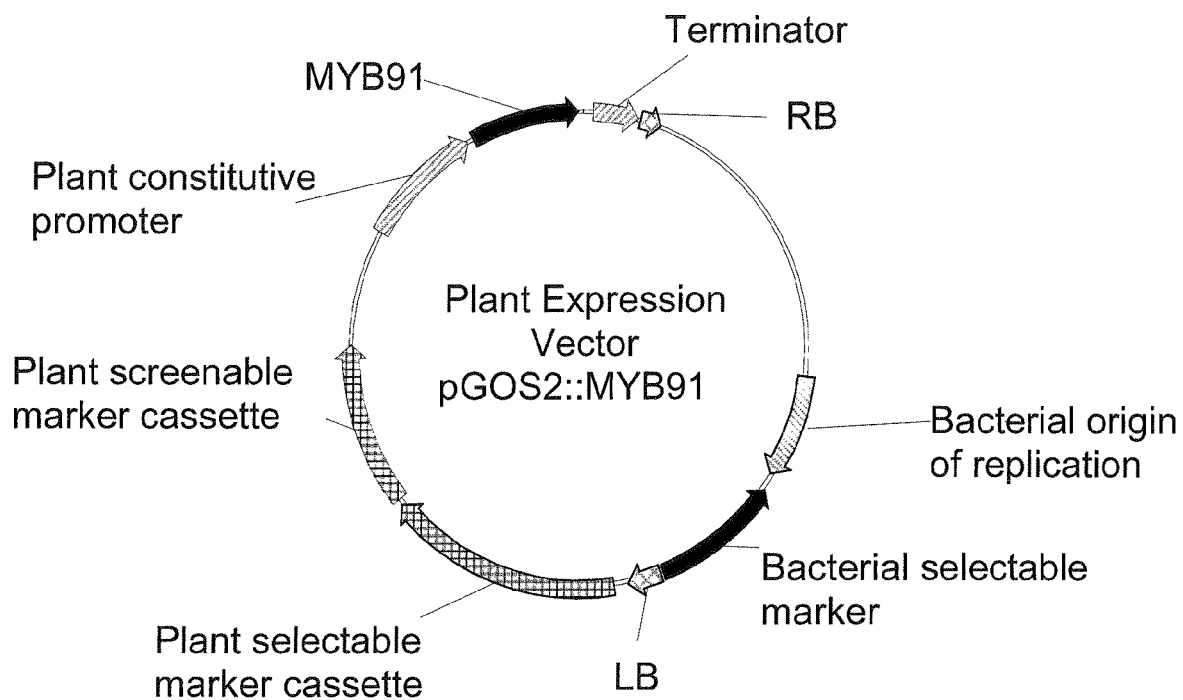
FIGURE 5 (continued)

```
Poptr_MYB91      RIEAEYREQLTGLRRDAETKEQKLSQWTAKHLRLTKFLEQMSCRPRLSEPNR-----
Medtr_MYB91__PHAN_ RIEGEYREQLAGLRRDAETKEQKLTEQWAAKHLRLTKFLEQVGCRSRHAESNGR-----
Pissa_MYB91      RIEGEYREQLAGLRRDAEAEQKLAEQWAAKHLRLTKFLEQVGCRSRHAEQNGR-----
Glyma_MYB91__PHANa_ RIEAEYREQLAGLRRDAESKEQKLAEQWAAKHLRLTKFLEQVGCRSRLTEPNGR-----
Glyma_MYB91__PHANb_ RIEAEYREQLAALRRDAENKEQKLAEQWDAAKHLRFTRLLEQLGCRAGLLEPNAR-----
Lotco_MYB91__PHANb_ RIENACREQLGGLRRDAESKEQKLAEKWTSKHLRLTRLLEQMKIQTGAP-----
Lotco_MYB91__PHANa_ RIEAEYREQLAGLRRDAETKEQKLAEQWTVKHSRLMKFMEQIGCRSRIAETNGR-----
Eucgr_MYB91      KIETEYREQLAGLRKDAESKEQKLAEQWTAHVQLSKLIEQIGFRPRIADHDRQ-----
Maldo_MYB91      RIEAEYREQLAGLRRDAEAEQKLAEQWAAKHLRLSQFLEQMCCRPRIVEPNGR-----
Lyces_MYB91      RIEAEYKEQLAGLRRDAEAEQKLAEQWTSKHMRLAKFLEQMGCQSRLAEPNGGR-----
Soltu_MYB91      RIEAEYKDQLAGLRRDAEAEQKLAEQWTSKHMRLAKFLEQMGCQSRLAEPNGGR-----
Nicta_MYB91      RIEAEYKEQLAGLRRDAEAEQKLAEQWASKHLRLSKFLEQMGCQSRLAEPNGGR-----
Vitvi_MYB91      RIEAEYREQLAGLRRDAESKEQKLAEQWSAKHLRLTKFIEQMGCPRPRLAEPNGR-----
Goshi_MYB91      RIEAEYREQLVGLRRDAEAEQKLAEQWAAKHLHLTKFLEQTGCRPRVVEPNQ-----
Aqufo_MYB91      RIEAEYREQLAGLRRDADAQKLADQWAGKHMRLTKFLEQMGCPRPRLIEPNGR-----
Escca_MYB91      RMEADYRDQLAGLRRDAEAEQKLADQWAAKHLRLMKFLEQIGCRP-PSEPSGR-----
Arath_AS1_MYB91  KIEGEYREQLVGLRRDAEAEQKLADQWTSRHIRLTKFLEQMGCRRLDRP-----
Carhi_MYB91      KIDGEYREQLVGLRRDAEAKDQKLADQWTSKHIRLTKFLEQNMGCRLDRP-----
Brana_MYB91      KIEGEYREQLVGLRRDAEAEQKLADQWTSKHIRLTKFLEQHMGCQRLLDRP-----
Antma_MYB91      RIEAEYREQLAGLRRDAEAEQKLAEQWAAKHLRLTKFLEQTGYRSIAGELNGR-----
Orysa_MYB91      RVEAEYREKMAGLRRDAEAEQKLAEQWAAKHARLAKFLDQVAACRRWPPVEINGGGGG
Zeama_MYB91__RS2_ RVERDHRKVAELRRDAQVKEEKMAEQWAAKHARVAKFVEQMGGCSRWSWSSATDMNC---
Moral_MYB91      RIEGEYREQLLALQORDAEAEAKLVEAWCGKHVKLAKLLDQIGAHCCNATNGFTAFPNP
:::  :::  *:::*: *: *: : * :* :. :::*
XXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX
```

Conserved Domain (CD)

```
Poptr_MYB91      -----
Medtr_MYB91__PHAN_ -----
Pissa_MYB91      -----
Glyma_MYB91__PHANa_ -----
Glyma_MYB91__PHANb_ -----
Lotco_MYB91__PHANb_ -----
Lotco_MYB91__PHANa_ -----
Eucgr_MYB91      -----
Maldo_MYB91      -----
Lyces_MYB91      -----
Soltu_MYB91      -----
Nicta_MYB91      -----
Vitvi_MYB91      -----
Goshi_MYB91      -----
Aqufo_MYB91      -----
Escca_MYB91      -----
Arath_AS1_MYB91  -----
Carhi_MYB91      -----
Brana_MYB91      -----
Antma_MYB91      -----
Orysa_MYB91      PGGGR
Zeama_MYB91__RS2_ -----
Moral_MYB91      N-----
```

FIGURE 5 (continued)

**FIGURE 6**

MEKTL~~SLVLILPLLIMLLLVGTHAKIII~~

ESPAPQPQPPNTLPMNGTTPGSLHPQDC

LPKCTYRCSNTQYRKPCMFFCQKCCAAC

LCVPAGTYGNKQFCPCYNNWKTKRGGPK

CP

FIGURE 7

CLUSTAL 2.0.9 multiple sequence alignment

```
Os05g0432200 -----
AK110640 -----
TA53297_4565 -----
TA52915_4565 -----
scaff_41.75 -----
TA52374_4081 -----
TA5035_4679 -----
Os09g0414900 -----
GASA6 -----
scaff_XVII.377 -----
TA56938_4081 -----
GASA4 -----
Os05g0376800 -----
scaff_VI.397 -----
scaff_I.1483 -----
BG128975 -----
BG130916 -----
TA52635_4081_SEQID2_ -----
TA5923_4679 -----
Os06g0266800 -----
TA100367_4565 -----
CA725087 -----
TA77646_4565 -----
TA92393_4565 -----
CK153563 -----
BI208422 -----
TA37180_4081 -----
scaff_II.2328 -----
scaff_II.2330 -----
GASA5 -----
GASA12 -----
Os10g0115550 -----
TA101332_4565 -----
TA56201_4081 -----
AJ785329 -----
AK105729 -----
Os03g0760800 -----
TA66036_4565 -----
BM136027 -----
CA705831 -----
CA593033 -----
TA66038_4565 -----
CD899399 -----
Os03g0607200 -----
scaff_IX.735 -----
scaff_I.2410 -----
Pop_GASA_ -----
scaff_40.379 -----
TA45751_4081 -----
scaff_205.30 -----
TA69823_4565 MYVGFNXXWRFTXNDKXHIINVKAXXCHICSNQNKELPAPKSSNDDFTLSLCDISMQGTG
TA69821_4565 -----
Os07g0592000 -----
Os04g0465300 -----
scaff_II.204 -----
scaff_II.202 -----
TA35962_4081 -----
scaff_II.203 -----
BE353147 -----
TA41886_4081 -----
scaff_XII.704 -----
scaff_XV.507 -----
TA48119_4081 -----
Mt_GASA_ -----
scaff_I.1926 -----
scaff_XIX.758 -----
TA36295_4081 -----
TA95153_4565 -----
TA51752_4565 -----
```

FIGURE 8

```
Os05g0432200 -----MASMAKSLLCISLVAILLL-----
AK110640 -----MASMAKSLLCISLVAILLL-----
TA53297_4565 -----MAGQARAFMCVALVVLLLL-----
TA52915_4565 -----MAGKARVFMCVALLVLLLL-----
scaff_41.75 -----MVSAKTTFTLAILCLALMH-----
TA52374_4081 -----MAISKFLLVTMVLISLLVFRPVE-----
TA5035_4679 -----
Os09g0414900 -----WSFREMALAGRLLVLFAIALLAISIA-----
GASA6 -----MAKLITSFLLLTILF-----
scaff_XVII.377 -----MAK-FVAVFLLALIAISML-----
TA56938_4081 -----MAK-IVSVLLALLVISML-----
GASA4 -----MAKSYGAIFLLTLIVLFML-----
Os05g0376800 -----MEGVGVGVIRIRALLCCIAMAMLSSYQ-----
scaff_VI.397 -----
scaff_I.1483 -----MGKSSIAIFLCSSLVLVLGQNG-----
BG128975 -----MAGKMSIVLVFLLVFLTQNG-----
BG130916 -----
TA52635_4081_SEQID2_ -----MEKTLSLVLILPLLIMLLLVGTH-----
TA5923_4679 -----MANSTCILLSLHLLLIATAIQ-----
Os06g0266800 -----
TA100367_4565 -----MVTKVICFLVLASVLLAVAFVPSAL-----
CA725087 -----MAKISFLLVALLVLAVAFP-----
TA77646_4565 -----MAKISFLLVALLVLAVAFP-----
TA92393_4565 -----MAKISFLLVALLVLAVGFP-----
CK153563 -----MAKISFLFVALLVLAVAFP-----
BI208422 -----
TA37180_4081 -----MAKSGYNASFLLIS-----
scaff_II.2328 -----MASLSRNS--LLVVL-----
scaff_II.2330 -----MDPETALELVKQGATLLLLDVPQYTLVGI-----
GASA5 -----MANCIRRNALFFLT-----
GASA12 -----MMKLIVVFVISLLFATQFS-----
Os10g0115550 -----MDPASRSLSIIFFLVAVTF-----
TA101332_4565 -----MACVARTLSIP-FLALAFF-----
TA56201_4081 -----MRLHIFLALLIMAS-----
AJ785329 -----
AK105729 -----MDTLHNTPTLKLLAWSLGPFTSTMKLN-----TTTTLALLLL-----
Os03g0760800 -----MKLN-----TTTTLALLLL-----
TA66036_4565 -----MKLGP-----TATTVALLLV-----
BM136027 -----MKLGP-----TATTVALLLV-----
CA705831 -----MKCLR-----TTTTLALLLL-----
CA593033 -----MKCLR-----TTTTLALLLL-----
TA66038_4565 -----MKKLRTTTATTTLALILLL-----
CD899399 -----MKKLHTTTATTTLALLLL-----
Os03g0607200 -----MKTRR-----AALLMLLLLV-----
scaff_IX.735 -----
scaff_I.2410 -----MQAPSLF-----VFIYLVLE-----
Pop_GASA -----MKKLF-----FVTLLLC-----
scaff_40.379 -----MKPVF-----AAIFLLC-----
TA45751_4081 -----
scaff_205.30 -----MKLSF-----AALLLLSV-----
TA69823_4565 SRNQTRELRCIFTSFPQPNKRLLLPNISWTKLKKRRTAMKPLP-----VTLALLA-----
TA69821_4565 -----MKPLP-----VTLALLA-----
Os07g0592000 -----MRVPPLR-----ATTALLAT-----
Os04g0465300 -----MAPGKLAVFALLASLLLN-----
scaff_II.204 -----MAISKLLIASLVSLVLHLAE-----
scaff_II.202 -----MAISKLLIASLVVSLVL-----
TA35962_4081 -----MAISKALFASLLSLLEQVQS-----
scaff_II.203 -----MLIWFRFS-----
BE353147 -----MTVQKAFVAMLIASFLLVHFANA-----
TA41886_4081 -----MASLKGFALLIASLVLVHFTYA-----
scaff_XII.704 -----
scaff_XV.507 -----MAVRSLLALMVVFVCL-----
TA48119_4081 -----MAMALRVLLLVFLTVKAQDS-----
Mt_GASA -----MERKITLLILMVALLFCMT-----
scaff_I.1926 -----MAFKAVCLMVVAFVLVTAKASYM-----
scaff_XIX.758 -----MQIT-----FNMQMD-----
TA36295_4081 -----MKIFTFLFILLIQVFANAATE-----
TA95153_4565 -----MAPGKQ-----LLPPLLLMLL-----
TA51752_4565 -----MAPTQORLLTRRLLPPLLLLLLLA-----
```

FIGURE 8 (continued)

Os05g0432200	-----VETTAPHGQAY-----
AK110640	-----VETTAPHGQAY-----
TA53297_4565	-----VETTAPSGQAH-----
TA52915_4565	-----VETTAPSGQAH-----
scaff_41.75	-----ELQIRTVBAGKIN-----
TA52374_4081	ANGNGDG-----DNLAVHTAGPEGANNPTYIP-----
TA5035_4679	-----MLVESAKNGY-----
Os09g0414900	-----EHKALAKGSTSEHDDNVYQV-----
GASA6	-----TFVCLTMSKEAEYHPESY-----
scaff_XVII.377	-----QTLVVASHGRGGHHNNKN-----
TA56938_4081	-----ANTVMAANGK--HHHYAK-----
GASA4	-----QTMVMASSGS--NVKWSQK-----
Os05g0376800	QQQAEASYMPWPATPPPPAAAAANSTSTAAANNSSSSSSTTAPPQOPTAF-----
scaff_VI.397	-----MSRKPSIN-----ANITEAPTQPQPNTNSNRPP-----
scaff_I.1483	-----ALKTPISASQTQRQGNH-----
BG128975	VSR-----ANIMRDEQQQQRRNNQ-----
BG130916	-----MVLVRGRPPSRLSTK-----
TA52635_4081_SEQID2_	-----AKIIIESPAPQPQPPNTLP-----
TA5923_4679	VKHAHAP-----TLQPVNSTAPTAPQPNYPS-----
Os06g0266800	-----MAGGRGRGGGGGGGVAG-----
TA100367_4565	RQQ-----VKKGGGGEGGGGGSVSGS-----
CA725087	-----VEVMGGGNGGAGGGG-----
TA77646_4565	-----VEVMGGGNGGAGGGG-----
TA92393_4565	-----VEVMGGGGGGGGGGGG-----
CK153563	-----VKVMGVXXXG-----
BI208422	-----MFLILLTFSNVVEGY-----
TA37180_4081	-----MFLILLTFSNVVEGY-----
scaff_II.2328	-----SLCLLITFSNVAEIHG-----
scaff_II.2330	DTQ-----VLEIXXXXXXXXXXX-----
GASA5	-----LFLLSVSNLVQAARGG-----
GASA12	-----NGDELESQAQAPAIHKNG-----
Os10g0115550	-----VVEVSGQKNEAVYHLFG-----
TA101332_4565	-----VAEVSGSMNVESYKPG-----
TA56201_4081	-----MSRAQPPVGPPTTCP-----
AJ785329	-----
AK105729	-----LLLASSSLQVSMAG-----
Os03g0760800	-----LLLASSSLQVSMAG-----
TA66036_4565	-----LLLASSSLRAATAG-----
BM136027	-----LLLASSSLRAATGG-----
CA705831	-----VFLAASSLRAAMAG-----
CA593033	-----VFLAASSLRAAMAG-----
TA66038_4565	-----VLIAATSLRVAMAG-----
CD899399	-----VLLAATSLRVAMAG-----
Os03g0607200	-----VVAAASWPQPCDAA-----
scaff_IX.735	-----
scaff_I.2410	-----ILI-----MLVISCG-----
Pop_GASA	-----LLFSSSFLEPVMTK-----
scaff_40.379	-----LVFSSSLFEVTMAA-----
TA45751_4081	-----MAG-----
scaff_205.30	-----VLLSSFLRFTMAVPNHVA-----
TA69823_4565	-----LFLAASYQDLAVAA-ADA-----
TA69821_4565	-----LFLVASYQDLTVAADADA-----
Os07g0592000	-----LLVAASFQDLTVAA-----
Os04g0465300	-----TIKAADYPPAPPLGPPP-----
scaff_II.204	-----ADQKVNSNQAAASHVPG-N-----
scaff_II.202	-----QQVNASPAAGSIPG-K-----
TA35962_4081	I-----QTDQVSSNAISEGADSYK-----
scaff_II.203	-----
BE353147	-----QKVDYSKPPASAPQGPQ-----
TA41886_4081	L-----QEVISGKPPAPSPQPPK-----
scaff_XII.704	-----
scaff_XV.507	-----AELVVRRGN-----
TA48119_4081	IIDL-----KEVEEDKQHVGLSQALRVFTRGAN-----
Mt_GASA	-----KVLCADSSVHIQDQFTHFEVVKGP-N-----
scaff_I.1926	NEDF-----KEKAVFSKSVVPASTPAPPEVKSPTFAPPVVTPTSTPLYKPPTPAP-----
scaff_XIX.758	-----EESDVVAIDKKHY-----
TA36295_4081	Q-----IEAGNEGALHKKIHP-----
TA95153_4565	-HHQPAAG--ASDPPVT--HGGMRASTARSLQQQQQQ-----
TA51752_4565	AHLQPAAASSASDPLVTTTTAHGSMRAS-SRSLQQQ-----

FIGURE 8 (continued)

Os05g0432200	-----
AK110640	-----
TA53297_4565	-----
TA52915_4565	-----
scaff_41.75	-----
TA52374_4081	-----
TA5035_4679	-----
Os09g0414900	-----
GASA6	-----
scaff_XVII.377	-----
TA56938_4081	-----
GASA4	-----
Os05g0376800	-----
scaff_VI.397	-----
scaff_I.1483	-----
BG128975	-----
BG130916	-----
TA52635_4081_SEQID2_	-----
TA5923_4679	-----
Os06g0266800	-----
TA100367_4565	-----
CA725087	-----
TA77646_4565	-----
TA92393_4565	-----
CK153563	-----
BI208422	-----
TA37180_4081	-----
scaff_II.2328	-----
scaff_II.2330	-----
GASA5	-----
GASA12	-----
Os10g0115550	-----
TA101332_4565	-----
TA56201_4081	-----
AJ785329	-----
AK105729	-----
Os03g0760800	-----
TA66036_4565	-----
BM136027	-----
CA705831	-----
CA593033	-----
TA66038_4565	-----
CD899399	-----
Os03g0607200	-----
scaff_IX.735	-----
scaff_I.2410	-----
Pop_GASA_	-----
scaff_40.379	-----
TA45751_4081	-----
scaff_205.30	-----
TA69823_4565	-----
TA69821_4565	-----
Os07g0592000	-----
Os04g0465300	-----
scaff_II.204	-----
scaff_II.202	-----
TA35962_4081	-----
scaff_II.203	-----
BE353147	-----
TA41886_4081	-----
scaff_XII.704	-----
scaff_XV.507	-----
TA48119_4081	-----
Mt_GASA_	-----
scaff_I.1926	PVKTPPPAPPVNPPTPVKPPTTPAPPVYKPPSPAPPVNPPTPVKPPTTPAPPVYKPPSPA
scaff_XIX.758	-----
TA36295_4081	-----
TA95153_4565	-----
TA51752_4565	-----

FIGURE 8 (continued)

Os05g0432200	-----AIDCGAKCGY
AK110640	-----AIDCGAKCGY
TA53297_4565	-----AVDCGSACS
TA52915_4565	-----AVDCGSACS
scaff_41.75	-----CKSKCEY
TA52374_4081	-----TSECGTACEA
TA5035_4679	-----GQGSILRS-----YQCSGQCAR
Os09g0414900	-----SKGGQGSILKS-----YQCSPOCAY
GASA6	-----GPGSLKS-----YQCGGQCTR
scaff_XVII.377	-----KYGPGSLKS-----FQCPSPQCTR
TA56938_4081	-----KYGPGSLKP-----SQCLPQCTR
GASA4	-----RYGPGSLKR-----TQCPSECDR
Os05g0376800	-----PMYGVTPGSLRP-----QECGGRCA
scaff_VI.397	-----YGTTOGSLNP-----QECGPRCTG
scaff_I.1483	-----AMYGATOGSLRP-----QECAPRCTT
BG128975	-----LYGVSEGRLHP-----QDCQPKCTY
BG130916	-----MHRT-----
TA52635_4081_SEQID2_	-----MNGTTPGSLHP-----QDCLPKCTY
TA5923_4679	-----HGFTEGSLQP-----QECGGRCDV
Os06g0266800	-----GGNLRP-----WECSPKCAG
TA100367_4565	-----GGGNLNP-----WECSPKCGS
CA725087	-----KLKP-----WECSSKCSS
TA77646_4565	-----KLKP-----WECSSKCSS
TA92393_4565	-----NLKP-----WECSSKCSS
CK153563	-----KLKP-----WECPSKCSS
BI208422	-----NKLRP-----TDCKPRCTY
TA37180_4081	-----NKLRP-----TDCKPRCTY
scaff_II.2328	-----AKLRP-----SCKKPRCNY
scaff_II.2330	-----XXXXXXXXXEATSLISPASTRCNY
GASA5	-----GKLKP-----QQCNSKCSF
GASA12	-----GEGSLKP-----BECPKACEY
Os10g0115550	-----GEGSLTK-----NECPGKCSY
TA101332_4565	-----AEGSVPL-----KECPAKCKI
TA56201_4081	-----VPK-----DKCEBACNV
AJ785329	-----MDI-----
AK105729	-----SDPCDGKCKV
Os03g0760800	-----SDPCDGKCKV
TA66036_4565	-----SAFCDGKCGV
BM136027	-----SAFCDGKCGV
CA705831	-----SAFCDGKCGV
CA593033	-----SAFCDGKCGV
TA66038_4565	-----SAFCDGKCGV
CD899399	-----SAFCDGKCGV
Os03g0607200	-----SGFCGSKCAV
scaff_IX.735	-----
scaff_I.2410	-----VAFCTKKCNT
Pop_GASA_	-----SSFCAKKCDT
scaff_40.379	-----SGFCDGKCSV
TA45751_4081	-----SYFCDGKCKL
scaff_205.30	-----SPPPPS-----PAIPSFCDPKCKA
TA69823_4565	-----DADGVG-----SGAPVLDSVCEGCKN
TA69821_4565	-----DAAGAGDVGAVPVPDSVCEGCKN
Os07g0592000	-----DGGG-----GVVPVPDSVCDACKCQK
Os04g0465300	-----HKIVDP-----GKDCVGACDA
scaff_II.204	-----NIDCGGACHA
scaff_II.202	-----NIDCGGACKD
TA35962_4081	-----KIDCGGACAA
scaff_II.203	-----DCGSACKA
BE353147	-----PLDCIGACKY
TA41886_4081	-----PIDCTGSCKT
scaff_XII.704	-----MDSIRY-----CGGLCKQ
scaff_XV.507	-----RRLMQDID-----CGGLCKQ
TA48119_4081	-----RRLVQDIVLKVAKYLNNGDI-----ALAPAPAPPPSPLDGGLCKY
Mt_GASA_	-----RRLAFVD-----CGTRCNV
scaff_I.1926	PPVNPPTPVPPVKPPTAPAPPVYKPPSPAPTVPVPPVKPPTGPMPPPVTRSDCTPLCGQ
scaff_XIX.758	-----KRINCGYLAR
TA36295_4081	-----KRIHCGYACAR
TA95153_4565	-----QPPRLDCPKVCLG
TA51752_4565	-----PPFRLDCPKVCLG

FIGURE 8 (continued)

* ** * ** *

FIGURE 8 (continued)

Os05g0432200	----KCP-----
AK110640	----KCP-----
TA53297_4565	----KCP-----
TA52915_4565	----KCP-----
scaff_41.75	----KCP-----
TA52374_4081	----KCP-----
TA5035_4679	----KCP-----
Os09g0414900	----KCP-----
GASA6	----KCP-----
scaff_XVII.377	----KCP-----
TA56938_4081	----KCP-----
GASA4	----KCP-----
Os05g0376800	----KCP-----
scaff_VI.397	----KCP-----
scaff_I.1483	----KCP-----
BG128975	----KCP-----
BG130916	----KCP-----
TA52635_4081_SEQID2_	----KCP-----
TA5923_4679	----KCP-----
Os06g0266800	----KCP-----
TA100367_4565	----KCP-----
CA725087	----KCPRIEFPPSSSSGGATCG----
TA77646_4565	----KCP-----
TA92393_4565	----KCP-----
CK153563	----KCP-----
BI208422	----KCP-----
TA37180_4081	----KCP-----
scaff_II.2328	----KCP-----
scaff_II.2330	----KCP-----
GASA5	----KCP-----
GASA12	----KCP-----
Os10g0115550	----VCELGIEEKRNDTGE----
TA101332_4565	----KCP-----
TA56201_4081	----KCP-----
AJ785329	----KCP-----
AK105729	--RPKCP-----
Os03g0760800	--RPKCP-----
TA66036_4565	--RPKCP-----
BM136027	--RPKCP-----
CA705831	KIRQQSPXIDSPTPMGSKQ----
CA593033	MIRQHSPSIDSPTAMGSKKHIXLKLHATLSNQVL
TA66038_4565	--RPKCP-----
CD899399	--RPKCP-----
Os03g0607200	--RPKCP-----
scaff_IX.735	--KPKCP-----
scaff_I.2410	--KPKCP-----
Pop_GASA_	--KPKCP-----
scaff_40.379	--KPKCP-----
TA45751_4081	--KSKCP-----
scaff_205.30	--TSKCP-----
TA69823_4565	--RPKCP-----
TA69821_4565	--RPKCP-----
Os07g0592000	--RPKCP-----
Os04g0465300	----KCP-----
scaff_II.204	----KCP-----
scaff_II.202	----KCP-----
TA35962_4081	----KCP-----
scaff_II.203	----KCP-----
BE353147	----KCP-----
TA41886_4081	----KCP-----
scaff_XII.704	----KCP-----
scaff_XV.507	----KCP-----
TA48119_4081	----KCP-----
Mt_GASA_	----KCP-----
scaff_I.1926	----KCP-----
scaff_XIX.758	----KCP-----
TA36295_4081	----KCP-----
TA95153_4565	----KCP-----
TA51752_4565	----KCP-----

FIGURE 8 (continued)

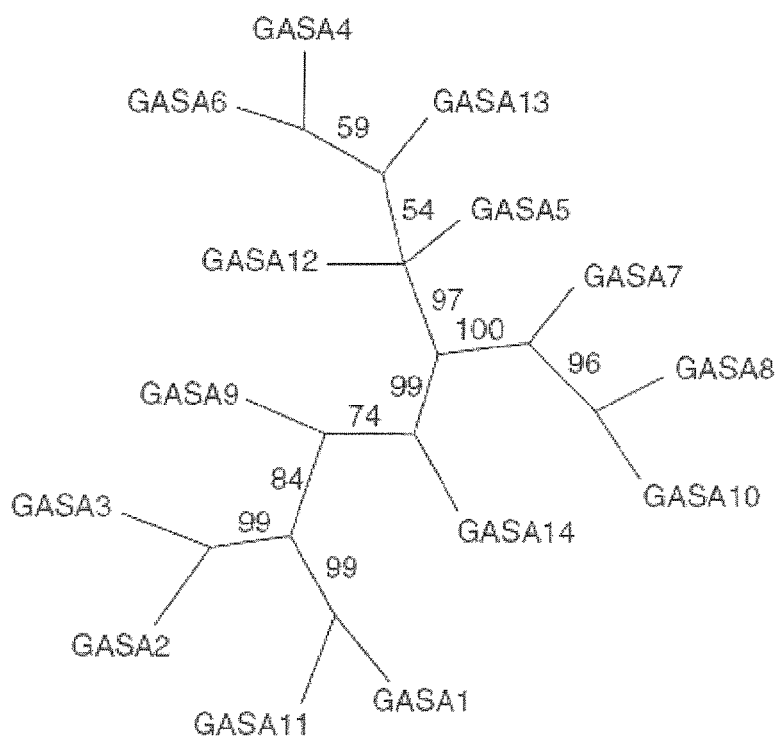


FIGURE 9
Terminator

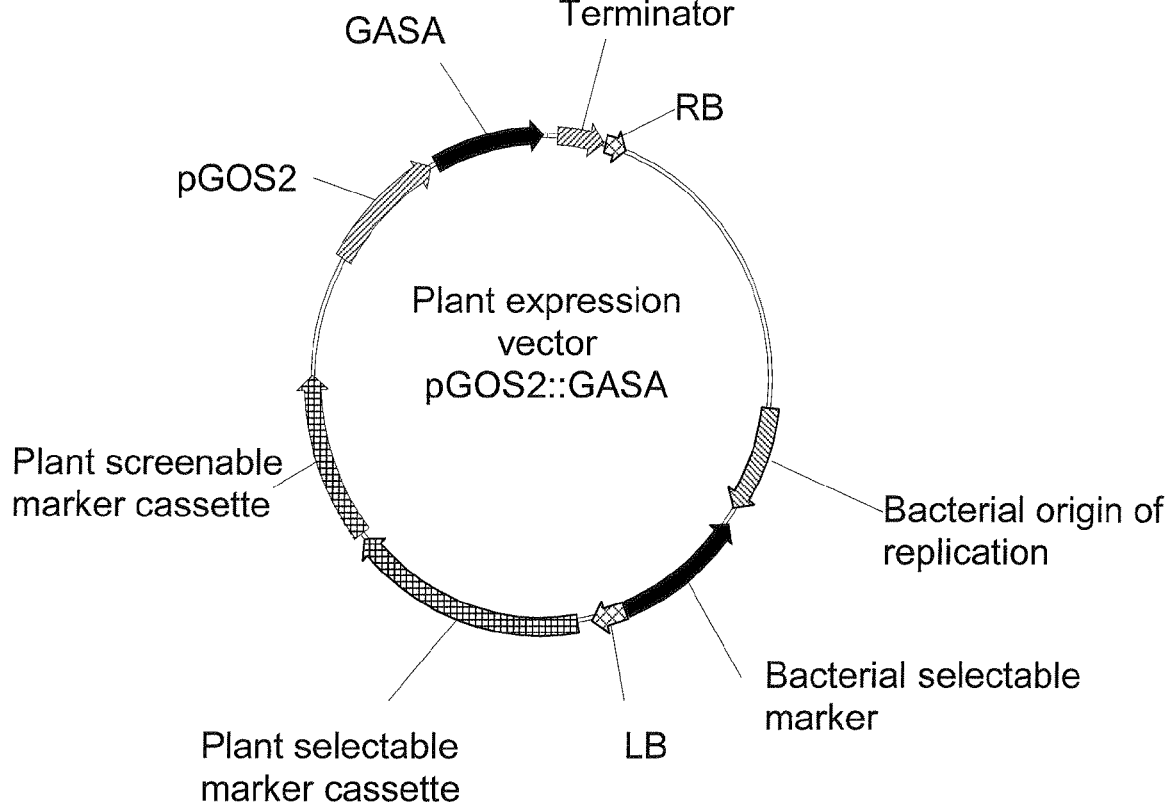


FIGURE 10

	1	50
seqidno02, PRT, Oryzasativa>	(1)	-----
seqidno2, PRT, Arabidopsisthaliana>	(1)	-----
seqidno14, PRT, Arabidopsisthaliana>	(1)	-----
seqidno24, PRT, Arabidopsisthaliana>	(1)	-----
seqidno228, PRT, Zeamays>	(1)	-----
seqidno84, PRT, Oryzasativa>	(1)	-----
seqidno102, PRT, Oryzasativa>	(1)	-----
seqidno108, PRT, Oryzasativa>	(1)	-----
seqidno126, PRT, Oryzasativa>	(1)	-----
seqidno180, PRT, Oryzasativa>	(1)	-----
seqidno22, PRT, Arabidopsisthaliana>	(1)	-----
seqidno54, PRT, Arabidopsisthaliana>	(1)	-----
seqidno120, PRT, Oryzasativa>	(1)	-----MAPPQER-DYIGLSP-----
seqidno144, PRT, Oryzasativa>	(1)	-----MSPPLEL-DYIGLSPPPPP--FSS
seqidno72, PRT, Oryzasativa>	(1)	-----MSPPLEL-DYIGLSPPPPP--FSS
seqidno210, PRT, Zeamays>	(1)	-----MSPPLDL-DYIGLSP-----
seqidno174, PRT, Oryzasativa>	(1)	-----MPPPLEARDYIGLGATPAS--SSS
seqidno198, PRT, Zeamays>	(1)	-----
seqidno138, PRT, Oryzasativa>	(1)	-----
seqidno192, PRT, Oryzasativa>	(1)	-----
seqidno234, PRT, Zeamays>	(1)	-----
seqidno156, PRT, Oryzasativa>	(1)	-----
seqidno90, PRT, Oryzasativa>	(1)	-----
seqidno162, PRT, Oryzasativa>	(1)	-----
seqidno216, PRT, Zeamays>	(1)	-----
seqidno36, PRT, Arabidopsisthaliana>	(1)	-----
seqidno48, PRT, Arabidopsisthaliana>	(1)	-----
seqidno66, PRT, Arabidopsisthaliana>	(1)	MSPEEELQSNVSVASSSPTSNCISRNTLGGLKEHNYLGLSDCSSVGSSTL
Consensus	(1)	-----
	51	100
seqidno02, PRT, Oryzasativa>	(1)	-----
seqidno2, PRT, Arabidopsisthaliana>	(1)	-----
seqidno14, PRT, Arabidopsisthaliana>	(1)	-----MYCS
seqidno24, PRT, Arabidopsisthaliana>	(1)	-----
seqidno228, PRT, Zeamays>	(1)	-----
seqidno84, PRT, Oryzasativa>	(1)	-----
seqidno102, PRT, Oryzasativa>	(1)	-----
seqidno108, PRT, Oryzasativa>	(1)	-----
seqidno126, PRT, Oryzasativa>	(1)	-----
seqidno180, PRT, Oryzasativa>	(1)	-----
seqidno22, PRT, Arabidopsisthaliana>	(1)	-----
seqidno54, PRT, Arabidopsisthaliana>	(1)	-----
seqidno120, PRT, Oryzasativa>	(15)	-----AAAAALATELRLGLPGTAEAESEGGGG-----
seqidno144, PRT, Oryzasativa>	(22)	SSAAAAARADDVDLKGTELRLGLPGSESPDRHPAAIA-----
seqidno72, PRT, Oryzasativa>	(22)	SSAAAAARADDVDLKGTELRLGLPGSESPDRHPAAIA-----
seqidno210, PRT, Zeamays>	(15)	--AAAAAAHDDLKGTELRLGLPGSGSPDRR-----
seqidno174, PRT, Oryzasativa>	(23)	SCCASTPVAEVVGAHLALRLGLPGSESPARAEAEAVV-----
seqidno198, PRT, Zeamays>	(1)	-----
seqidno138, PRT, Oryzasativa>	(1)	-----MAADLAFEATELRLGLPGGGGDGDA-----
seqidno192, PRT, Oryzasativa>	(1)	-----MAADLAFEATELRLGLPGGGGDGDA-----
seqidno234, PRT, Zeamays>	(1)	-----MATTTDLGFEATELRLGLPGGGGGGEP-----
seqidno156, PRT, Oryzasativa>	(1)	-----
seqidno90, PRT, Oryzasativa>	(1)	-----MAGLGFDETELRLGLPGAG-----
seqidno162, PRT, Oryzasativa>	(1)	-----MAGADVVDVGTTELRLGLPGGGG-G-----
seqidno216, PRT, Zeamays>	(1)	-----MAGADVVDVGTTELRLGLPGGG-----
seqidno36, PRT, Arabidopsisthaliana>	(1)	-----MINFEATELRLGLPGGN-----
seqidno48, PRT, Arabidopsisthaliana>	(1)	-----MIGQLMNLKATELCLGLPGGA-----
seqidno66, PRT, Arabidopsisthaliana>	(51)	SPLAEDDKATISLKATELTLGLPGSQSPARDTELNLLSPAKLDEKPFPPPL
Consensus	(51)	L LGLPG

FIGURE 11

```
seqidno02, PRT, Oryzasativa>
seqidno2, PRT, Arabidopsisthaliana>
seqidno14, PRT, Arabidopsisthaliana>
seqidno24, PRT, Arabidopsisthaliana>
seqidno228, PRT, Zeamays>
seqidno84, PRT, Oryzasativa>
seqidno102, PRT, Oryzasativa>
seqidno108, PRT, Oryzasativa>
seqidno126, PRT, Oryzasativa>
seqidno180, PRT, Oryzasativa>
seqidno22, PRT, Arabidopsisthaliana>
seqidno54, PRT, Arabidopsisthaliana>
seqidno120, PRT, Oryzasativa>
seqidno144, PRT, Oryzasativa>
seqidno72, PRT, Oryzasativa>
seqidno210, PRT, Zeamays>
seqidno174, PRT, Oryzasativa>
seqidno198, PRT, Zeamays>
seqidno138, PRT, Oryzasativa>
seqidno192, PRT, Oryzasativa>
seqidno234, PRT, Zeamays>
seqidno156, PRT, Oryzasativa>
seqidno90, PRT, Oryzasativa>
seqidno162, PRT, Oryzasativa>
seqidno216, PRT, Zeamays>
seqidno36, PRT, Arabidopsisthaliana>
seqidno48, PRT, Arabidopsisthaliana>
seqidno66, PRT, Arabidopsisthaliana>
Consensus

101
(1) -----MEEKKRLELRLAPPCHQF
(1) -----MEEKKRLELRLAPPCHQF
(5) DPPHPLHLVASDKQQKDHKLILSWKKPTMDSDDLGVFPNSPKYHPYYSQT
(1) -----MDPNTPADFFKGSSEKPHYYYSQT
(1) -----MRETRTESYSASINKAPTEKKQESTTSGCRLFGIEI
(1) -----MELELGLAPPNSGHLVVDLSSSSSSSGGGS
(1) -----MSVETERSTESSAASGLDFEDTALTLLRPGSSS
(1) -----
(1) -----MSTSSGADSSPPVSGLDYDDTALTLLRPGSSS
(1) -----MSTSSGADSSPPVSGLDYDDTALTLLRPGSSS
(1) -----MSPEEYVRVWPDSDGLGGTELTLALPGTPT
(1) -----MEVTNGLNLKDTLRLGLPGAQ-
(44) -----GTDAAPLTLELLPKGGAKRGFADAIVGGPAGQRR
(58) -----AAAATATTLELLPAKGAKRVPDEAALTPTPT-
(58) -----AAAATATTLELLPAKGAKRVPDEAALTPTPT-
(44) -----VVAATATLDLLPAKGAKRGFSEAPTSPG-
(60) -----VDAALTGLGFAPPFRGGAKRGFVDSLDRSEGR-RA
(1) -----
(26) -----A-----AAAAR-----SSSGKRGFAETIDLKLKLEPA
(26) -----A-----AAAAR-----SSSGKRGFAETIDLKLKLEPA
(28) -----A-----LGGEGRSSSSASGKRGFAETIDLKLKLEPA
(1) -----
(20) -----E-----LAARS-----SGKRGFAETIDLKLKLEPA
(23) -----AEEAAAKAARGFEETIDLKLKLEPA
(21) -----ADAATAAKRGFEDTIDLKLKLEPA
(18) -----HGGEMAGKNNG--KRGFSETVDLKLNLSSST
(22) -----EAVESPAKSAVSGKRGFSETVDLKLNLQSN
(101) LPSKDEICSSSQKNNASGNKRGFSDTMDQFAEAKSSVYTEKNMFPFAA-
(101) GKR F E L L L

151
(19) TS-----NNNIN
(19) TS-----NNNIN
(55) TFFGG-----VIDLGLS
(24) KKGKG-----VIDLGLS
(37) G-----SSAVSP
(31) GS-----APVSASSAGKRG-----FREAPQET
(35) SSSSSS-----SSSSSS-----PSEPDRKRASA
(1) -----
(33) SSSS-----TADPERKRAAH
(33) SSSS-----TADPERKRAAH
(31) NAS-----EGPKKFG-----NKRRFLE
(23) -----EEQQLE-----LSCVR
(78) E-----AAGGK-----AAAA
(89) -----AAAGKKG-----AAREG
(89) -----AAAGKKG-----AAREG
(75) -----AASGK-----G
(93) A-----ATAGDDE-----RGVRE
(1) -----
(53) AAAVDDDDDKKEAAADDREKKVDIVGADNDDASPPAAAAAGMKRSPSQS
(53) AAAVDDDDDKKEAAADDREKKVDIVGADNDDASPPAAAAAGMKRSPSQS
(59) AVVEAEEEEEDHGVAVALEK-----EEE-AGMKRSPSQS
(1) -----
(45) APAAVSGEEGAQEDKEDADA-----AAAADEKMSMKRSASQS
(49) G-----MEEA--AAGKAEAP-----AAEKA
(45) G-----MEEA--AAAAAAKPE-----P--AAEKA
(46) A-----MD-----SVSKV
(52) K-----EG-----SVDLK
(150) -----ATQSVT-----KKDVP
(151) -----
```

FIGURE 11 (continued)

	201	250
seqidno02, PRT, Oryzasativa>	(26) GSKQKSSTKETSFLSNNRVEVAPVVGWPPVRSRRNLTAQLKEEMKKKES	
seqidno2, PRT, Arabidopsisthaliana>	(26) GSKQKSSTKETSFLSNNRVEVAPVVGWPPVRSRRNLTAQLKEEMKKKES	
seqidno14, PRT, Arabidopsisthaliana>	(67) LRTIQHEIYHSSG-----Q	
seqidno24, PRT, Arabidopsisthaliana>	(36) LRTIQHETYLPPARMIGLDGYGELIDWSQPSYNSITQLKSEDTHGQRLAQ	
seqidno228, PRT, Zeamays>	(44) VVTVASVGHDP PPPALSVDAESDQLSQPSHANKATDAPAASSDRSPNETE	
seqidno84, PRT, Oryzasativa>	(53) LLLFDDGSCCNTSDDDCRRRKKT VVGWPPVSSARR---ACG-----	
seqidno102, PRT, Oryzasativa>	(59) TDDDPDNLGSTATESPSPKARVVGWPPVRAFRKNALALAAASS----	
seqidno108, PRT, Oryzasativa>	(1) ---MAR-----RGGRRARVVGWPPVRAFRKNALALAAASS----	
seqidno126, PRT, Oryzasativa>	(48) ADHADAK-----PSPKARAVGWPPVRAFRKNALREDSAR-----	
seqidno180, PRT, Oryzasativa>	(49) ADHADAK-----PSPKARAVGWPPVRAFRKNALREDAAR-----	
seqidno22, PRT, Arabidopsisthaliana>	(48) TVDLKLGEAHENNYISSMVTNDQLVGWPPVATARKTVR-----	
seqidno54, PRT, Arabidopsisthaliana>	(34) SNN-KRKNNDSTEE SAPPPAKTQIVGWPPVRSNRK-----	
seqidno120, PRT, Oryzasativa>	(88) AAEAEHEEEKKKAQAP--AAKAQVVGWPP IRSYRKNMTAMSQPALKGKDD	
seqidno144, PRT, Oryzasativa>	(101) EEVGAEEDDKVAAPPQPAKAQVVGWPP IRSYRKNMTATNQIKSN-KED	
seqidno72, PRT, Oryzasativa>	(101) EEVGAEEDDKVAAPPQPAKAQVVGWPP IRSYRKNMTATNQIKSN-KED	
seqidno210, PRT, Zeamays>	(81) KKVAEEEDDKVAATPQPVAKAQVVGWPP IRSYRKNMTSTTQLKGS-KED	
seqidno174, PRT, Oryzasativa>	(106) EEEEEKGLGEAAAGAPRAAKAQVVGWPPVRSYRKNTLAASATKTKGEDQ	
seqidno198, PRT, Zeamays>	(1) -----	
seqidno138, PRT, Oryzasativa>	(103) --SVVTAADPE---KPRAPKAQVVGWPPVRSYRKNILAVQADK--GKDA	
seqidno192, PRT, Oryzasativa>	(103) --SVVTAADPE---KPRAPKAQVVGWPPVRSYRKNILAVQADK--GKDA	
seqidno234, PRT, Zeamays>	(93) SVAAAVLADPAE--KPRAPKAQVVGWPPVRSFRKNIMSVQSDKGAGKDA	
seqidno156, PRT, Oryzasativa>	(1) -----MSIRAQVVGWPPVRSFRKNVLAEKCKA-----	
seqidno90, PRT, Oryzasativa>	(83) --SVVTAEPDPD---KPRAPKAQVVGWPPVRSFRKNVLAEKCKA-----	
seqidno216, PRT, Zeamays>	(67) KRPAEAAAADAE---KPPAPKAQAVGWPPVRSFRKNIMTVQSVKSKKEEE	
seqidno36, PRT, Arabidopsisthaliana>	(65) KRPAEAPAADAE---KPPAPKAQAVGWPPVRSYRRNMTVQSVKSKKEEE	
seqidno48, PRT, Arabidopsisthaliana>	(54) DLENMKEK-----VVKPPAKAQVVGWPPVRSFRKNVMSGQKPTTGDATE	
seqidno66, PRT, Arabidopsisthaliana>	(60) NVSAVPKEKTTLLKDP SKPPAKAQVVGWPPVRSYRKNMTTQKTS SSGAEAA	
Consensus	(161) QNIPKGQSSSTNNSSSPPAAKAQIVGWPPVRSYRKNLTATTCNSD----	
	(201) A KAQVVGWPPVRSYRKN LA	
	251	300
seqidno02, PRT, Oryzasativa>	(76) DEEK-----ELYVKINMEGVPIGRKVNLSAYNNYQQLSH	
seqidno2, PRT, Arabidopsisthaliana>	(76) DEEK-----ELYVKINMEGVPIGRKVNLSAYNNYQQLSH	
seqidno14, PRT, Arabidopsisthaliana>	(81) RYCSNEG-----YRRKWGVVKTMDGLVVGKVKCVLDHGSYSTLAH	
seqidno24, PRT, Arabidopsisthaliana>	(86) GYYNNEGE-----SRGKYAYVKVNLGLVVGKVKCVLDQGAATLAL	
seqidno228, PRT, Zeamays>	(94) -SR-----QARSCTKVMQGVAVGRAVDLTRLDGYDDLRR	
seqidno84, PRT, Oryzasativa>	(91) -----GANYVVKVKEGDAIGRKVDLALHSSYDELA	
seqidno102, PRT, Oryzasativa>	(105) -----SKAKFVKVAVDGA PYLRKVLDL EAYRGYDQLLA	
seqidno108, PRT, Oryzasativa>	(34) -----SKAKFVKVAVDGA PYLRKVLDL EAYRGYDQLLA	
seqidno126, PRT, Oryzasativa>	(83) -----AKLVKVAVDGA PYLRKVLDL AAHAGYAPLLR	
seqidno180, PRT, Oryzasativa>	(84) -----AKLVKVAVDGA PYLRKVLDL AAHAGYAPLLR	
seqidno22, PRT, Arabidopsisthaliana>	(86) -----R---KYVKVALDGAAYLRKVLDL GMYDCYQGLFT	
seqidno54, PRT, Arabidopsisthaliana>	(68) ---NNNN-----KNVS YVKV SMDGAPYLRKID LKMYKNYP ELLK	
seqidno120, PRT, Oryzasativa>	(136) GEAKQAPA-----SGCLYVKV SMDGAPYLRKVLD LKMYKNYK ELSL	
seqidno144, PRT, Oryzasativa>	(150) VDAKQG-----QGFLYVKV SMDGAPYLRKVLD LKTYKNYK DMSL	
seqidno72, PRT, Oryzasativa>	(150) VDAKQG-----QGFLYVKV SMDGAPYLRKVLD LKTYKNYK DMSL	
seqidno210, PRT, Zeamays>	(130) AEAKQD-----QGFLYVKV SMDGAPYLRKID LKTYKNYK DLSL	
seqidno174, PRT, Oryzasativa>	(156) GKSEVG-----C--CYVKV SMDGAPYLRKVLD LKTYSSYEDLSL	
seqidno198, PRT, Zeamays>	(1) -----MYVKV SMDGAPYLRKVID I KMYSSYEDLSV	
seqidno138, PRT, Oryzasativa>	(146) ADGGGDKS---GAGAAA---AAFVKV SMDGAPYLRKVLD LKMYKSYLELSK	
seqidno192, PRT, Oryzasativa>	(146) ADGGGDKS---GAGAAA---AAFVKV SMDGAPYLRKVLD LKMYKSYLELSK	
seqidno234, PRT, Zeamays>	(141) AAANGDKS---SAAAGG---GA AFVKV SLDGAPYLRKVLD LKMYRSYQQLSK	
seqidno156, PRT, Oryzasativa>	(28) -----AALVKV SMDGAPYLRKIDV AMYKSYPELSM	
seqidno90, PRT, Oryzasativa>	(122) -----AALVKV SMDGAPYLRKIDV AMYKSYPELSM	
seqidno162, PRT, Oryzasativa>	(114) ADKQQQQP---AANASGSNSSAFVKV SMDGAPYLRKVLD LKMYNSYK DLSL	
seqidno216, PRT, Zeamays>	(112) PEKQKS-----AANAGG-NGSAFVKV SMDGAPYLRKVLD LKMYNSYK DLSV	
seqidno36, PRT, Arabidopsisthaliana>	(98) GNDKTS GSSGATSSASACATVAYVKV SMDGAPYLRKID LKLYKTYDLSN	
seqidno48, PRT, Arabidopsisthaliana>	(110) SSKAG-----NFGGGAAGALVKV SMDGAPYLRKVLD LKMYKSYDLSL	
seqidno66, PRT, Arabidopsisthaliana>	(207) -EVDGRPG-----SGALFVKV SMDGAPYLRKVLD LRSYTNYGELSS	
Consensus	(251) A YVKV SMDGAPYLRKVLD LKMYK Y DLS	

FIGURE 11 (continued)

	301	350
seqidno02, PRT, Oryzasativa>	(110) AVDQLFSKKDSWDLN-----RQYTLVYEDTEGDK	
seqidno2, PRT, Arabidopsisthaliana>	(110) AVDQLFSKKDSWDLN-----RQYTLVYEDTEGDK	
seqidno14, PRT, Arabidopsisthaliana>	(122) QLEDMFGMQSVSGLR-----LFQMESEFCLVYRDEEGLW	
seqidno24, PRT, Arabidopsisthaliana>	(128) QLNDMFGMQTVSGLR-----LFQTESEFSLVYRDREGIW	
seqidno228, PRT, Zeamays>	(128) KLEEMFDIPGELSAS-----LKKWKVIYTDDEDDM	
seqidno84, PRT, Oryzasativa>	(122) TLARMFPTNDHQGEK-----KMANDDHGDAAGPVVITYEDKGDGW	
seqidno102, PRT, Oryzasativa>	(137) ALQDKFFSHFTIIPRERGDEA-RRRGERQVRADVRGQGRRLDARRRRPLE	
seqidno108, PRT, Oryzasativa>	(66) ALQDKFFSHFTIR-----KLGNEEMKLVDVAVSGNEYVPTYEDKDGDW	
seqidno126, PRT, Oryzasativa>	(113) ALHGMFASCLAVR---GGG-GGDGEGTKLVLDLVTGAEYVPTYEDKDGDW	
seqidno180, PRT, Oryzasativa>	(114) ALHGMFASCLAVR---GGA-GGDGEGTKLVLDLVTGAEYVPTYEDKDGDW	
seqidno22, PRT, Arabidopsisthaliana>	(116) ALENMFQGIITICRVTELER-----KGEFVATYEDKDGDL	
seqidno54, PRT, Arabidopsisthaliana>	(104) ALENMFQFTVGEYSE-----REGYKGSFVPTYEDKDGDW	
seqidno120, PRT, Oryzasativa>	(176) ALEKMFSCFTVGHGESNGKSGRDGLSDCRLMDLKNGTTELVLTYEDKDDEW	
seqidno144, PRT, Oryzasativa>	(188) GLEKMFIFGSTGKEGAENQK-----DGEYVLTIEDKDGDW	
seqidno72, PRT, Oryzasativa>	(188) GLEKMFIFGSTGKEGAENQK-----DGEYVLTIEDKDGDW	
seqidno210, PRT, Zeamays>	(168) ALEKMFSGFSTG-----EMSRVTLLERMARQY	
seqidno174, PRT, Oryzasativa>	(192) ALEKMFSCFTIGRSSSHKTSKRDRDLTDGSRADALKDQEVVLTIEDKDDADW	
seqidno198, PRT, Zeamays>	(30) ALQKMFSCPIAGQSGSLHKSSSKDRLTNGSKVDALKDQEVVLTIEDKDDADW	
seqidno138, PRT, Oryzasativa>	(190) ALEKMFSSFTIGNCG-SHGV--NGMNESKIADLLNGSEYVPTYEDKDGDW	
seqidno192, PRT, Oryzasativa>	(190) ALEKMFSSFTIGNCG-SHGV--NGMNESKIADLLNGSEYVPTYEDKDGDW	
seqidno234, PRT, Zeamays>	(186) ALENMFSSFTIGSCG-SQGM--NGMNESKLVDLLNGSEYVPTYEDKDGDW	
seqidno156, PRT, Oryzasativa>	(58) AFQNMFTSFTIGKCG-SHOQ--LKESNK---LRDDLEYVPTYEDKDGDW	
seqidno90, PRT, Oryzasativa>	(152) AFQNMFTSFTIGKCG-SHOQ--LKESNK---LRDDLEYVPTYEDKDGDW	
seqidno162, PRT, Oryzasativa>	(161) ALQKMFGTFTAT---G---NNMN-----EVNGSDAVTIEDKDGDW	
seqidno216, PRT, Zeamays>	(156) ALKKMFSTFTTS---G---NNMNEGKLVDVPVSGADVVTYEDKDGDW	
seqidno36, PRT, Arabidopsisthaliana>	(148) ALSKMFSSFTIGNYGPQGMK--DFMNESKLIDLLNGSDVPTYEDKDGDW	
seqidno48, PRT, Arabidopsisthaliana>	(154) ALAKMFSSFTMGNYGAQGMK--DFMNESKLMNLLNSSEYVPSYEDKDGDW	
seqidno66, PRT, Arabidopsisthaliana>	(246) ALEKMFTTFTLQCGSNGAAGKMLSETKLDLLNGKDVLTIEDKDGDW	
Consensus	(301) ALEKMFSSFT G	D L G EYV TYEDKDGDW
seqidno02, PRT, Oryzasativa>	351	400
seqidno2, PRT, Arabidopsisthaliana>	(139) VLVGDVPWEMFVSTVKRLHVLKTSHASSLSPRKHGKE-----	
seqidno14, PRT, Arabidopsisthaliana>	(139) VLVGDVPWEMFVSTVKRLHVLKTSHAFSLSPRKHGKE-----	
seqidno24, PRT, Arabidopsisthaliana>	(156) RNAGDVPWNEFIESVRLRITRRNDVLPF-----	
seqidno228, PRT, Zeamays>	(162) RNVGDVPWKEFVESVDRMRIARRNDALLPF-----	
seqidno84, PRT, Oryzasativa>	(158) MLVGDDVPWSEFCRMVKRIYIYSYEEAKSLTPKAKLPAIGGDTGVKPDPSK	
seqidno102, PRT, Oryzasativa>	(161) MLVGDPVWDDFARSVKRLKILG-----	
seqidno108, PRT, Oryzasativa>	(186) NVCGDLPTSSSHEKL-----	
seqidno126, PRT, Oryzasativa>	(108) MLVGDPVWKMVFVETCQRLRLMKSSSEAVNLAPRSA-----	
seqidno180, PRT, Oryzasativa>	(158) MLVGDPVWK-----	
seqidno22, PRT, Arabidopsisthaliana>	(159) MLVGDPVWKMVFVESCKRIRLMKSSSEAVNLSPRRSSR-----	
seqidno54, PRT, Arabidopsisthaliana>	(151) MLVGDPVWMMFVESCKRMRLMKTGDAIGL-----	
seqidno120, PRT, Oryzasativa>	(139) MLVGDPVWDMFSSSCQKLRIMKGSEAPTAL-----	
seqidno144, PRT, Oryzasativa>	(226) MLVGDPVWRMFTDSCRRLRIMKGSDAVGLAPRATDKSKNRN-----	
seqidno72, PRT, Oryzasativa>	(223) MLVGDPVWEMFTDSCRRLRIMKGSDAIGLGCSQLRLVPLFVPKL-----	
seqidno210, PRT, Zeamays>	(223) MLVGDPVWEMFTDSCRRLRIMKGSDAIGLAPRAGEKSKNRN-----	
seqidno174, PRT, Oryzasativa>	(195) VIVFHFDVDGVRSTSRSL-----	
seqidno198, PRT, Zeamays>	(242) MLVGDLPWDLPFTTSCRKLRLMRGSDAAGIASDNLNNGNSYLLCPCSSSEIT	
seqidno138, PRT, Oryzasativa>	(80) MLVGDLPWDYGDQMITEG-----	
seqidno192, PRT, Oryzasativa>	(237) MLVGDPVWEMFVESCKRLRIMKGSEAIGLAPRAMEKCKNRS-----	
seqidno234, PRT, Zeamays>	(237) MLVGDPVWEMFVESCKRLRIMKGSEAIGLAPRAMEKCKNRS-----	
seqidno156, PRT, Oryzasativa>	(233) MLVGDPVWEMFVESCKRLRIMKGSEAIGLAPRAMEKCKNRS-----	
seqidno90, PRT, Oryzasativa>	(101) MLVGDPVWEMFVESCKRLRIMKGSEAIGLAPRAVEKCKS-----	
seqidno162, PRT, Oryzasativa>	(195) MLVGDPVWEMFVESCKRLRIMKGSEAIGLAPRAVEKCKS-----	
seqidno216, PRT, Zeamays>	(196) MLVGDPVWQMFVESCKRLRIMKGSEAIGLAPRAKDKYKNKS-----	
seqidno36, PRT, Arabidopsisthaliana>	(197) MLVGDPVWEMFVESCKRLRIMKSSSEIGLAPRTKDKCKNRS-----	
seqidno48, PRT, Arabidopsisthaliana>	(196) MLVGDPVWEMFVDSCKRIRIMKGSEAIGLAPRALEKCKNRS-----	
seqidno66, PRT, Arabidopsisthaliana>	(202) MLVGDPVWBE-----	
Consensus	(296) MLVGDPVWEMFIDVCKKLKIMKGCDAGLAAAPRAMEKSKMRA-----	
	(351) MLVGDPVWEMFVESCKRLRIMK SEAIGLAPR	

FIGURE 11 (continued)

	401	430
seqidno02, PRT, Oryzasativa>	(176)	-----
seqidno2, PRT, Arabidopsisthaliana>	(176)	-----
seqidno14, PRT, Arabidopsisthaliana>	(186)	-----
seqidno24, PRT, Arabidopsisthaliana>	(192)	-----
seqidno228, PRT, Zeamays>	(208)	LPPESDVPQSDSDNSAPVAADKD-----
seqidno84, PRT, Oryzasativa>	(183)	-----
seqidno102, PRT, Oryzasativa>	(201)	-----
seqidno108, PRT, Oryzasativa>	(142)	-----
seqidno126, PRT, Oryzasativa>	(167)	-----
seqidno180, PRT, Oryzasativa>	(195)	-----
seqidno22, PRT, Arabidopsisthaliana>	(180)	-----
seqidno54, PRT, Arabidopsisthaliana>	(169)	-----
seqidno120, PRT, Oryzasativa>	(267)	-----
seqidno144, PRT, Oryzasativa>	(267)	-----
seqidno72, PRT, Oryzasativa>	(264)	-----
seqidno210, PRT, Zeamays>	(213)	-----
seqidno174, PRT, Oryzasativa>	(292)	GTDRSEQIVARAFIIWTANVWTKFAVASFR
seqidno198, PRT, Zeamays>	(98)	-----
seqidno138, PRT, Oryzasativa>	(278)	-----
seqidno192, PRT, Oryzasativa>	(278)	-----
seqidno234, PRT, Zeamays>	(274)	-----
seqidno156, PRT, Oryzasativa>	(140)	-----
seqidno90, PRT, Oryzasativa>	(234)	-----
seqidno162, PRT, Oryzasativa>	(237)	-----
seqidno216, PRT, Zeamays>	(238)	-----
seqidno36, PRT, Arabidopsisthaliana>	(237)	-----
seqidno48, PRT, Arabidopsisthaliana>	(211)	-----
seqidno66, PRT, Arabidopsisthaliana>	(339)	-----
Consensus		

FIGURE 11 (continued)

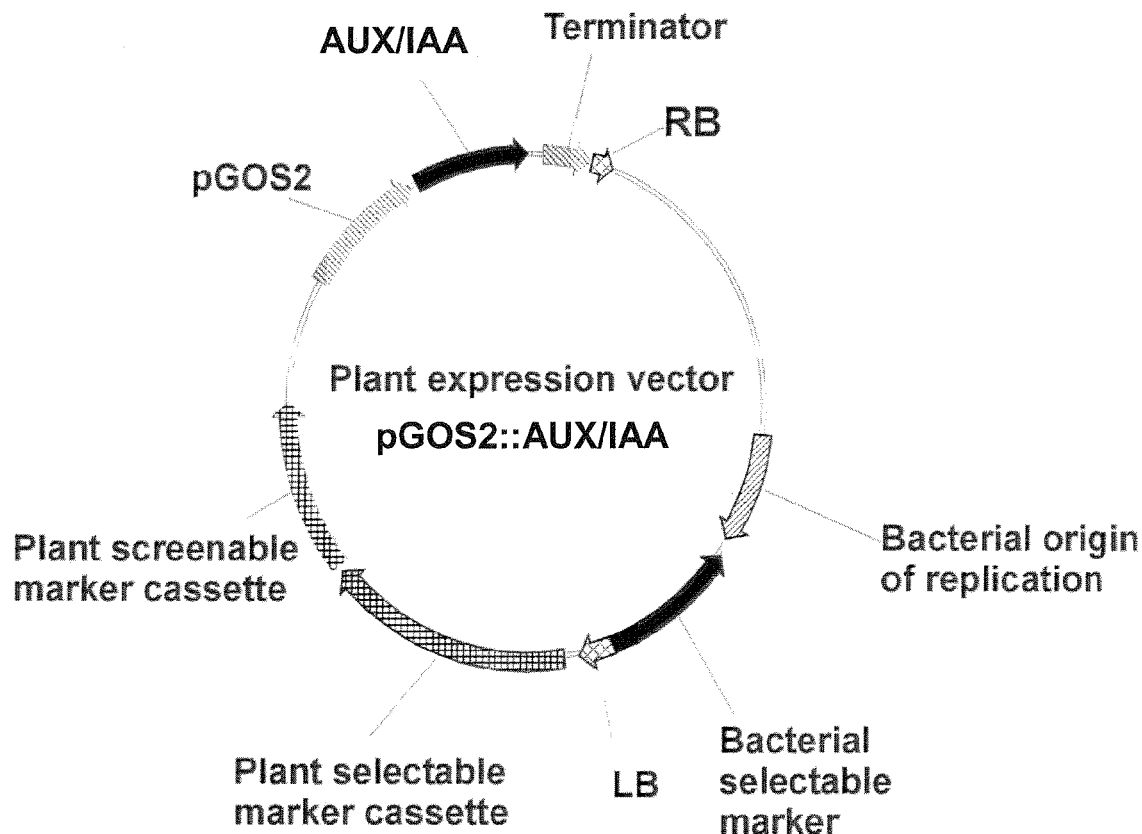


FIGURE 12

MNLKETELCLGLPGGTETVESPAKSGVGNKRGFSETVDL

Motif 1

Motif 2

KLNLQSNKQGHVDLNTNGAPKEKTFLKDPSKPPAKAQVV

GWPPVRNYRKNVMANQKSGEAEFEAMSSGGGTVAFVKVSM

Motif 3

DGAPYLRKVDLKMYSYKDLSDALAKMFSSFTMGSYGAQ

Motif 4

GMIDFMNESKVMDLLNSSEYVPSYEDKDGDWMLVGDVPW

Motif 5

PMFVESCKRLRIMKGSEAIGLAPRAMEKFKNRS

Motif 6

bold: AUX-IAA domain

FIGURE 13

CLUSTAL 2.0.9 multiple sequence alignment

```
AT3G23050.1  -----
AT3G23050.2  -----
AT4G14550.1  -----
Mt_TA20354   -----
Pt_566151    -----
Pt_720961    -----
Sl_TA40922   -----
AT1G04250.1  -----
Mt_TA27011   -----
Mt_TA22814   -----
Pt_643213    -----
Sl_TA48108   -----
Os_CB657009  -----
Os_TA41733   -----
AT3G04730.1  -----
Mt_TA20951   -----
Mt_TA25400   -----
Pt_584053    -----MSMPLEHDYIGISSEVSSMENTSG-----
Pt_711734    -----MSSIPKEHDYIGLS-ETPSMEKISDKLSSSSSTL-----
AT4G29080.1  -----MSVSVA AEHDYIGLS-EFPTMEATTMS-----
Mt_TA23062   MSLPRLGIGDEESKSNVTLLLEKSLHLNGSKPKFNYMGLPSSNCSSVDSSVP-----
AT3G23030.1  -----
AT4G14560.1  -----
Sl_TA38817   -----
Sl_TA43058   -----
Pt_726443    -----
Pt_564913    -----
Mt_TA20557   -----
Pt_831610    -----
Pt_798526    -----
Mt_TA31746   -----
Pt_823671    -----
Pt_595419    -----
Mt_TA20558   -----
AT1G04240.1  -----
Sl_TA42190   -----
```

FIGURE 14

AT3G23050.1	-----MIGQLMNLKATELCLGLPGG-----
AT3G23050.2	-----MIGQLMNLKATELCLGLPGG-----
AT4G14550.1	-----MNLKETELCLGLPGG-----
Mt_TA20354	-----MATMGHGLNLKETELCLGLPGGGGGGGG-G-----
Pt_566151	-----MATATVLGTEMADLNYKETELCLGLPGAVG-----V-----
Pt_720961	-----MTTSVLGTERDNLNYKETELCLGLPGAVG-----A-----
Sl_TA40922	-----MDLKETELCLGLPGGGGGGGGELI-----
AT1G04250.1	-----MMGSVELNLRKETELCLGLPGG-----
Mt_TA27011	-----MEVVG---MKKE-NMGFEETELRLGIGFLGN-----
Mt_TA22814	-----MEVVAG---MKKEEKMVFDETELRLGLGLPG-----
Pt_643213	-----MEVEK-----GTKMGFEETELRLGLPGNGG-----
Sl_TA48108	-----MSSNK-----LDFEETELRLGL---PGG-----
Os_CB657009	-----
Os_TA41733	-----MAADLAFEATELRLGLPGGGG-----
AT3G04730.1	-----MINFEATELRLGLPGGNH-----
Mt_TA20951	-----MTNVGDAERDKYSLINFEETELRLGLPGAGD-----
Mt_TA25400	-----
Pt_584053	-----TDTINISTTASKGLNLKATELRLGLPGSDSPERGNE-----NQQLGFS
Pt_711734	STEENINSNSNSNSNSTNTSLNLKETELRLGLPGYQSPERKLT-----LPAAGVS
AT4G29080.1	-----DKTKTRDNNNGLNFKATELRLGLPGSESPER-----VDSRFLA
Mt_TA23062	-----KIQSPKDETKSNLNLKATELRLGLPGSLSPERDSSDFCLRSSKQFDEKPLFP
AT3G23030.1	-----MAYEKVNELN-LKDTELCLGLPGRTEKI-----
AT4G14560.1	-----MEVTNGLN-LKDTELRLGLPGAQE-----
Sl_TA38817	-----MECVLAHEKDLN-LKATELRLGLPGRTDE-----
Sl_TA43058	-----MTSIVGNQKDLN-FKATELRLGLPGTEDQ-----
Pt_726443	-----MEGGVAYENDLN-LKATELRLGLPG-TSC-----
Pt_564913	-----MEGGVAYENDLN-LKATELRLGLPG-TGC-----
Mt_TA20557	-----MENKVAYENDLN-MKATELRLGLPG-TEQ-----
Pt_831610	-----MA-YESDLN-LKATELRLGLPGSDEP-----
Pt_798526	-----MERSMA-YERHLN-LKATELRLGLPGSDEP-----
Mt_TA31746	-----MENSLGNHQTEMN-LKATELRLGLPGSDEV-----
Pt_823671	-----MEFERDLN-LDATELRLGLPGTATKQS-----
Pt_595419	-----MEFERDLN-LEATELRLGLPGTATEQL-----
Mt_TA20558	-----MEF-----KATELRLGLPGTDEK-----
AT1G04240.1	-----MDEFVN-LKETELRLGLPGTDNVC-----
Sl_TA42190	-----MRIYEKDINDLEATELRLGLPGIIND-----

FIGURE 14 (continued)

AT3G23050.1	-----AEAVESPAKSAVGSKRGFSET-----VDLMLNLQSNKEGS-----
AT3G23050.2	-----AEAVESPAKSAVGSKRGFSET-----VDLMLNLQSNKEGS-----
AT4G14550.1	-----TETVESPAKSGVGSKRGFSET-----VDLKLNLQSNKQGH-----
Mt_TA20354	-----GSEVETPRAS--GKRGFSET-----VDLKLNLQTK-----
Pt_566151	-----KNEVETPNKAT--GKRGFAET-----VDLKLNLQAKEGVMDL---
Pt_720961	-----KNEVETPNKAT--GKRGFAET-----VDLKLNLQAKEGVMDL---
Sl_TA40922	-----RDNNNNNNKVN--GKRGFSET-----VDLKLNFHQASDDISC---
AT1G04250.1	-----DTVAPVTGN--KRGFSET-----VDLKLNLNNEPANK-----
Mt_TA27011	-----NG--SATATEGVVRKRGFSETETDDDTTMDLMLNLSSKEATA-----
Mt_TA22814	-----KTTEVVRKRGFSETSESESETNTVDLKLNLSTKEG-----
Pt_643213	-----G-----AEGEMVRKRGFSET-----VDLKLKLSKES-----
Sl_TA48108	-----ARKNVYGNDTCNVN--GKRGFVDLKLNLSSDIN-----
Os_CB657009	-----
Os_TA41733	-----DGDAAAAAARSSSGKRGFAET-----IDLKLEPAAAAVDDD---
AT3G04730.1	-----GGEMAG---KNNGKRGFSET-----VDLKLNL-----
Mt_TA20951	-----HGESPVK--NSCGKRGFSETA-----NVDLKLNLSPIN-----
Mt_TA25400	-----
Pt_584053	LNNNNSKD-----KSFVSGARRGFSVAIHGGSANWVFSGNAGSDPNF-----
Pt_711734	LFGKDIDTNNNTNGYPLRPLKLVSGTKRGFSDAIVGSSGKWVFSGNGSEVDLGKGAILE
AT4G29080.1	LNKS-----SCPVSGAKRVFSDAIN-DSNKWVFS--PGSTTATG-----
Mt_TA23062	LHPQKDDHLFES-----KPAVLGNKRGFSDAMNVFSEGKLPSSKMLENVAG-----
AT3G23030.1	-----KEEQEVSCVKSNNKRLFEETR-----
AT4G14560.1	-----EQQLELSCVRSNNKRKNDS-----
Sl_TA38817	-----ESDKEIVFHFKNKRALPE-----DEDC-----
Sl_TA43058	-----ESDQEISNSKNNKRALPESTHD-----EEDC-----
Pt_726443	-----TNEEQAVSGARNNRPLPET-----REER-----
Pt_564913	-----TNE-KGVSGARNNRPPPET-----REEG-----
Mt_TA20557	-----NEE---QAKISNKRLPTET-----SKDS-----
Pt_831610	-----EK-PSTTPSVRSNKRASPEISE-----ESRSKG-----
Pt_798526	-----EK-PSTTPSVRSNKRASPEISE-----ESRSKG-----
Mt_TA31746	-----EKLPCNFSLRNNKRSSPEEASDV-----DSISKSKLN-----
Pt_823671	-----EKQTPNSNLAKSNKRSLPDMNE-----EPAGSSREN-----
Pt_595419	-----EKQTPNSNVTKSNKRSLPDMNE-----DSAG--RRE-----
Mt_TA20558	-----DMKTIHGVSVKNNKRQLPQTSE-----ESVS-----
AT1G04240.1	-----EAKERVSCCNNNKRVLSTDTEK-----EIES-----
Sl_TA42190	-----ESSTSTSTSKNSRKRPSSSSVN-----

FIGURE 14 (continued)

AT3G23050.1	-----VDLKNVSAVPKEKTTL-KDPSKPPA-----
AT3G23050.2	-----VDLKNVSAVPKEKTTL-KDPSKPPA-----
AT4G14550.1	-----VDLN-TNGAPKEKTFL-KDPSKPPA-----
Mt_TA20354	-----DLNEKSAS-KEKTLL-KDPAKPPA-----
Pt_566151	-----NENIKNIASKDKNHLPADTI-KDPAKPPA-----
Pt_720961	-----NENIKNITSKDKNHLPAVTI-KDPAKPPA-----
Sl_TA40922	-----AMENNMKSSVTTTKEVVCN-KDPIKPPA-----
AT1G04250.1	-----EGSTTHDVVTFDSKEKSACPKDPAKPPA-----
Mt_TA27011	-----EVDPSDITTKTLQKEKTLLPADP-AKPPA-----
Mt_TA22814	-----ATDP----QFKPKEKALLLSDSGAKPPA-----
Pt_643213	-----GADPNHEKTSSLQREKNLLATDP-AKPPA-----
Sl_TA48108	-----NIKNSTHKTPAA-----
Os_CB657009	-----
Os_TA41733	-----DDKEEAAADDREKKVDIVGADNDDASPPAAAAAGGMKR
AT3G04730.1	-----STAMDSVSKVDLENMKEKVVKPPA-----
Mt_TA20951	-----DSASSSSTIASVAENK GKDTTTSATVSP-----
Mt_TA25400	-----
Pt_584053	SLRGANS-----KEGFPHSSKPVVQENKSQVDGANTNGHGAAPAS-----
Pt_711734	SPRGDNGNSQKSCVAGPAKKDDVAQSPKP-VQEKISQVAAANEN--SSAPAA-----
AT4G29080.1	-DVGSGSGP-----RTSVVKDGKS-TTFTKPAVPVKEKKSSATAPAS-----
Mt_TA23062	QKVKADEIA-----TVKIGLERPNGVGESKPGLNGSANNGNSTAPAS-----
AT3G23030.1	-----DEEESTPP-----
AT4G14560.1	-----TEESAPPP-----
Sl_TA38817	-----ESN-----SISD-PKTPP-----
Sl_TA43058	-----ESK-----SSSDHVKTPPP-----
Pt_726443	-----GAKGKSDPRHDDQETAPAP-----
Pt_564913	-----GANGKSDAQHDDQETASAPNTYSFDMHA-----
Mt_TA20557	-----GSK-----TSDD--AAPPS-----
Pt_831610	-----SSSVSSNVEN-GERDSAPP-----
Pt_798526	-----SSSLSSNVEN-SEGDDAPP-----
Mt_TA31746	-----SSNGSSHTTN-DDQDNAPP-----
Pt_823671	-----SSTVSSNDKKSHDQETAPP-----
Pt_595419	-----SSSVSSNDKKSHDQETAPP-----
Mt_TA20558	-----ISKVSNDQHVESSAAPP-----
AT1G04240.1	-----SSRKTETSPP-----
Sl_TA42190	-----ENEQQDSAPAP-----

FIGURE 14 (continued)

AT3G23050.1	-----KAQVVGWPPVRNYRKNMMTQOKTSSG-----
AT3G23050.2	-----KAQVVGWPPVRNYRKNMMTQOKTSSG-----
AT4G14550.1	-----KAQVVGWPPVRNYRKNVMAQKSGE-----
Mt_TA20354	-----KAQVVGWPPVRSYRKNMMAQKVNTE-----
Pt_566151	-----KAQVVGWPPVRSYRKNVLAQKNASEEGFRAQVVGWPPPLRS
Pt_720961	-----KAQVVGWPPVRSYRKNVMAQKNASEE-----
Sl_TA40922	-----KAQVVGWPPVRSFRKNVMAQKSNTEE-----
AT1G04250.1	-----KAQVVGWPPVRSYRKNVMVSCQKSSG-----
Mt_TA27011	-----KAQVVGWPPVRSYRKNMLAMQKS-----
Mt_TA22814	-----KAQVVGWPPVRSFRKNMFAAQKSNEGS-----
Pt_643213	-----KAQVVGWPPVRSFRKNMLAVQKSS-TD-----
Sl_TA48108	-----KAQVVGWPPVRSFRKNILTSQKLD-----
Os_CB657009	-----
Os_TA41733	SPSQSSVVTAAADPEKPRAPKAQVVGWPPVRSYRKNILAVQADKGKD-----
AT3G04730.1	-----KAQVVGWPPVRSFRKNVMSGQKPTTGD-----
Mt_TA20951	-----PPRAKAQVVGWPPVRSFRKNIVNVHQ-KSNS-----
Mt_TA25400	-----MMKLREN-----
Pt_584053	-----KAQVVGWPPIRSFRKNTMASHLS-----
Pt_711734	-----KAQVVGWPPIRSFRKNTMASSLV-----
AT4G29080.1	-----KAQVVGWPPIRSFRKNMMASSQSQKPG-----
Mt_TA23062	-----KAQVVGWPPIRSFRKNSLTTAS-----
AT3G23030.1	-----TKTQIVGWPPVRSRKNNNNS-----
AT4G14560.1	-----AKTQIVGWPPVRSNRKNNNNK-----
Sl_TA38817	-----VAKTQIVGWPPVRANRKNFSFPSKK-----
Sl_TA43058	-----VAKAQIVGWPPVRSNRKNIIQPKK-----
Pt_726443	-----KAQIVGWPPIRSFRKNLQPKKA-----
Pt_564913	-----TCRVQIVGWPPIRSFRKNLQPKKA-----
Mt_TA20557	-----KAKIVGWPPIRSFRKNLQPKKA-----
Pt_831610	-----AKAQVVGWPPIRSFRKNLQPKKN-----
Pt_798526	-----AKAQVVGWPPIRSFRKNLQPKKN-----
Mt_TA31746	-----SKAQVVGWPPIRSFRKNLQPKKG-----
Pt_823671	-----IKAQVVGWPPIRSFRKNLQAKK-----
Pt_595419	-----TKTQVVGWPPIRSFRKNLQARK-----
Mt_TA20558	-----AKAKIVGWPPIRSFRKNLQPKK-----
AT1G04240.1	-----RKAQIVGWPPVRSYRKNNIQSKKNE-----
Sl_TA42190	-----KAQVVGWPPVRSYRKNHVSQKLS-----

FIGURE 14 (continued)

AT3G23050.1	-----AEEASSEKAGNFGGGAAGAGLVKVSMDGAPYLRKVDLKMYKSYQDLSDALAKMF
AT3G23050.2	-----AEEASSEKAGNFGGGAAGAGLVKVSMDGAPYLRKVDLKMYKSYQDLSDALAKMF
AT4G14550.1	-----ABEAMSS-----GGGTV--AFVKVSMDGAPYLRKVDLKMYTSYKDLSDALAKMF
Mt_TA20354	-----DTEKTTS-----NTTAAAFVKVSMDGAPYLRKVDLTMYKTYKDLSDALAKMF
Pt_566151	YRK NVLTQKNASEEGDKASTGGSSAAFVKVCMDGAPYLRKVDLKMYKSYQELSDALAKMF
Pt_720961	-----GEKASTGGSSAAFVKVCMDGAPYLRKVDLKMYRSYQELSDALAKMF
Sl_TA40922	-----SEKTTAAAFVKVCMDGAPYLRKVDLKMYKSYQELSDALAKMF
AT1G04250.1	-----GPEAAAFVKVSMDGAPYLRKIDLRMYKSYDELSNALSNMF
Mt_TA27011	--ESEKNSSSNFN-----AITFVKVSMDGAPYLRKVDLKMYTSYSQSLGSKMF
Mt_TA22814	-BESEKKNS-NNN-----PISFVKVSMDGAPYLRKVDLKMYKSYPELSDALAKMF
Pt_643213	-QECEKVPG---G-----NATFVKVSMDGAPYLRKVDLKMYKTYQELSDALGKMF
Sl_TA48108	---RENDN-----ILVKVSMDGAPYLRKVDLNMYKSYQELFDALTKMF
Os_CB657009	-----
Os_TA41733	-AADGGGDKSGAG-----AAAAAFVKVSMDGAPYLRKVDLKMYKSYLELSKALEKMF
AT3G04730.1	-ATEGNDKTSGSSGATSSASACATVAVVKVSMDGAPYLRKIDLKLYKTYQDLSNALSKMF
Mt_TA20951	-ETEVDKSISSGG-----GNGAFVKVSMDGAPYLRKVDLKLYKSYQELSDALAKMF
Mt_TA25400	QNFD-----CLYVKVSMDGAPYLRKVDLKTYNNYMESSALEKMF
Pt_584053	KNDDGAEVKSGSG-----CLYVKVSMDGAPYLRKVDLKTFGSYMESSALEKMF
Pt_711734	KNNEDVEGKSGYG-----CLYVKVSMDGAPYLRKVDLKTYSNYLESSALEKMF
AT4G29080.1	NNSETEEAEAKSGPE-----QPCLYVKVSMGAPYLRKIDLKTYKSYLELSALEKMF
Mt_TA23062	KNTTEVDGKLGSGG-----AVFVKVSMDGAPYLRKVDLKNTYAYSELSSSLEKMF
AT3G23030.1	-----VSYVKVSMDGAPYLRKIDLKTYKNYPPELLKALENMF
AT4G14560.1	-----NVSYVKVSMDGAPYLRKIDLKMYKNYPPELLKALENMF
Sl_TA38817	-----AEAECGMYVKVSMDGAPYLRKIDLKLYKGYPPELLKALEKMF
Sl_TA43058	-----TESESGMYVKVSMDGAPYLRKIDLKMYKCYQELLKALENMF
Pt_726443	-----EAEAAAAGMYVKVSMDGAPYLRKIDLKVYKGYPPELLKALENMF
Pt_564913	-----EDEAAAAGMYVKVSMDGAPYLRKIDLKVYKGYPPELLKALENMF
Mt_TA20557	-----EAEASGIYVKVSLDGAPYLRKIDLRVYGGYAQLLKALESMF
Pt_831610	-----DQVDGAGMYVKVSVDGAPYLRKIDLKVYKSYPELLKALENMF
Pt_798526	-----DRVDGAGMYVKVSVDGAPYLRKIDLKVYRSYPELLKALED MF
Mt_TA31746	-----EEVG---MYLKVSMAGAPYLRKIDLKVYKSYSELLKVLENMF
Pt_823671	-----LEAEAAAGLYVKVSMDGAPYLRKIDLKVYKGYPPELLKALEEMF
Pt_595419	-----LEAEAAAGLYVKVSMDGAPYLRKIDLKVYKGYPPELLLEVVEEMF
Mt_TA20558	-----EAEVGGIYVKVSMDGAPYLRKIDLR IYGGYPPELLKALETMF
AT1G04240.1	-----SEHEGQGIYVKVSMDGAPYLRKIDLS CYKGYSELLKALEVMF
Sl_TA42190	-----SDNNSSGMYLKVSMGAPYLRKIDLQVYKSYQELLKALQSMF

FIGURE 14 (continued)

AT3G23050.1 SSFTMGNYGAQ--GMIDFMNES-KLMNLLNSSEYVPSYEDKDGDWMLVGDVPWEMFVESC
AT3G23050.2 SSFTMGNYGAQ--GMIDFMNES-KLMNLLNSSEYVPSYEDKDGDWMLVGDVPWE-----
AT4G14550.1 SSFTMGSYGAQ--GMIDFMNES-KVMDLLNSSEYVPSYEDKDGDWMLVGDVPWPMFVESC
Mt_TA20354 SSFTTGNYGAQ--GMIDFMNES-KLMDLLNSSEYVPTYEDKDGDWMLVGDVPWEMFVGSC
Pt_566151 SSFTMGNYGAQ--GMIDFMNES-KLMDLLNSSEYVPSYEDKDGDWMLVGDVPWEMFVDSC
Pt_720961 SSFTMGNYGAQ--GMIDFMNES-KLMDLLNSSEYVPSYEDKDGDWMLVGDVPWEMFVNSC
Sl_TA40922 SSFTNGNYGSQ--GMIDFMNES-KLMDLLNSSEYVPTYEDKDGDWMLVGDVPREMFGDSC
AT1G04250.1 SSFTMGKHGEGE-GMIDFMNER-KLMDLVNSWDYVPSYEDKDGDWMLVGDVPWPMFVDTC
Mt_TA27011 SSFTIGNCESQ--GMKDFMNES-KLMDLLNSDYVPTYEDKDGDWMLVGDVPWEMFVESC
Mt_TA22814 NSFTTGNCSQ--GIKDFMNESNKLMDLLNSDYVPTYEDKDGDWMLVGDVPWDMFIDSC
Pt_643213 SSFTIGNCGSH--GLKDFLNES-KLIDLLNGTDYVPTYEDKDGDWMLVGDVPWDMFVESC
Sl_TA48108 NSFTI----VQ--GMKDFMHEG-KLMDLLNSDYVPTYEDKDGDWMLVGDVPWGMFVDSC
Os_CB657009 -----MNES-KIADLLNGSEYVPTYEDKDGDWMFVGDVPWEMFVESC
Os_TA41733 SSFTIGNCGSH--GVNG-MNES-KIADLLNGSEYVPTYEDKDGDWMLVGDVPWEMFVESC
AT3G04730.1 SSFTIGNYGPQ--GMKDFMNES-KLIDLLNGSDYVPTYEDKDGDWMLVGDVPWEMFVDSC
Mt_TA20951 SSFTIDNCGSQ--VTKDFMNES-KLIDLLNGSDYVPTYEDKDGDWMLVGDVPWEMFVQSC
Mt_TA25400 TCFTIGQCNSPGLPGKDGLSES-SLRDLLHGSEYVLTIEDKDGDWMLVGDVPWGMFADSC
Pt_584053 SCFTIGQCGSHVPGQDGLSES-RLMDLLHGSEYVLTIEDKDNDWMLVGDVPWKMFTDSC
Pt_711734 SCFTIGQCGSHGLRGQDGLTES-RLKDILHGSEYVLTIEDKDGDWMLVGDVPWDMFTNSC
AT4G29080.1 SCFTIGQFGSHGGCGRDGLNES-RLTDLLRGSEYVVTIEDKDSWMLVGDVPWEMFICSC
Mt_TA23062 SCFTIGQCESH---GNQMLNET-KLRDLLHGSEYVITYEDKDGDWMLVGDVPWEMFIDTC
AT3G23030.1 K-VMIGEYCE-----EG-----YKSGGFVPTYEDKDGDWMLVGDVPWDMFSSSC
AT4G14560.1 K-FTVGEYSE-----EG-----YKSGGFVPTYEDKDGDWMLVGDVPWDMFSSSC
Sl_TA38817 K-LSIGEYSE-----EG-----YKGSEFAPAYEDKDGDMLVGDVPFEMFLSSC
Sl_TA43058 K-LTIGEYSE-----EG-----YKGSEFAPAYEDKDGDMLVGDVPWEMFMSSC
Pt_726443 K-LTIGEYSE-----EG-----YKGSEYAPTYEDKDGDWMLIGDVPWDMFLSSC
Pt_564913 K-LTIGEYSE-----EG-----YKGSEYAPTYEDKDGDWMLVGDVPWDMFLSSC
Mt_TA20557 K-LTIGNYSEK---EG-----YKGSEYEPTYEDKDGDWMLVGDVPWEMFVTSC
Pt_831610 K-LTIGEYSEN---EG-----YNGSEFAPTYEDKDGDWMLVGDVPWDMFISSC
Pt_798526 K-LTIGEYSEK---EG-----YNGSDFAPTYEDKDGDWMLVGDVPWDMFISTC
Mt_TA31746 K-CTIGEYSE-----EG-----YNGSEFVPTYEDKDGDWMLVGDVPWEMFMSSC
Pt_823671 K-SKVGEYSE-----EG-----YNGSEHVPTYEDKDGDWMLVGDVPWDMFINSC
Pt_595419 K-FKVGEYSE-----EG-----YNGSEYVPTYEDKDGDWMLVGDVPWEMFINSC
Mt_TA20558 K-LTIGEYSE-----EG-----YKGSEYAPTYEDKDGDWMLVGDVPWDMFVTSC
AT1G04240.1 K-FSVGEYFER---DG-----YKGSDFVPTYEDKDGDWMLIGDVPWEMFICTC
Sl_TA42190 K-CTIGVYSE-----EG-----YNGSDYAPTYEDKDGDWMLVGDVPWEMFISSC

.. . :*****.* *::*****

FIGURE 14 (continued)

AT3G23050.1	KRLRIMKGSEAVGLAPRAMEKYCKNRS-----
AT3G23050.2	-----
AT4G14550.1	KRLRIMKGSEAIGLAPRAMEKFKNRS-----
Mt_TA20354	KRLRIMKGSEAIGLAPRAMEKCKNRS-----
Pt_566151	KRLRIMKGSEAIGLAPRAMEKCKSRT-----
Pt_720961	KRLRIMKGSEAIGLAPRAMEKCKSRT-----
Sl_TA40922	KRLRIMKGSEAIGLAPRAMEKCKSRI-----
AT1G04250.1	KRLRLMKGSDAIGLAPRAMEKCKSRA-----
Mt_TA27011	KRLRIMKGKEAIGYSTKSYGKMQEQLDLLVALVRHLLHLLSYFGTCRMFSIVNLCNVIW
Mt_TA22814	KRLRIMKGKEAIGLAPRAMEKCKNRS-----
Pt_643213	KRLRIMKGTEATGLAPRAMEKCKNRSYK-----
Sl_TA48108	KRLRIMKGTEAIGLAPRAMEKCKNRNG-----
Os_CB657009	KRLRIMKGSEAIGLAPRAMEKCKNRS-----
Os_TA41733	KRLRIMKGSEAIGLAPRAMEKCKNRS-----
AT3G04730.1	KRLRIMKGSEAIGLAPRALEKCKNRS-----
Mt_TA20951	KRLRIMKGSEAIGLAPRAVEKCKNRS-----
Mt_TA25400	RRLRIMKGSDAIGLAPRAMEKCKSRQN-----
Pt_584053	RRLRIMKGSEAIGLAPRAMEKCKSRN-----
Pt_711734	RRLRIMKGSEAIGLAPRAMEKCKNRN-----
AT4G29080.1	KKLRLIMKSSEAIGLAPRVMEKCKSRN-----
Mt_TA23062	RRLRIMKSSDAIGLAPRAVEKCKSRN-----
AT3G23030.1	KRLRIMKGSDAPALDSSL-----
AT4G14560.1	QKLRLIMKGSEAP---TAL-----
Sl_TA38817	KRLRIMKGSEARGLGCGV-----
Sl_TA43058	KRLRIMKGSETRGLGCGV-----
Pt_726443	KKLRIIKGSEATG-----
Pt_564913	KKLRLIMKGSEAIGLGCGA-----
Mt_TA20557	KRLRIMKGTEARGV-----
Pt_831610	KRLRIMKGSEARGLGC-----
Pt_798526	KRLRIMKGSEARGLGC-----
Mt_TA31746	KRLRIMKGSEAKGLGCF-----
Pt_823671	KRLRIMKESEARGLGCAV-----
Pt_595419	KRLRIMKESEARGLGCAV-----
Mt_TA20558	KRLRIMKGTEARGLGCGV-----
AT1G04240.1	KRLRIMKGSEAKGLGCGV-----
Sl_TA42190	KRLRIIKGSEAKGLACL-----

FIGURE 14 (continued)

AT3G23050.1	-----
AT3G23050.2	-----
AT4G14550.1	-----
Mt_TA20354	-----
Pt_566151	-----
Pt_720961	-----
Sl_TA40922	-----
AT1G04250.1	-----
Mt_TA27011	FLFFDKIVWFVIHI
Mt_TA22814	-----
Pt_643213	-----
Sl_TA48108	-----
Os_CB657009	-----
Os_TA41733	-----
AT3G04730.1	-----
Mt_TA20951	-----
Mt_TA25400	-----
Pt_584053	-----
Pt_711734	-----
AT4G29080.1	-----
Mt_TA23062	-----
AT3G23030.1	-----
AT4G14560.1	-----
Sl_TA38817	-----
Sl_TA43058	-----
Pt_726443	-----
Pt_564913	-----
Mt_TA20557	-----
Pt_831610	-----
Pt_798526	-----
Mt_TA31746	-----
Pt_823671	-----
Pt_595419	-----
Mt_TA20558	-----
AT1G04240.1	-----
Sl_TA42190	-----

FIGURE 14 (continued)

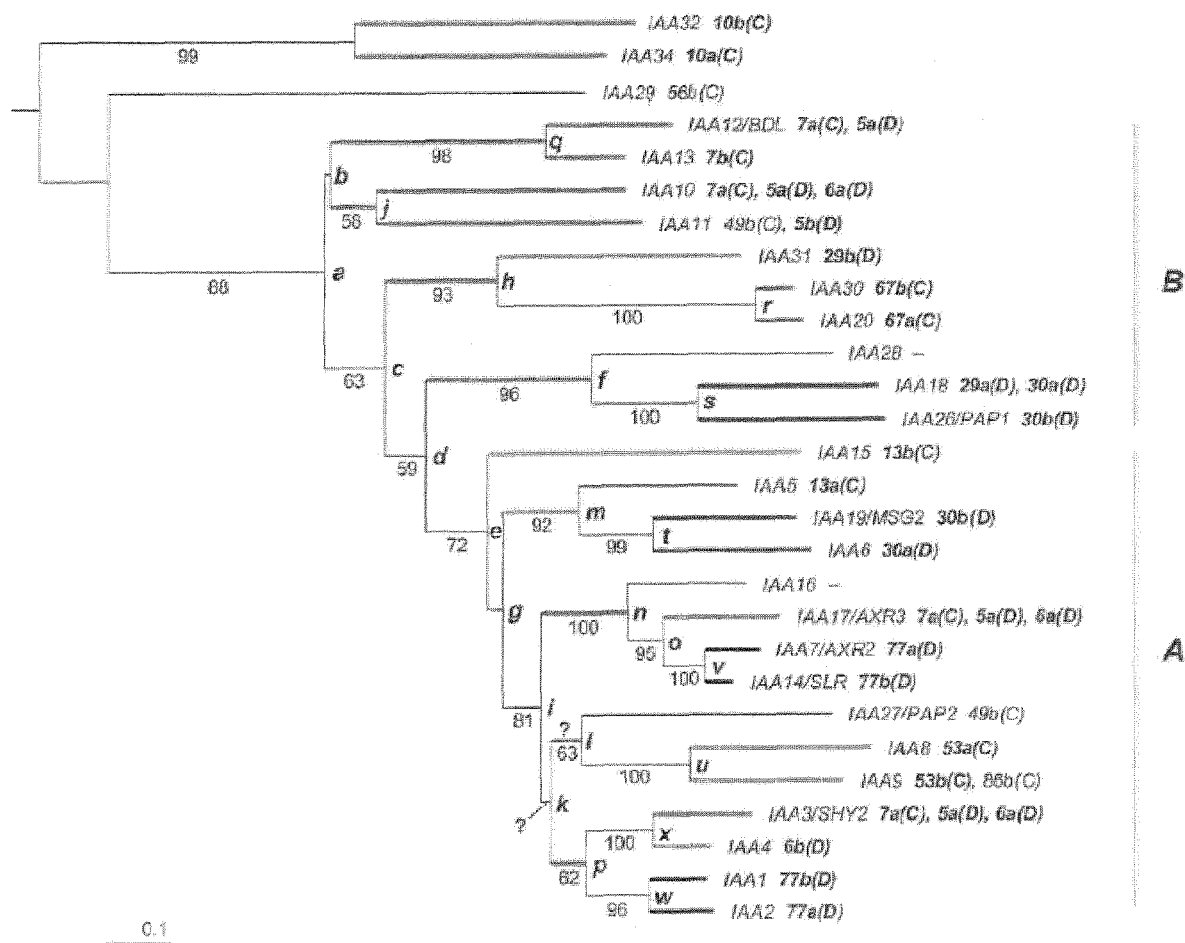
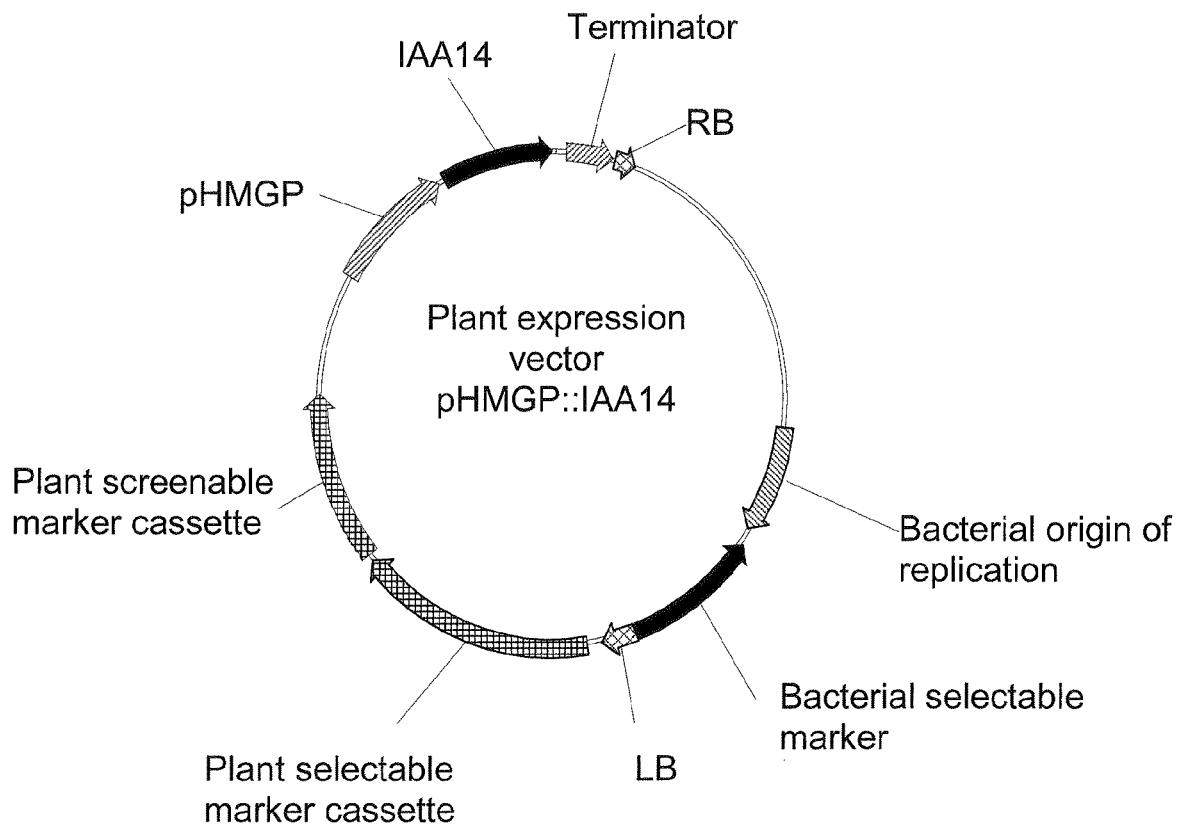


FIGURE 15

**FIGURE 16**

PLANTS HAVING ENHANCED YIELD-RELATED TRAITS AND A METHOD FOR MAKING THE SAME

RELATED APPLICATIONS

This application is a national stage application (under 35 U.S.C. §371) of PCT/EP2009/062174, filed Sep. 21, 2009, which claims benefit of European application 08165001.2, filed Sep. 24, 2008; U.S. Provisional Application 61/099,629, filed Sep. 24, 2008; U.S. Provisional Application 61/103,301, filed Oct. 7, 2008; European Application 08166008.6, filed Oct. 7, 2008; European Application 08167387.3, filed Oct. 23, 2008; European Application 08167390.7, filed Oct. 23, 2008; U.S. Provisional Application 61/107,680, filed Oct. 23, 2008; U.S. Provisional Application 61/107,695, filed Oct. 23, 2008; European Application 09100261.8, filed Apr. 29, 2009; and U.S. Provisional Application 61/180,953, filed May 26, 2009.

SUBMISSION OF SEQUENCE LISTING

The Sequence Listing associated with this application is filed in electronic format via EFS-Web and hereby incorporated by reference into the specification in its entirety. The name of the text file containing the Sequence Listing is Sequence_Listing_13987_00144. The size of the text file is 1,438 KB, and the text file was created on Mar. 19, 2013.

The present invention relates generally to the field of molecular biology and concerns a method for enhancing various economically important yield-related traits in plants. More specifically, the present invention concerns a method for enhancing yield-related traits in plants by modulating expression in a plant of a nucleic acid encoding an ASPAT (Asparatate AminoTransferase) polypeptide. The present invention also concerns plants having modulated expression of a nucleic acid encoding an ASPAT polypeptide, which plants have enhanced yield-related traits relative to control plants. The invention also provides hitherto unknown ASPAT-encoding nucleic acids and constructs comprising the same, useful in performing the methods of the invention.

Furthermore, the present invention relates generally to the field of molecular biology and concerns a method for increasing various plant yield-related traits by increasing expression in a plant of a nucleic acid sequence encoding a MYB91 like transcription factor (MYB91) polypeptide. The present invention also concerns plants having increased expression of a nucleic acid sequence encoding an MYB91 polypeptide, which plants have increased yield-related traits relative to control plants. The invention additionally relates to nucleic acid sequences, nucleic acid constructs, vectors and plants containing said nucleic acid sequences.

Even furthermore, the present invention relates generally to the field of molecular biology and concerns a method for improving various plant growth characteristics by modulating expression in a plant of a nucleic acid encoding a GASA (Gibberellic Acid-Stimulated *Arabidopsis*). The present invention also concerns plants having modulated expression of a nucleic acid encoding a GASA, which plants have improved growth characteristics relative to corresponding wild type plants or other control plants. The invention also provides constructs useful in the methods of the invention.

Yet furthermore, the present invention relates generally to the field of molecular biology and concerns a method for enhancing various economically important yield-related traits in plants. More specifically, the present invention concerns a method for enhancing yield-related traits in plants by

modulating expression in a plant of a nucleic acid encoding an AUX/IAA (auxin/indoleacetic acid) polypeptide. The present invention also concerns plants having modulated expression of a nucleic acid encoding IAA polypeptide, which plants have enhanced yield-related traits relative to control plants. The invention also provides constructs comprising AUX/IAA-encoding nucleic acids, useful in performing the methods of the invention.

The ever-increasing world population and the dwindling supply of arable land available for agriculture fuels research towards increasing the efficiency of agriculture. Conventional means for crop and horticultural improvements utilise selective breeding techniques to identify plants having desirable characteristics. However, such selective breeding techniques have several drawbacks, namely that these techniques are typically labour intensive and result in plants that often contain heterogeneous genetic components that may not always result in the desirable trait being passed on from parent plants. Advances in molecular biology have allowed mankind to modify the germplasm of animals and plants. Genetic engineering of plants entails the isolation and manipulation of genetic material (typically in the form of DNA or RNA) and the subsequent introduction of that genetic material into a plant. Such technology has the capacity to deliver crops or plants having various improved economic, agronomic or horticultural traits.

A trait of particular economic interest is increased yield. Yield is normally defined as the measurable produce of economic value from a crop. This may be defined in terms of quantity and/or quality. Yield is directly dependent on several factors, for example, the number and size of the organs, plant architecture (for example, the number of branches), seed production, leaf senescence and more. Root development, nutrient uptake, stress tolerance and early vigour may also be important factors in determining yield. Optimizing the above-mentioned factors may therefore contribute to increasing crop yield.

Seed yield is a particularly important trait, since the seeds of many plants are important for human and animal nutrition. Crops such as corn, rice, wheat, canola and soybean account for over half the total human caloric intake, whether through direct consumption of the seeds themselves or through consumption of meat products raised on processed seeds. They are also a source of sugars, oils and many kinds of metabolites used in industrial processes. Seeds contain an embryo (the source of new shoots and roots) and an endosperm (the source of nutrients for embryo growth during germination and during early growth of seedlings). The development of a seed involves many genes, and requires the transfer of metabolites from the roots, leaves and stems into the growing seed. The endosperm, in particular, assimilates the metabolic precursors of carbohydrates, oils and proteins and synthesizes them into storage macromolecules to fill out the grain.

Plant biomass is yield for forage crops like alfalfa, silage corn and hay. Many proxies for yield have been used in grain crops. Chief amongst these are estimates of plant size. Plant size can be measured in many ways depending on species and developmental stage, but include total plant dry weight, above-ground dry weight, above-ground fresh weight, leaf area, stem volume, plant height, rosette diameter, leaf length, root length, root mass, tiller number and leaf number. Many species maintain a conservative ratio between the size of different parts of the plant at a given developmental stage. These allometric relationships are used to extrapolate from one of these measures of size to another (e.g. Tittone et al 2005 *Agric Ecosys & Environ* 105: 213). Plant size at an early developmental stage will typically correlate with plant size

later in development. A larger plant with a greater leaf area can typically absorb more light and carbon dioxide than a smaller plant and therefore will likely gain a greater weight during the same period (Fasoula & Tollenaar 2005 Maydica 50:39). This is in addition to the potential continuation of the micro-environmental or genetic advantage that the plant had to achieve the larger size initially. There is a strong genetic component to plant size and growth rate (e.g. ter Steege et al 2005 Plant Physiology 139:1078), and so for a range of diverse genotypes plant size under one environmental condition is likely to correlate with size under another (Hittalmani et al 2003 Theoretical Applied Genetics 107:679). In this way a standard environment is used as a proxy for the diverse and dynamic environments encountered at different locations and times by crops in the field.

Another important trait for many crops is early vigour. Improving early vigour is an important objective of modern rice breeding programs in both temperate and tropical rice cultivars. Long roots are important for proper soil anchorage in water-seeded rice. Where rice is sown directly into flooded fields, and where plants must emerge rapidly through water, longer shoots are associated with vigour. Where drill-seeding is practiced, longer mesocotyls and coleoptiles are important for good seedling emergence. The ability to engineer early vigour into plants would be of great importance in agriculture. For example, poor early vigour has been a limitation to the introduction of maize (*Zea mays* L.) hybrids based on Corn Belt germplasm in the European Atlantic.

Harvest index, the ratio of seed yield to aboveground dry weight, is relatively stable under many environmental conditions and so a robust correlation between plant size and grain yield can often be obtained (e.g. Rebetzke et al 2002 Crop Science 42:739). These processes are intrinsically linked because the majority of grain biomass is dependent on current or stored photosynthetic productivity by the leaves and stem of the plant (Gardener et al 1985 Physiology of Crop Plants. Iowa State University Press, pp 68-73). Therefore, selecting for plant size, even at early stages of development, has been used as an indicator for future potential yield (e.g. Tittonell et al 2005 Agric Ecosys & Environ 105: 213). When testing for the impact of genetic differences on stress tolerance, the ability to standardize soil properties, temperature, water and nutrient availability and light intensity is an intrinsic advantage of greenhouse or plant growth chamber environments compared to the field. However, artificial limitations on yield due to poor pollination due to the absence of wind or insects, or insufficient space for mature root or canopy growth, can restrict the use of these controlled environments for testing yield differences. Therefore, measurements of plant size in early development, under standardized conditions in a growth chamber or greenhouse, are standard practices to provide indication of potential genetic yield advantages.

A further important trait is that of improved abiotic stress tolerance. Abiotic stress is a primary cause of crop loss worldwide, reducing average yields for most major crop plants by more than 50% (Wang et al., Planta (2003) 218: 1-14). Abiotic stresses may be caused by drought, salinity, extremes of temperature, chemical toxicity and oxidative stress. The ability to improve plant tolerance to abiotic stress would be of great economic advantage to farmers worldwide and would allow for the cultivation of crops during adverse conditions and in territories where cultivation of crops may not otherwise be possible.

Crop yield may therefore be increased by optimising one of the above-mentioned factors.

Depending on the end use, the modification of certain yield traits may be favoured over others. For example for applica-

tions such as forage or wood production, or bio-fuel resource, an increase in the vegetative parts of a plant may be desirable, and for applications such as flour, starch or oil production, an increase in seed parameters may be particularly desirable. Even amongst the seed parameters, some may be favoured over others, depending on the application. Various mechanisms may contribute to increasing seed yield, whether that is in the form of increased seed size or increased seed number.

One approach to increasing yield (seed yield and/or biomass) in plants may be through modification of the inherent growth mechanisms of a plant, such as the cell cycle or various signalling pathways involved in plant growth or in defense mechanisms.

Concerning ASPAT polypeptides, it has now been found that various yield-related traits may be improved in plants by modulating expression in a plant of a nucleic acid encoding an ASPAT (Aspartate AminoTransferase) in a plant.

Concerning MYB91 polypeptides, it has now been found that various yield-related traits may be increased in plants relative to control plants, by increasing expression in a plant of a nucleic acid sequence encoding a MYB91 like transcription factor (MYB91) polypeptide. The increased yield-related traits comprise one or more of: increased plant height, increased harvest index (HI), and increased Thousand Kernel Weight (TKW).

Concerning GASA polypeptides, it has now been found that various growth characteristics may be improved in plants by modulating expression in a plant of a nucleic acid encoding a GASA (Gibberellic Acid-Stimulated *Arabidopsis*) in a plant.

Concerning AUX/IAA polypeptides it has now been found that various growth characteristics may be improved in plants by modulating expression in a plant of a nucleic acid encoding an AUX/IAA polypeptide in a plant.

BACKGROUND

1. Aspartate AminoTransferase (ASPAT)

The capacity for growth, development and yield production of a plant is influenced by the regulation of carbon and nitrogen metabolisms and the N/C ratio in a the plant Lawlor 2002 Journal of Experimental Botany, Vol. 53, No. 370, pp. 773-787.

The enzyme Aspartate aminotransferase (ASPAT enzyme) catalyzes catalyses the reversible reaction of transamination between aspartate and 2-oxoglutarate to generate glutamate and oxaloacetate using pyridoxal 5'-phosphate (PLP) as essential cofactor in a reaction that can be express as: L-aspartate+2-oxoglutarate=oxaloacetate+L-glutamate.

The enzyme plays a key role in the metabolic regulation of carbon and nitrogen metabolism in all organisms. Structurally and functionally the ASPAT enzyme is conserved in all organisms. In eukaryots the enzyme plays a critical role in the interchanges of carbon and nitrogen pools between subcellular compartments.

Aspartate aminotransferases are classified into the group I of the aminotransferase superfamily (Jensen and Gu, 1996). Further, Aspartate Aminotransferases have been classified in four subgroups. Subgroup 1a includes the ASPATs from eubacteria and eukaryotes, whereas subgroup 1b comprises the enzymes from some eubacteria including cyanobacteria and archaeobacteria. A new group of ASPAT enzymes was described by De La Torre et al. 2006 Plant J. 2006, 46(3):414-25.

In plants, genes have been identified encoding ASPAT polypeptides that are targeted to different subcellular com-

partments and assembled into functional ASPAT Isoenzymes in the mitochondria, the cytosol, the peroxisome and the chloroplast.

2. MYB91 Like Transcription Factor (MYB91)

DNA-binding proteins are proteins that comprise any of many DNA-binding domains and thus have a specific or general affinity to DNA. DNA-binding proteins include for example transcription factors that modulate the process of transcription, nucleases that cleave DNA molecules, and histones that are involved in DNA packaging in the cell nucleus.

Transcription factors are usually defined as proteins that show sequence-specific DNA binding affinity and that are capable of activating and/or repressing transcription. The *Arabidopsis thaliana* genome codes for at least 1533 transcriptional regulators, accounting for ~5.9% of its estimated total number of genes (Riechmann et al. (2000) Science 290: 2105-2109). The Database of Rice Transcription Factors (DRTF) is a collection of known and predicted transcription factors of *Oryza sativa* L. ssp. indica and *Oryza sativa* L. ssp. japonica, and currently contains 2,025 putative transcription factors (TF) gene models in indica and 2,384 in japonica, distributed in 63 families (Gao et al. (2006) Bioinformatics 2006, 22(10):1286-7).

One of these families is the MYB domain family of transcription factors, characterized by a highly conserved DNA-binding domain, the MYB domain. The MYB domain was originally described in the oncogene (v-myb) of avian myeloblastosis virus (Klempnauer et al. (1982) Cell 33, 453-63). Many vertebrates contain three genes related to v-Myb c-Myb, A-Myb and B-Myb and other similar genes have been identified in insects, plants, fungi and slime molds. The encoded proteins are crucial to the control of proliferation and differentiation in a number of cell types. MYB proteins contain one to four imperfect direct repeats of a conserved sequence of 50-53 amino acids which encodes a helix-turn-helix structure involved in DNA binding (Rosinski and Atchley (1998) J Mol Evol 46, 74-83). Three regularly spaced tryptophan residues, which form a tryptophan cluster in the three-dimensional helix-turn-helix structure, are characteristic of a MYB repeat. The three repeats in c-Myb are referred to as R1, R2 and R3; and repeats from other MYB proteins are categorised according to their similarity to R1, R2 or R3. Since there is limited sequence conservation outside of the MYB domain, MYB proteins have been clustered into subgroups based on conserved motifs identified outside of the MYB coding region (Jiang et al. (2004) Genome Biology 5, R46).

AtMYB91 belongs to the R2R3-MYB gene family (Li and Parish, Plant J. 8, 963-972, 1995), which is a large gene family (with reportedly 126 genes in *Arabidopsis thaliana* (Zimmerman et al., Plant J. 40, 22-34, 2004)). Members of this group are involved in various processes, including secondary metabolism, cell morphogenesis, regulation of meristem formation, flower and seed development, cell cycle, defense and stress responses, light and hormone signalling (Chen et al., Cell Res. 16, 797-798, 2006). AtMYB91 is also named AS1 asymmetric leaves 1, and is closely related to Antirrhinum PHAN phantastica and to maize ROUGH SHEATH2 (RS2) polypeptides (Sun et al. (2002) Planta 214 (5):694-702), all having an evolutionarily conserved role in specification of leaf cell identity, in particular in dorsal-ventral identity. In *Arabidopsis*, AS1 is expressed in leaf founder cells, where it functions as a heterodimer with the structurally unrelated AS2 proteins to repress activity of KNOTTED 1-like homeobox (KNOX) genes.

3. Gibberellic Acid-Stimulated *Arabidopsis* (GASA)

GASA (Gibberellic Acid-Stimulated *Arabidopsis*) proteins are plant-specific and are expressed during a variety of physiological processes. Several GASA-like genes are hormone responsive, expression of tomato gene GAST1, the first member of the family to be characterized, was induced upon application of exogenous gibberellin in a gibberellin-deficient background (Shi et al. Plant J. 2, 153-159, 1992). A related tomato gene, RSI-1, shares high sequence identity with GAST1 and is activated during lateral root formation (Taylor and Scheuring, Mol. Gen. Genet. 243, 148-157, 1994). GASA1 to GASA4 from *Arabidopsis* were first identified based on their similarity to tomato GAST1 (Herzog et al. Plant Mol. Biol. 27, 743-752, 1995). Expression data indicated that GASA1 accumulates in flower buds and immature siliques, GASA2 and GASA3 in siliques and dry seeds, and GASA4 in growing roots and flower buds. GASA4 is reported to be expressed in all meristematic regions (Aubert et al., Plant Mol. Biol. 36, 871-883, 1998).

Functionally, the GASA proteins are not well characterised. GASA proteins are reportedly involved in pathogen responses and in plant development. Plants ectopically expressing GEG, a GASA homologue from *Gerbera hybrida*, showed shorter corollas with decreased cell length compared with the wild type, indicating a role for GEG as an inhibitor of cell elongation. Overexpression of *Arabidopsis* GASA4 resulted in plants having increased seed weight (Roxrud et al, Plant Cell Physiol. 48, 471-483, 2007). However, these plants in addition had occasional meristem identity changes with reversion from floral meristems development to normal indeterminate inflorescence development. Furthermore, modulated GASA4 expression caused a significant increase of branching. Overexpression of *Arabidopsis* GASA4 also increased tolerance to heat stress (Ko et al., Plant Physiol. Biochem. 45, 722-728, 2007).

4. Auxin/Indoleacetic Acid Genes (AUX/IAA)

The AUX/IAA (auxin/indoleacetic acid) genes encode a family of proteins whose expression is tightly regulated by auxin. The plant hormone auxin is involved in various processes like cell division, cell expansion and differentiation, patterning of embryos, vasculature or other tissues, regulation of growth of primary and lateral root or shoot meristems. AUX/IAA proteins furthermore are usually expressed in a tissue-specific manner.

AUX/IAA proteins typically have four conserved amino acid sequence motifs (domains I, II, III and IV) and have nuclear localisation signal sequences. Domains I and II are postulated to destabilize the protein and may be involved in protein turnover. Domains III and IV are postulated to be involved in protein-protein interactions: AUX/IAA proteins can form homodimers and are known to associate with ARF proteins. The AUX/IAA-ARF complexes are likely to be involved in auxin mediated gene expression. The Aux/IAA proteins are negative regulators of the auxin response factors (ARFs) that regulate expression of auxin-responsive genes. Aux/IAA proteins bind to the DNA-bound ARF partner proteins and repress ARF activity. In the auxin activated status, Aux/IAA proteins are ubiquitinated via interactions with the auxin-modified SCFTIR1 complex and subsequently degraded by 26S proteasome action. An overview of roles and activities of AUX/IAA proteins is given by Reed (Trends in Plant Science 6, 420-425, 2001). The structure and expression analysis of early auxin-responsive Aux/IAA gene family in rice (*Oryza sativa*) has recently been reported by Jain et al. 2006 Funct Integr Genomics. 2006 January; 6(1):47-59.

IAA14 is a AUX/IAA protein that acts as a transcriptional repressor in lateral root formation. A gain of function muta-

tion in IAA14 blocks early pericycle divisions that initiate lateral root development (Fukaki et al., Plant J. 29, 153-168, 2002).

SUMMARY

1. Aspartate Amino Transferase (ASPAT)

Surprisingly, it has now been found that modulating expression of a nucleic acid encoding an ASPAT polypeptide gives plants having enhanced yield-related traits relative to control plants.

According one embodiment, there is provided a method for enhancing yield-related traits relative to control plants, comprising modulating expression of a nucleic acid encoding an ASPAT polypeptide in a plant.

2. MYB91 Like Transcription Factor (MYB91)

Surprisingly, it has now been found that increasing expression in a plant of a nucleic acid sequence encoding a MYB91 like transcription factor (MYB91) polypeptide as defined herein, gives plants having increased yield-related traits relative to control plants.

According to one embodiment, there is provided a method for increasing yield-related traits in plants relative to control plants, comprising increasing expression in a plant of a nucleic acid sequence encoding a MYB91 like transcription factor (MYB91) as defined herein. The increased yield-related traits comprise one or more of: increased plant height, increased harvest index (HI), and increased Thousand Kernel Weight (TKW).

3. Gibberellic Acid-Stimulated *Arabidopsis* (GASA)

Surprisingly, it has now been found that modulating expression of a nucleic acid encoding a GASA polypeptide gives plants having enhanced yield-related traits, in particular increased yield relative to control plants.

According one embodiment, there is provided a method for improving yield related traits of a plant, relative to control plants, comprising modulating expression of a nucleic acid encoding a GASA polypeptide in a plant.

4. Auxin/Indoleacetic Acid Genes (AUX/IAA)

Surprisingly, it has now been found that modulating expression of a nucleic acid encoding an AUX/IAA polypeptide gives plants having enhanced yield-related traits, in particular increased yield relative to control plants.

According one embodiment, there is provided a method for improving yield related traits of a plant relative to control plants, comprising modulating expression of a nucleic acid encoding an AUX/IAA polypeptide in a plant, wherein the yield related traits do not encompass increased root growth.

DEFINITIONS

Polypeptide(s)/Protein(s)

The terms “polypeptide” and “protein” are used interchangeably herein and refer to amino acids in a polymeric form of any length, linked together by peptide bonds.

Polynucleotide(s)/Nucleic Acid(s)/Nucleic Acid Sequence(s)/Nucleotide Sequence(s)

The terms “polynucleotide(s)”, “nucleic acid sequence(s)”, “nucleotide sequence(s)”, “nucleic acid(s)”, “nucleic acid molecule” are used interchangeably herein and refer to nucleotides, either ribonucleotides or deoxyribonucleotides or a combination of both, in a polymeric unbranched form of any length.

Control Plant(s)

The choice of suitable control plants is a routine part of an experimental setup and may include corresponding wild type plants or corresponding plants without the gene of interest.

The control plant is typically of the same plant species or even of the same variety as the plant to be assessed. The control plant may also be a nullizygote of the plant to be assessed. Nullizygotes are individuals missing the transgene by segregation. A “control plant” as used herein refers not only to whole plants, but also to plant parts, including seeds and seed parts.

Homologue(s)

“Homologues” of a protein encompass peptides, oligopeptides, polypeptides, proteins and enzymes having amino acid substitutions, deletions and/or insertions relative to the unmodified protein in question and having similar biological and functional activity as the unmodified protein from which they are derived.

A deletion refers to removal of one or more amino acids from a protein.

An insertion refers to one or more amino acid residues being introduced into a predetermined site in a protein. Insertions may comprise N-terminal and/or C-terminal fusions as well as intra-sequence insertions of single or multiple amino acids. Generally, insertions within the amino acid sequence will be smaller than N- or C-terminal fusions, of the order of about 1 to 10 residues. Examples of N- or C-terminal fusion proteins or peptides include the binding domain or activation domain of a transcriptional activator as used in the yeast two-hybrid system, phage coat proteins, (histidine)-6-tag, glutathione S-transferase-tag, protein A, maltose-binding protein, dihydrofolate reductase, Tag•100 epitope, c-myc epitope, FLAG®-epitope, lacZ, CMP (calmodulin-binding peptide), HA epitope, protein C epitope and VSV epitope.

A substitution refers to replacement of amino acids of the protein with other amino acids having similar properties (such as similar hydrophobicity, hydrophilicity, antigenicity, propensity to form or break α -helical structures or β -sheet structures). Amino acid substitutions are typically of single residues, but may be clustered depending upon functional constraints placed upon the polypeptide; insertions will usually be of the order of about 1 to 10 amino acid residues. The amino acid substitutions are preferably conservative amino acid substitutions. Conservative substitution tables are well known in the art (see for example Creighton (1984) Proteins. W.H. Freeman and Company (Eds) and Table 1 below).

TABLE 1

Examples of conserved amino acid substitutions	
Residue	Conservative Substitutions
Ala	Ser
Arg	Lys
Asn	Gln; His
Asp	Glu
Gln	Asn
Cys	Ser
Glu	Asp
Gly	Pro
His	Asn; Gln
Ile	Leu; Val
Leu	Ile; Val
Lys	Arg; Gln
Met	Leu; Ile
Phe	Met; Leu; Tyr
Ser	Thr; Gly
Thr	Ser; Val
Trp	Tyr
Tyr	Trp; Phe
Val	Ile; Leu

Amino acid substitutions, deletions and/or insertions may readily be made using peptide synthetic techniques well known in the art, such as solid phase peptide synthesis and the like, or by recombinant DNA manipulation. Methods for the

manipulation of DNA sequences to produce substitution, insertion or deletion variants of a protein are well known in the art. For example, techniques for making substitution mutations at predetermined sites in DNA are well known to those skilled in the art and include M13 mutagenesis, T7-Gen in vitro mutagenesis (USB, Cleveland, Ohio), QuickChange Site Directed mutagenesis (Stratagene, San Diego, Calif.), PCR-mediated site-directed mutagenesis or other site-directed mutagenesis protocols.

Derivatives

"Derivatives" include peptides, oligopeptides, polypeptides which may, compared to the amino acid sequence of the naturally-occurring form of the protein, such as the protein of interest, comprise substitutions of amino acids with non-naturally occurring amino acid residues, or additions of non-naturally occurring amino acid residues. "Derivatives" of a protein also encompass peptides, oligopeptides, polypeptides which comprise naturally occurring altered (glycosylated, acylated, prenylated, phosphorylated, myristoylated, sulphated etc.) or non-naturally altered amino acid residues compared to the amino acid sequence of a naturally-occurring form of the polypeptide. A derivative may also comprise one or more non-amino acid substituents or additions compared to the amino acid sequence from which it is derived, for example a reporter molecule or other ligand, covalently or non-covalently bound to the amino acid sequence, such as a reporter molecule which is bound to facilitate its detection, and non-naturally occurring amino acid residues relative to the amino acid sequence of a naturally-occurring protein. Furthermore, "derivatives" also include fusions of the naturally-occurring form of the protein with tagging peptides such as FLAG, HIS6 or thioredoxin (for a review of tagging peptides, see Terpe, Appl. Microbiol. Biotechnol. 60, 523-533, 2003).

Orthologue(s)/Paralogue(s)

Orthologues and paralogues encompass evolutionary concepts used to describe the ancestral relationships of genes. Paralogues are genes within the same species that have originated through duplication of an ancestral gene; orthologues are genes from different organisms that have originated through speciation, and are also derived from a common ancestral gene.

Domain

The term "domain" refers to a set of amino acids conserved at specific positions along an alignment of sequences of evolutionarily related proteins. While amino acids at other positions can vary between homologues, amino acids that are highly conserved at specific positions indicate amino acids that are likely essential in the structure, stability or function of a protein. Identified by their high degree of conservation in aligned sequences of a family of protein homologues, they can be used as identifiers to determine if any polypeptide in question belongs to a previously identified polypeptide family.

Motif/Consensus sequence/Signature

The term "motif" or "consensus sequence" or "signature" refers to a short conserved region in the sequence of evolutionarily related proteins. Motifs are frequently highly conserved parts of domains, but may also include only part of the domain, or be located outside of conserved domain (if all of the amino acids of the motif fall outside of a defined domain).

Hybridisation

The term "hybridisation" as defined herein is a process wherein substantially homologous complementary nucleotide sequences anneal to each other. The hybridisation process can occur entirely in solution, i.e. both complementary nucleic acids are in solution. The hybridisation process can also occur with one of the complementary nucleic acids immobilised to a matrix such as magnetic beads, Sepharose beads or any other resin.

The hybridisation process can furthermore occur with one of the complementary nucleic acids immobilised to a solid support such as a nitro-cellulose or nylon membrane or immobilised by e.g. photolithography to, for example, a siliceous glass support (the latter known as nucleic acid arrays or microarrays or as nucleic acid chips). In order to allow hybridisation to occur, the nucleic acid molecules are generally thermally or chemically denatured to melt a double strand into two single strands and/or to remove hairpins or other secondary structures from single stranded nucleic acids.

The term "stringency" refers to the conditions under which a hybridisation takes place. The stringency of hybridisation is influenced by conditions such as temperature, salt concentration, ionic strength and hybridisation buffer composition. Generally, low stringency conditions are selected to be about 30° C. lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. Medium stringency conditions are when the temperature is 20° C. below T_m , and high stringency conditions are when the temperature is 10° C. below T_m . High stringency hybridisation conditions are typically used for isolating hybridising sequences that have high sequence similarity to the target nucleic acid sequence. However, nucleic acids may deviate in sequence and still encode a substantially identical polypeptide, due to the degeneracy of the genetic code. Therefore medium stringency hybridisation conditions may sometimes be needed to identify such nucleic acid molecules.

The T_m is the temperature under defined ionic strength and pH, at which 50% of the target sequence hybridises to a perfectly matched probe. The T_m is dependent upon the solution conditions and the base composition and length of the probe. For example, longer sequences hybridise specifically at higher temperatures. The maximum rate of hybridisation is obtained from about 16° C. up to 32° C. below T_m . The presence of monovalent cations in the hybridisation solution reduce the electrostatic repulsion between the two nucleic acid strands thereby promoting hybrid formation; this effect is visible for sodium concentrations of up to 0.4M (for higher concentrations, this effect may be ignored). Formamide reduces the melting temperature of DNA-DNA and DNA-RNA duplexes with 0.6 to 0.7° C. for each percent formamide, and addition of 50% formamide allows hybridisation to be performed at 30 to 45° C., though the rate of hybridisation will be lowered. Base pair mismatches reduce the hybridisation rate and the thermal stability of the duplexes. On average and for large probes, the T_m decreases about 1° C. per % base mismatch. The T_m may be calculated using the following equations, depending on the types of hybrids:

1) DNA-DNA hybrids (Meinkoth and Wahl, Anal. Biochem., 138: 267-284, 1984):

$$T_m = 81.5^\circ \text{C.} + 16.6 \times \log_{10} [\text{Na}^+]^a + 0.41 \times \% [G/C]^b - 500 \times [L]^{-1} - 0.61 \times \% \text{ formamide}$$

2) DNA-RNA or RNA-RNA hybrids:

$$T_m = 79.8 + 18.5 (\log_{10} [\text{Na}^+]^a) + 0.58 (\% G/C)^b + 11.8 (\% G/C)^2 - 820/L^c$$

3) oligo-DNA or oligo-RNAs hybrids:

$$\text{For } <20 \text{ nucleotides: } T_m = 2(I_n)$$

$$\text{For } 20-35 \text{ nucleotides: } T_m = 22 + 1.46(I_n)$$

^a or for other monovalent cation, but only accurate in the 0.01-0.4 M range.

^b only accurate for % GC in the 30% to 75% range.

^c L=length of duplex in base pairs.

^d oligo, oligonucleotide; I_n = effective length of primer = $2 \times (\text{no. of G/C}) + (\text{no. of NT})$.

Non-specific binding may be controlled using any one of a number of known techniques such as, for example, blocking

the membrane with protein containing solutions, additions of heterologous RNA, DNA, and SDS to the hybridisation buffer, and treatment with Rnase. For non-homologous probes, a series of hybridizations may be performed by varying one of (i) progressively lowering the annealing temperature (for example from 68° C. to 42° C.) or (ii) progressively lowering the formamide concentration (for example from 50% to 0%). The skilled artisan is aware of various parameters which may be altered during hybridisation and which will either maintain or change the stringency conditions.

Besides the hybridisation conditions, specificity of hybridisation typically also depends on the function of post-hybridisation washes. To remove background resulting from non-specific hybridisation, samples are washed with dilute salt solutions. Critical factors of such washes include the ionic strength and temperature of the final wash solution: the lower the salt concentration and the higher the wash temperature, the higher the stringency of the wash. Wash conditions are typically performed at or below hybridisation stringency. A positive hybridisation gives a signal that is at least twice of that of the background. Generally, suitable stringent conditions for nucleic acid hybridisation assays or gene amplification detection procedures are as set forth above. More or less stringent conditions may also be selected. The skilled artisan is aware of various parameters which may be altered during washing and which will either maintain or change the stringency conditions.

For example, typical high stringency hybridisation conditions for DNA hybrids longer than 50 nucleotides encompass hybridisation at 65° C. in 1×SSC or at 42° C. in 1×SSC and 50% formamide, followed by washing at 65° C. in 0.3×SSC. Examples of medium stringency hybridisation conditions for DNA hybrids longer than 50 nucleotides encompass hybridisation at 50° C. in 4×SSC or at 40° C. in 6×SSC and 50% formamide, followed by washing at 50° C. in 2×SSC. The length of the hybrid is the anticipated length for the hybridising nucleic acid. When nucleic acids of known sequence are hybridised, the hybrid length may be determined by aligning the sequences and identifying the conserved regions described herein. 1×SSC is 0.15M NaCl and 15 mM sodium citrate; the hybridisation solution and wash solutions may additionally include 5×Denhardt's reagent, 0.5-1.0% SDS, 100 µg/ml denatured, fragmented salmon sperm DNA, 0.5% sodium pyrophosphate.

For the purposes of defining the level of stringency, reference can be made to Sambrook et al. (2001) *Molecular Cloning: a laboratory manual*, 3rd Edition, Cold Spring Harbor Laboratory Press, CSH, New York or to *Current Protocols in Molecular Biology*, John Wiley & Sons, N.Y. (1989 and yearly updates).

Splice Variant

The term "splice variant" as used herein encompasses variants of a nucleic acid sequence in which selected introns and/or exons have been excised, replaced, displaced or added, or in which introns have been shortened or lengthened. Such variants will be ones in which the biological activity of the protein is substantially retained; this may be achieved by selectively retaining functional segments of the protein. Such splice variants may be found in nature or may be manmade. Methods for predicting and isolating such splice variants are well known in the art (see for example Foissac and Schiex (2005) *BMC Bioinformatics* 6: 25).

Allelic Variant

Alleles or allelic variants are alternative forms of a given gene, located at the same chromosomal position. Allelic variants encompass Single Nucleotide Polymorphisms (SNPs), as well as Small Insertion/Deletion Polymorphisms (IN-

DELs). The size of INDELs is usually less than 100 bp. SNPs and INDELs form the largest set of sequence variants in naturally occurring polymorphic strains of most organisms. Gene Shuffling/Directed Evolution

Gene shuffling or directed evolution consists of iterations of DNA shuffling followed by appropriate screening and/or selection to generate variants of nucleic acids or portions thereof encoding proteins having a modified biological activity (Castle et al., (2004) *Science* 304(5674): 1151-4; U.S. Pat. Nos. 5,811,238 and 6,395,547).

Regulatory Element/Control Sequence/Promoter

The terms "regulatory element", "control sequence" and "promoter" are all used interchangeably herein and are to be taken in a broad context to refer to regulatory nucleic acid sequences capable of effecting expression of the sequences to which they are ligated. The term "promoter" typically refers to a nucleic acid control sequence located upstream from the transcriptional start of a gene and which is involved in recognising and binding of RNA polymerase and other proteins, thereby directing transcription of an operably linked nucleic acid. Encompassed by the aforementioned terms are transcriptional regulatory sequences derived from a classical eukaryotic genomic gene (including the TATA box which is required for accurate transcription initiation, with or without a CCAAT box sequence) and additional regulatory elements (i.e. upstream activating sequences, enhancers and silencers) which alter gene expression in response to developmental and/or external stimuli, or in a tissue-specific manner. Also included within the term is a transcriptional regulatory sequence of a classical prokaryotic gene, in which case it may include a -35 box sequence and/or -10 box transcriptional regulatory sequences. The term "regulatory element" also encompasses a synthetic fusion molecule or derivative that confers, activates or enhances expression of a nucleic acid molecule in a cell, tissue or organ.

A "plant promoter" comprises regulatory elements, which mediate the expression of a coding sequence segment in plant cells. Accordingly, a plant promoter need not be of plant origin, but may originate from viruses or micro-organisms, for example from viruses which attack plant cells. The "plant promoter" can also originate from a plant cell, e.g. from the plant which is transformed with the nucleic acid sequence to be expressed in the inventive process and described herein. This also applies to other "plant" regulatory signals, such as "plant" terminators. The promoters upstream of the nucleotide sequences useful in the methods of the present invention can be modified by one or more nucleotide substitution(s), insertion(s) and/or deletion(s) without interfering with the functionality or activity of either the promoters, the open reading frame (ORF) or the 3'-regulatory region such as terminators or other 3' regulatory regions which are located away from the ORF. It is furthermore possible that the activity of the promoters is increased by modification of their sequence, or that they are replaced completely by more active promoters, even promoters from heterologous organisms. For expression in plants, the nucleic acid molecule must, as described above, be linked operably to or comprise a suitable promoter which expresses the gene at the right point in time and with the required spatial expression pattern.

For the identification of functionally equivalent promoters, the promoter strength and/or expression pattern of a candidate promoter may be analysed for example by operably linking the promoter to a reporter gene and assaying the expression level and pattern of the reporter gene in various tissues of the plant. Suitable well-known reporter genes include for example beta-glucuronidase or beta-galactosidase. The promoter activity is assayed by measuring the enzy-

matic activity of the beta-glucuronidase or beta-galactosidase. The promoter strength and/or expression pattern may then be compared to that of a reference promoter (such as the one used in the methods of the present invention). Alternatively, promoter strength may be assayed by quantifying mRNA levels or by comparing mRNA levels of the nucleic acid used in the methods of the present invention, with mRNA levels of housekeeping genes such as 18S rRNA, using methods known in the art, such as Northern blotting with densitometric analysis of autoradiograms, quantitative real-time PCR or RT-PCR (Heid et al., 1996 *Genome Methods* 6: 986-994). Generally by “weak promoter” is intended a promoter that drives expression of a coding sequence at a low level. By “low level” is intended at levels of about 1/10,000 transcripts to about 1/100,000 transcripts, to about 1/500,000 transcripts per cell. Conversely, a “strong promoter” drives expression of a coding sequence at high level, or at about 1/10 transcripts to about 1/100 transcripts to about 1/1000 transcripts per cell. Generally, by “medium strength promoter” is intended a promoter that drives expression of a coding sequence at a lower level than a strong promoter, in particular at a level that is in all instances below that obtained when under the control of a 35S CaMV promoter.

Operably Linked

The term “operably linked” as used herein refers to a functional linkage between the promoter sequence and the gene of interest, such that the promoter sequence is able to initiate transcription of the gene of interest.

Constitutive Promoter

A “constitutive promoter” refers to a promoter that is transcriptionally active during most, but not necessarily all, phases of growth and development and under most environmental conditions, in at least one cell, tissue or organ. Table 2a below gives examples of constitutive promoters.

TABLE 2a

Examples of constitutive promoters	
Gene Source	Reference
Actin	McElroy et al, <i>Plant Cell</i> , 2: 163-171, 1990
HMGP	WO 2004/070039
CAMV 35S	Odell et al, <i>Nature</i> , 313: 810-812, 1985
CaMV 19S	Nilsson et al., <i>Physiol. Plant.</i> 100: 456-462, 1997
GOS2	de Pater et al, <i>Plant J Nov</i> ; 2(6): 837-44, 1992, WO 2004/065596
Ubiquitin	Christensen et al, <i>Plant Mol. Biol.</i> 18: 675-689, 1992
Rice cyclophilin	Buchholz et al, <i>Plant Mol Biol.</i> 25(5): 837-43, 1994
Maize H3 histone	Lepetit et al, <i>Mol. Gen. Genet.</i> 231: 276-285, 1992
Alfalfa H3 histone	Wu et al. <i>Plant Mol. Biol.</i> 11: 641-649, 1988
Actin 2	An et al, <i>Plant J.</i> 10(1): 107-121, 1996
34S FMV	Sanger et al., <i>Plant. Mol. Biol.</i> , 14, 1990: 433-443
Rubisco small subunit	U.S. Pat. No. 4,962,028
OCS	Leisner (1988) <i>Proc Natl Acad Sci USA</i> 85(5): 2553
SAD1	Jain et al., <i>Crop Science</i> , 39 (6), 1999: 1696
SAD2	Jain et al., <i>Crop Science</i> , 39 (6), 1999: 1696
nos	Shaw et al. (1984) <i>Nucleic Acids Res.</i> 12(20): 7831-7846
V-ATPase	WO 01/14572
Super promoter	WO 95/14098
G-box proteins	WO 94/12015

Ubiquitous Promoter

A ubiquitous promoter is active in substantially all tissues or cells of an organism.

Developmentally-Regulated Promoter

A developmentally-regulated promoter is active during certain developmental stages or in parts of the plant that undergo developmental changes.

Inducible Promoter

An inducible promoter has induced or increased transcription initiation in response to a chemical (for a review see Gatz 1997, *Annu. Rev. Plant Physiol. Plant Mol. Biol.*, 48:89-108), environmental or physical stimulus, or may be “stress-inducible”, i.e. activated when a plant is exposed to various stress conditions, or a “pathogen-inducible” i.e. activated when a plant is exposed to exposure to various pathogens.

Organ-Specific/Tissue-Specific Promoter

An organ-specific or tissue-specific promoter is one that is capable of preferentially initiating transcription in certain organs or tissues, such as the leaves, roots, seed tissue etc. For example, a “root-specific promoter” is a promoter that is transcriptionally active predominantly in plant roots, substantially to the exclusion of any other parts of a plant, whilst still allowing for any leaky expression in these other plant parts. Promoters able to initiate transcription in certain cells only are referred to herein as “cell-specific”.

Examples of root-specific promoters are listed in Table 2b below:

TABLE 2b

Examples of root-specific promoters	
Gene Source	Reference
RCc3	<i>Plant Mol Biol.</i> 1995 January; 27(2): 237-48
<i>Arabidopsis</i> PHT1	Kovama et al., 2005; Mudge et al. (2002, <i>Plant J.</i> 31: 341)
<i>Medicago</i> phosphate transporter	Xiao et al., 2006
<i>Arabidopsis</i> Pyk10 root-expressible genes	Nitz et al. (2001) <i>Plant Sci</i> 161 (2): 337-346
tobacco auxin-inducible gene	Tingey et al., <i>EMBO J.</i> 6: 1, 1987.
β -tubulin	Van der Zaal et al., <i>Plant Mol. Biol.</i> 16, 983, 1991.
tobacco root-specific genes	Oppenheimer, et al., <i>Gene</i> 63: 87, 1988.
<i>B. napus</i> G1-3b gene	Conkling, et al., <i>Plant Physiol.</i> 93: 1203, 1990.
SbPRP1	U.S. Pat. No. 5,401,836
LRX1	Suzuki et al., <i>Plant Mol. Biol.</i> 21: 109-119, 1993.
BTG-26 <i>Brassica napus</i>	Baumberger et al. 2001, <i>Genes & Dev.</i> 15: 1128
LeAMT1 (tomato)	US 20050044585
The LeNRT1-1 (tomato)	Lauter et al. (1996, <i>PNAS</i> 3: 8139)
class I patatin gene (potato)	Lauter et al. (1996, <i>PNAS</i> 3: 8139)
KDC1 (<i>Daucus carota</i>)	Liu et al., <i>Plant Mol. Biol.</i> 153: 386-395, 1991.
TobRB7 gene	Downey et al. (2000, <i>J. Biol. Chem.</i> 275: 39420)
OsRAB5a (rice)	W Song (1997) PhD Thesis, North Carolina State University, Raleigh, NC USA
ALF5 (<i>Arabidopsis</i>)	Wang et al. 2002, <i>Plant Sci.</i> 163: 273
NRT2; 1Np (<i>N. plumbaginifolia</i>)	Diener et al. (2001, <i>Plant Cell</i> 13: 1625)
	Quesada et al. (1997, <i>Plant Mol. Biol.</i> 34: 265)

A seed-specific promoter is transcriptionally active predominantly in seed tissue, but not necessarily exclusively in seed tissue (in cases of leaky expression). The seed-specific promoter may be active during seed development and/or during germination. The seed specific promoter may be endosperm/aleurone/embryo specific. Examples of seed-specific promoters (endosperm/aleurone/embryo specific) are shown in Table 2c to Table 2f below. Further examples of seed-specific promoters are given in Qing Qu and Takaiwa (Plant Biotechnol. J. 2, 113-125, 2004), which disclosure is incorporated by reference herein as if fully set forth.

TABLE 2c

Examples of seed-specific promoters	
Gene source	Reference
seed-specific genes	Simon et al., Plant Mol. Biol. 5: 191, 1985; Scofield et al., J. Biol. Chem. 262: 12202, 1987; Baszczynski et al., Plant Mol. Biol. 14: 633, 1990.
Brazil Nut albumin	Pearson et al., Plant Mol. Biol. 18: 235-245, 1992.
legumin	Ellis et al., Plant Mol. Biol. 10: 203-214, 1988.
glutelin (rice)	Takaiwa et al., Mol. Gen. Genet. 208: 15-22, 1986; Takaiwa et al., FEBS Letts. 221: 43-47, 1987.
zein	Matzke et al Plant Mol Biol, 14(3): 323-32 1990
napA	Stalberg et al, Planta 199: 515-519, 1996.
wheat LMW and HMW glutenin-1	Mol Gen Genet 216: 81-90, 1989; NAR 17: 461-2, 1989
wheat SPA	Albani et al, Plant Cell, 9: 171-184, 1997
wheat α , β , γ -gliadins	EMBO J. 3: 1409-15, 1984
barley Itr1 promoter	Diaz et al. (1995) Mol Gen Genet 248(5): 592-8
barley B1, C, D, hordein	Theor Appl Gen 98: 1253-62, 1999; Plant J 4: 343-55, 1993; Mol Gen Genet 250: 750-60, 1996
barley DOF	Mena et al, The Plant Journal, 116(1): 53-62, 1998
blz2	EP99106056.7
synthetic promoter	Vicente-Carbajosa et al., Plant J, 13: 629-640, 1998.
rice prolamin NRP33	Wu et al, Plant Cell Physiology 39(8) 885-889, 1998
rice α -globulin G1b-1	Wu et al, Plant Cell Physiology 39(8) 885-889, 1998
rice OSH1	Sato et al, Proc. Natl. Acad. Sci. USA, 93: 8117-8122, 1996
rice α -globulin REB/OHP-1	Nakase et al. Plant Mol. Biol. 33: 513-522, 1997
rice ADP-glucose pyrophos- phorylase	Trans Res 6: 157-68, 1997
maize ESR gene family	Plant J 12: 235-46, 1997
<i>sorghum</i> α -kafirin	DeRose et al., Plant Mol. Biol 32: 1029-35, 1996
KNOX	Postma-Haarsma et al, Plant Mol. Biol. 39: 257-71, 1999
rice oleosin	Wu et al, J. Biochem. 123: 386, 1998
sunflower oleosin	Cummins et al., Plant Mol. Biol. 19: 873-876, 1992
PRO0117, putative rice 40S ribosomal protein	WO 2004/070039
PRO0136, rice alanine aminotransferase	unpublished
PRO0147, trypsin inhibitor	unpublished
ITR1 (barley)	
PRO0151, rice WSI18	WO 2004/070039
PRO0175, rice RAB21	WO 2004/070039
PRO005	WO 2004/070039
PRO0095	WO 2004/070039
α -amylase (Amy32b)	Lanahan et al, Plant Cell 4: 203-211, 1992; Skriver et al, Proc Natl Acad Sci USA 88: 7266-7270, 1991
cathepsin β -like gene	Cejudo et al, Plant Mol Biol 20: 849-856, 1992
Barley Ltp2	Kalla et al., Plant J. 6: 849-60, 1994
Chi26	Leah et al., Plant J. 4: 579-89, 1994
Maize B-Peru	Selinger et al., Genetics 149: 1125-38, 1998

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TABLE 2d

examples of endosperm-specific promoters	
Gene source	Reference
glutelin (rice)	Takaiwa et al. (1986) Mol Gen Genet 208: 15-22; Takaiwa et al. (1987) FEBS Letts. 221: 43-47
zein	Matzke et al., (1990) Plant Mol Biol 14(3): 323-32
wheat LMW and HMW	Colot et al. (1989) Mol Gen Genet 216: 81-90, Anderson et al. (1989) NAR 17: 461-2
glutenin-1	
wheat SPA	Albani et al. (1997) Plant Cell 9: 171-184
wheat gliadins	Rafalski et al. (1984) EMBO 3: 1409-15
barley Itr1 promoter	Diaz et al. (1995) Mol Gen Genet 248(5): 592-8
barley B1, C, D, hordein	Cho et al. (1999) Theor Appl Genet 98: 1253-62; Muller et al. (1993) Plant J 4: 343-55; Sorenson et al. (1996) Mol Gen Genet 250: 750-60
barley DOF	Mena et al, (1998) Plant J 116(1): 53-62
blz2	Onate et al. (1999) J Biol Chem 274(14): 9175-82
synthetic promoter	Vicente-Carbajosa et al. (1998) Plant J 13: 629-640
rice prolamin NRP33	Wu et al, (1998) Plant Cell Physiol 39(8) 885-889

TABLE 2d-continued

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65

examples of endosperm-specific promoters	
Gene source	Reference
rice globulin G1b-1	Wu et al. (1998) Plant Cell Physiol 39(8) 885-889
rice globulin REB/OHP-1	Nakase et al. (1997) Plant Molec Biol 33: 513-522
rice ADP-glucose pyrophosphorylase	Russell et al. (1997) Trans Res 6: 157-68
maize ESR gene family	Opsahl-Ferstad et al. (1997) Plant J 12: 235-46
<i>sorghum</i> kafirin	DeRose et al. (1996) Plant Mol Biol 32: 1029-35

TABLE 2e

Examples of embryo specific promoters:	
Gene source	Reference
rice OSH1	Sato et al, Proc. Natl. Acad. Sci. USA, 93: 8117-8122, 1996

TABLE 2e-continued

Examples of embryo specific promoters:	
Gene source	Reference
KNOX	Postma-Haarsma et al, Plant Mol. Biol. 39: 257-71, 1999
PRO0151	WO 2004/070039
PRO0175	WO 2004/070039
PRO005	WO 2004/070039
PRO0095	WO 2004/070039

TABLE 2f

Examples of aleurone-specific promoters:	
Gene source	Reference
α -amylase (Amy32b)	Lanahan et al, Plant Cell 4: 203-211, 1992; Skriver et al, Proc Natl Acad Sci USA 88: 7266-7270, 1991
cathepsin β -like gene	Cejudo et al, Plant Mol Biol 20: 849-856, 1992
Barley Ltp2	Kalla et al., Plant J. 6: 849-60, 1994
Chi26	Leah et al., Plant J. 4: 579-89, 1994
Maize B-Peru	Selinger et al., Genetics 149: 1125-38, 1998

A green tissue-specific promoter as defined herein is a promoter that is transcriptionally active predominantly in green tissue, substantially to the exclusion of any other parts of a plant, whilst still allowing for any leaky expression in these other plant parts.

Examples of green tissue-specific promoters which may be used to perform the methods of the invention are shown in Table 2g below.

TABLE 2g

Examples of green tissue-specific promoters		
Gene	Expression	Reference
Maize Orthophosphate dikinase	Leaf specific	Fukavama et al., 2001
Maize Phosphoenolpyruvate carboxylase	Leaf specific	Kausch et al., 2001
Rice Phosphoenolpyruvate carboxylase	Leaf specific	Liu et al., 2003
Rice small subunit Rubisco	Leaf specific	Nomura et al., 2000
rice beta expansin EXBP9	Shoot specific	WO 2004/070039
Pigeonpea small subunit Rubisco	Leaf specific	Panguluri et al., 2005
Pea RBCS3A	Leaf specific	

Another example of a tissue-specific promoter is a meristem-specific promoter, which is transcriptionally active predominantly in meristematic tissue, substantially to the exclusion of any other parts of a plant, whilst still allowing for any leaky expression in these other plant parts. Examples of green meristem-specific promoters which may be used to perform the methods of the invention are shown in Table 2h below.

TABLE 2h

Examples of meristem-specific promoters		
Gene source	Expression pattern	Reference
rice OSH1	Shoot apical meristem, from embryo globular stage to seedling stage	Sato et al. (1996) Proc. Natl. Acad. Sci. USA, 93: 8117-8122
Rice metallothionein	Meristem specific	BAD87835.1
WAK1 & WAK2	Shoot and root apical meristems, and in expanding leaves and sepals	Wagner & Kohorn (2001) Plant Cell 13(2): 303-318

Terminator

The term "terminator" encompasses a control sequence which is a DNA sequence at the end of a transcriptional unit which signals 3' processing and polyadenylation of a primary transcript and termination of transcription. The terminator can be derived from the natural gene, from a variety of other plant genes, or from T-DNA. The terminator to be added may be derived from, for example, the nopaline synthase or octopine synthase genes, or alternatively from another plant gene, or less preferably from any other eukaryotic gene.

Modulation

The term "modulation" means in relation to expression or gene expression, a process in which the expression level is changed by said gene expression in comparison to the control plant, the expression level may be increased or decreased. The original, unmodulated expression may be of any kind of expression of a structural RNA (rRNA, tRNA) or mRNA with subsequent translation. The term "modulating the activity" shall mean any change of the expression of the inventive nucleic acid sequences or encoded proteins, which leads to increased yield and/or increased growth of the plants.

Expression

The term "expression" or "gene expression" means the transcription of a specific gene or specific genes or specific genetic construct. The term "expression" or "gene expression" in particular means the transcription of a gene or genes or genetic construct into structural RNA (rRNA, tRNA) or mRNA with or without subsequent translation of the latter into a protein. The process includes transcription of DNA and processing of the resulting mRNA product.

Increased Expression/Overexpression

The term "increased expression" or "overexpression" as used herein means any form of expression that is additional to the original wild-type expression level.

Methods for increasing expression of genes or gene products are well documented in the art and include, for example, overexpression driven by appropriate promoters, the use of transcription enhancers or translation enhancers. Isolated nucleic acids which serve as promoter or enhancer elements may be introduced in an appropriate position (typically upstream) of a non-heterologous form of a polynucleotide so as to upregulate expression of a nucleic acid encoding the polypeptide of interest. For example, endogenous promoters may be altered in vivo by mutation, deletion, and/or substitution (see, Kmiec, U.S. Pat. No. 5,565,350; Zarling et al., WO9322443), or isolated promoters may be introduced into a plant cell in the proper orientation and distance from a gene of the present invention so as to control the expression of the gene.

If polypeptide expression is desired, it is generally desirable to include a polyadenylation region at the 3'-end of a polynucleotide coding region. The polyadenylation region can be derived from the natural gene, from a variety of other plant genes, or from T-DNA. The 3' end sequence to be added may be derived from, for example, the nopaline synthase or octopine synthase genes, or alternatively from another plant gene, or less preferably from any other eukaryotic gene.

An intron sequence may also be added to the 5' untranslated region (UTR) or the coding sequence of the partial coding sequence to increase the amount of the mature message that accumulates in the cytosol. Inclusion of a spliceable intron in the transcription unit in both plant and animal expression constructs has been shown to increase gene expression at both the mRNA and protein levels up to 1000-fold (Buchman and Berg (1988) Mol. Cell. Biol. 8: 4395-4405; Callis et al. (1987) Genes Dev 1:1183-1200). Such intron enhancement of gene expression is typically greatest

when placed near the 5' end of the transcription unit. Use of the maize introns Adh1-S intron 1, 2, and 6, the Bronze-1 intron are known in the art. For general information see: The Maize Handbook, Chapter 116, Freeling and Walbot, Eds., Springer, N.Y. (1994).

Endogenous Gene

Reference herein to an "endogenous" gene not only refers to the gene in question as found in a plant in its natural form (i.e., without there being any human intervention), but also refers to that same gene (or a substantially homologous nucleic acid/gene) in an isolated form subsequently (re)introduced into a plant (a transgene). For example, a transgenic plant containing such a transgene may encounter a substantial reduction of the transgene expression and/or substantial reduction of expression of the endogenous gene. The isolated gene may be isolated from an organism or may be manmade, for example by chemical synthesis.

Decreased Expression

Reference herein to "decreased expression" or "reduction or substantial elimination" of expression is taken to mean a decrease in endogenous gene expression and/or polypeptide levels and/or polypeptide activity relative to control plants. The reduction or substantial elimination is in increasing order of preference at least 10%, 20%, 30%, 40% or 50%, 60%, 70%, 80%, 85%, 90%, or 95%, 96%, 97%, 98%, 99% or more reduced compared to that of control plants. Methods for decreasing expression are known in the art and the skilled person would readily be able to adapt the known methods for silencing so as to achieve reduction of expression of an endogenous gene in a whole plant or in parts thereof through the use of an appropriate promoter, for example.

For the reduction or substantial elimination of expression an endogenous gene in a plant, a sufficient length of substantially contiguous nucleotides of a nucleic acid sequence is required. In order to perform gene silencing, this may be as little as 20, 19, 18, 17, 16, 15, 14, 13, 12, 11, 10 or fewer nucleotides, alternatively this may be as much as the entire gene (including the 5' and/or 3' UTR, either in part or in whole). The stretch of substantially contiguous nucleotides may be derived from the nucleic acid encoding the protein of interest (target gene), or from any nucleic acid capable of encoding an orthologue, paralogue or homologue of the protein of interest. Preferably, the stretch of substantially contiguous nucleotides is capable of forming hydrogen bonds with the target gene (either sense or antisense strand), more preferably, the stretch of substantially contiguous nucleotides has, in increasing order of preference, 50%, 60%, 70%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99%, 100% sequence identity to the target gene (either sense or antisense strand). A nucleic acid sequence encoding a (functional) polypeptide is not a requirement for the various methods discussed herein for the reduction or substantial elimination of expression of an endogenous gene.

Examples of various methods for the reduction or substantial elimination of expression in a plant of an endogenous gene, or for lowering levels and/or activity of a protein, are known to the skilled in the art. A skilled person would readily be able to adapt the known methods for silencing, so as to achieve reduction of expression of an endogenous gene in a whole plant or in parts thereof through the use of an appropriate promoter, for example.

This reduction or substantial elimination of expression may be achieved using routine tools and techniques. A preferred method for the reduction or substantial elimination of endogenous gene expression is by introducing and expressing in a plant a genetic construct into which the nucleic acid (in this case a stretch of substantially contiguous nucleotides

derived from the gene of interest, or from any nucleic acid capable of encoding an orthologue, paralogue or homologue of any one of the protein of interest) is cloned as an inverted repeat (in part or completely), separated by a spacer (non-coding DNA).

In such a preferred method, expression of the endogenous gene is reduced or substantially eliminated through RNA-mediated silencing using an inverted repeat of a nucleic acid or a part thereof (in this case a stretch of substantially contiguous nucleotides derived from the gene of interest, or from any nucleic acid capable of encoding an orthologue, paralogue or homologue of the protein of interest), preferably capable of forming a hairpin structure. The inverted repeat is cloned in an expression vector comprising control sequences. A non-coding DNA nucleic acid sequence (a spacer, for example a matrix attachment region fragment (MAR), an intron, a polylinker, etc.) is located between the two inverted nucleic acids forming the inverted repeat. After transcription of the inverted repeat, a chimeric RNA with a self-complementary structure is formed (partial or complete). This double-stranded RNA structure is referred to as the hairpin RNA (hpRNA). The hpRNA is processed by the plant into siRNAs that are incorporated into an RNA-induced silencing complex (RISC). The RISC further cleaves the mRNA transcripts, thereby substantially reducing the number of mRNA transcripts to be translated into polypeptides. For further general details see for example, Grierson et al. (1998) WO 98/53083; Waterhouse et al. (1999) WO 99/53050).

Performance of the methods of the invention does not rely on introducing and expressing in a plant a genetic construct into which the nucleic acid is cloned as an inverted repeat, but any one or more of several well-known "gene silencing" methods may be used to achieve the same effects.

One such method for the reduction of endogenous gene expression is RNA-mediated silencing of gene expression (downregulation). Silencing in this case is triggered in a plant by a double stranded RNA sequence (dsRNA) that is substantially similar to the target endogenous gene. This dsRNA is further processed by the plant into about 20 to about 26 nucleotides called short interfering RNAs (siRNAs). The siRNAs are incorporated into an RNA-induced silencing complex (RISC) that cleaves the mRNA transcript of the endogenous target gene, thereby substantially reducing the number of mRNA transcripts to be translated into a polypeptide. Preferably, the double stranded RNA sequence corresponds to a target gene.

Another example of an RNA silencing method involves the introduction of nucleic acid sequences or parts thereof (in this case a stretch of substantially contiguous nucleotides derived from the gene of interest, or from any nucleic acid capable of encoding an orthologue, paralogue or homologue of the protein of interest) in a sense orientation into a plant. "Sense orientation" refers to a DNA sequence that is homologous to an mRNA transcript thereof. Introduced into a plant would therefore be at least one copy of the nucleic acid sequence. The additional nucleic acid sequence will reduce expression of the endogenous gene, giving rise to a phenomenon known as co-suppression. The reduction of gene expression will be more pronounced if several additional copies of a nucleic acid sequence are introduced into the plant, as there is a positive correlation between high transcript levels and the triggering of co-suppression.

Another example of an RNA silencing method involves the use of antisense nucleic acid sequences. An "antisense" nucleic acid sequence comprises a nucleotide sequence that is complementary to a "sense" nucleic acid sequence encoding a protein, i.e. complementary to the coding strand of a double-

stranded cDNA molecule or complementary to an mRNA transcript sequence. The antisense nucleic acid sequence is preferably complementary to the endogenous gene to be silenced. The complementarity may be located in the "coding region" and/or in the "non-coding region" of a gene. The term "coding region" refers to a region of the nucleotide sequence comprising codons that are translated into amino acid residues. The term "non-coding region" refers to 5' and 3' sequences that flank the coding region that are transcribed but not translated into amino acids (also referred to as 5' and 3' untranslated regions).

Antisense nucleic acid sequences can be designed according to the rules of Watson and Crick base pairing. The antisense nucleic acid sequence may be complementary to the entire nucleic acid sequence (in this case a stretch of substantially contiguous nucleotides derived from the gene of interest, or from any nucleic acid capable of encoding an orthologue, paralogue or homologue of the protein of interest), but may also be an oligonucleotide that is antisense to only a part of the nucleic acid sequence (including the mRNA 5' and 3' UTR). For example, the antisense oligonucleotide sequence may be complementary to the region surrounding the translation start site of an mRNA transcript encoding a polypeptide. The length of a suitable antisense oligonucleotide sequence is known in the art and may start from about 50, 45, 40, 35, 30, 25, 20, 15 or 10 nucleotides in length or less. An antisense nucleic acid sequence according to the invention may be constructed using chemical synthesis and enzymatic ligation reactions using methods known in the art. For example, an antisense nucleic acid sequence (e.g., an antisense oligonucleotide sequence) may be chemically synthesized using naturally occurring nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase the physical stability of the duplex formed between the antisense and sense nucleic acid sequences, e.g., phosphorothioate derivatives and acridine substituted nucleotides may be used. Examples of modified nucleotides that may be used to generate the antisense nucleic acid sequences are well known in the art. Known nucleotide modifications include methylation, cyclization and 'caps' and substitution of one or more of the naturally occurring nucleotides with an analogue such as inosine. Other modifications of nucleotides are well known in the art.

The antisense nucleic acid sequence can be produced biologically using an expression vector into which a nucleic acid sequence has been subcloned in an antisense orientation (i.e., RNA transcribed from the inserted nucleic acid will be of an antisense orientation to a target nucleic acid of interest). Preferably, production of antisense nucleic acid sequences in plants occurs by means of a stably integrated nucleic acid construct comprising a promoter, an operably linked antisense oligonucleotide, and a terminator.

The nucleic acid molecules used for silencing in the methods of the invention (whether introduced into a plant or generated in situ) hybridize with or bind to mRNA transcripts and/or genomic DNA encoding a polypeptide to thereby inhibit expression of the protein, e.g., by inhibiting transcription and/or translation. The hybridization can be by conventional nucleotide complementarity to form a stable duplex, or, for example, in the case of an antisense nucleic acid sequence which binds to DNA duplexes, through specific interactions in the major groove of the double helix. Antisense nucleic acid sequences may be introduced into a plant by transformation or direct injection at a specific tissue site. Alternatively, antisense nucleic acid sequences can be modified to target selected cells and then administered systemically. For example, for systemic administration, antisense nucleic acid

sequences can be modified such that they specifically bind to receptors or antigens expressed on a selected cell surface, e.g., by linking the antisense nucleic acid sequence to peptides or antibodies which bind to cell surface receptors or antigens. The antisense nucleic acid sequences can also be delivered to cells using the vectors described herein.

According to a further aspect, the antisense nucleic acid sequence is an a-anomeric nucleic acid sequence. An a-anomeric nucleic acid sequence forms specific double-stranded hybrids with complementary RNA in which, contrary to the usual b-units, the strands run parallel to each other (Gaultier et al. (1987) Nucl Ac Res 15: 6625-6641). The antisense nucleic acid sequence may also comprise a 2'-o-methylribonucleotide (Inoue et al. (1987) Nucl Ac Res 15, 6131-6148) or a chimeric RNA-DNA analogue (Inoue et al. (1987) FEBS Lett. 215, 327-330).

The reduction or substantial elimination of endogenous gene expression may also be performed using ribozymes. Ribozymes are catalytic RNA molecules with ribonuclease activity that are capable of cleaving a single-stranded nucleic acid sequence, such as an mRNA, to which they have a complementary region. Thus, ribozymes (e.g., hammerhead ribozymes (described in Haselhoff and Gerlach (1988) Nature 334, 585-591) can be used to catalytically cleave mRNA transcripts encoding a polypeptide, thereby substantially reducing the number of mRNA transcripts to be translated into a polypeptide. A ribozyme having specificity for a nucleic acid sequence can be designed (see for example: Cech et al. U.S. Pat. No. 4,987,071; and Cech et al. U.S. Pat. No. 5,116,742). Alternatively, mRNA transcripts corresponding to a nucleic acid sequence can be used to select a catalytic RNA having a specific ribonuclease activity from a pool of RNA molecules (Bartel and Szostak (1993) Science 261, 1411-1418). The use of ribozymes for gene silencing in plants is known in the art (e.g., Atkins et al. (1994) WO 94/00012; Lenne et al. (1995) WO 95/03404; Lutziger et al. (2000) WO 00/00619; Prinsen et al. (1997) WO 97/13865 and Scott et al. (1997) WO 97/38116).

Gene silencing may also be achieved by insertion mutagenesis (for example, T-DNA insertion or transposon insertion) or by strategies as described by, among others, Angell and Baulcombe ((1999) Plant J 20(3): 357-62), (Amplicon VIGS WO 98/36083), or Baulcombe (WO 99/15682).

Gene silencing may also occur if there is a mutation on an endogenous gene and/or a mutation on an isolated gene/nucleic acid subsequently introduced into a plant. The reduction or substantial elimination may be caused by a non-functional polypeptide. For example, the polypeptide may bind to various interacting proteins; one or more mutation(s) and/or truncation(s) may therefore provide for a polypeptide that is still able to bind interacting proteins (such as receptor proteins) but that cannot exhibit its normal function (such as signalling ligand).

A further approach to gene silencing is by targeting nucleic acid sequences complementary to the regulatory region of the gene (e.g., the promoter and/or enhancers) to form triple helical structures that prevent transcription of the gene in target cells. See Helene, C., Anticancer Drug Res. 6, 569-84, 1991; Helene et al., Ann. N.Y. Acad. Sci. 660, 27-36 1992; and Maher, L. J. Bioassays 14, 807-15, 1992.

Other methods, such as the use of antibodies directed to an endogenous polypeptide for inhibiting its function in plants, or interference in the signalling pathway in which a polypeptide is involved, will be well known to the skilled man. In particular, it can be envisaged that manmade molecules may be useful for inhibiting the biological function of a target

polypeptide, or for interfering with the signalling pathway in which the target polypeptide is involved.

Alternatively, a screening program may be set up to identify in a plant population natural variants of a gene, which variants encode polypeptides with reduced activity. Such natural variants may also be used for example, to perform homologous recombination.

Artificial and/or natural microRNAs (miRNAs) may be used to knock out gene expression and/or mRNA translation. Endogenous miRNAs are single stranded small RNAs of typically 19-24 nucleotides long. They function primarily to regulate gene expression and/or mRNA translation. Most plant microRNAs (miRNAs) have perfect or near-perfect complementarity with their target sequences. However, there are natural targets with up to five mismatches. They are processed from longer non-coding RNAs with characteristic fold-back structures by double-strand specific RNases of the Dicer family. Upon processing, they are incorporated in the RNA-induced silencing complex (RISC) by binding to its main component, an Argonaute protein. mRNAs serve as the specificity components of RISC, since they base-pair to target nucleic acids, mostly mRNAs, in the cytoplasm. Subsequent regulatory events include target mRNA cleavage and destruction and/or translational inhibition. Effects of miRNA over-expression are thus often reflected in decreased mRNA levels of target genes.

Artificial microRNAs (amiRNAs), which are typically 21 nucleotides in length, can be genetically engineered specifically to negatively regulate gene expression of single or multiple genes of interest. Determinants of plant microRNA target selection are well known in the art. Empirical parameters for target recognition have been defined and can be used to aid in the design of specific amiRNAs, (Schwab et al., Dev. Cell 8, 517-527, 2005). Convenient tools for design and generation of amiRNAs and their precursors are also available to the public (Schwab et al., Plant Cell 18, 1121-1133, 2006).

For optimal performance, the gene silencing techniques used for reducing expression in a plant of an endogenous gene requires the use of nucleic acid sequences from monocotyledonous plants for transformation of monocotyledonous plants, and from dicotyledonous plants for transformation of dicotyledonous plants. Preferably, a nucleic acid sequence from any given plant species is introduced into that same species. For example, a nucleic acid sequence from rice is transformed into a rice plant. However, it is not an absolute requirement that the nucleic acid sequence to be introduced originates from the same plant species as the plant in which it will be introduced. It is sufficient that there is substantial homology between the endogenous target gene and the nucleic acid to be introduced.

Described above are examples of various methods for the reduction or substantial elimination of expression in a plant of an endogenous gene. A person skilled in the art would readily be able to adapt the aforementioned methods for silencing so as to achieve reduction of expression of an endogenous gene in a whole plant or in parts thereof through the use of an appropriate promoter, for example.

Selectable Marker (Gene)/Reporter Gene

"Selectable marker", "selectable marker gene" or "reporter gene" includes any gene that confers a phenotype on a cell in which it is expressed to facilitate the identification and/or selection of cells that are transfected or transformed with a nucleic acid construct of the invention. These marker genes enable the identification of a successful transfer of the nucleic acid molecules via a series of different principles. Suitable markers may be selected from markers that confer antibiotic or herbicide resistance, that introduce a new metabolic trait or

that allow visual selection. Examples of selectable marker genes include genes conferring resistance to antibiotics (such as nptII that phosphorylates neomycin and kanamycin, or hpt, phosphorylating hygromycin, or genes conferring resistance to, for example, bleomycin, streptomycin, tetracyclin, chloramphenicol, ampicillin, gentamycin, geneticin (G418), spectinomycin or blasticidin), to herbicides (for example bar which provides resistance to Basta®; aroA or gox providing resistance against glyphosate, or the genes conferring resistance to, for example, imidazolinone, phosphinothricin or sulfonylurea), or genes that provide a metabolic trait (such as manA that allows plants to use mannose as sole carbon source or xylose isomerase for the utilisation of xylose, or antinutritive markers such as the resistance to 2-deoxyglucose). Expression of visual marker genes results in the formation of colour (for example β -glucuronidase, GUS or β -galactosidase with its coloured substrates, for example X-Gal), luminescence (such as the luciferin/luciferase system) or fluorescence (Green Fluorescent Protein, GFP, and derivatives thereof). This list represents only a small number of possible markers. The skilled worker is familiar with such markers. Different markers are preferred, depending on the organism and the selection method.

It is known that upon stable or transient integration of nucleic acids into plant cells, only a minority of the cells takes up the foreign DNA and, if desired, integrates it into its genome, depending on the expression vector used and the transfection technique used. To identify and select these integrants, a gene coding for a selectable marker (such as the ones described above) is usually introduced into the host cells together with the gene of interest. These markers can for example be used in mutants in which these genes are not functional by, for example, deletion by conventional methods. Furthermore, nucleic acid molecules encoding a selectable marker can be introduced into a host cell on the same vector that comprises the sequence encoding the polypeptides of the invention or used in the methods of the invention, or else in a separate vector. Cells which have been stably transfected with the introduced nucleic acid can be identified for example by selection (for example, cells which have integrated the selectable marker survive whereas the other cells die). The marker genes may be removed or excised from the transgenic cell once they are no longer needed. Techniques for marker gene removal are known in the art, useful techniques are described above in the definitions section.

Since the marker genes, particularly genes for resistance to antibiotics and herbicides, are no longer required or are undesired in the transgenic host cell once the nucleic acids have been introduced successfully, the process according to the invention for introducing the nucleic acids advantageously employs techniques which enable the removal or excision of these marker genes. One such method is what is known as co-transformation. The co-transformation method employs two vectors simultaneously for the transformation, one vector bearing the nucleic acid according to the invention and a second bearing the marker gene(s). A large proportion of transformants receives or, in the case of plants, comprises (up to 40% or more of the transformants), both vectors. In case of transformation with *Agrobacteria*, the transformants usually receive only a part of the vector, i.e. the sequence flanked by the T-DNA, which usually represents the expression cassette. The marker genes can subsequently be removed from the transformed plant by performing crosses. In another method, marker genes integrated into a transposon are used for the transformation together with desired nucleic acid (known as the Ac/Ds technology). The transformants can be crossed with a transposase source or the transformants are

transformed with a nucleic acid construct conferring expression of a transposase, transiently or stable. In some cases (approx. 10%), the transposon jumps out of the genome of the host cell once transformation has taken place successfully and is lost. In a further number of cases, the transposon jumps to a different location. In these cases the marker gene must be eliminated by performing crosses. In microbiology, techniques were developed which make possible, or facilitate, the detection of such events. A further advantageous method relies on what is known as recombination systems; whose advantage is that elimination by crossing can be dispensed with. The best-known system of this type is what is known as the Cre/lox system. Cre1 is a recombinase that removes the sequences located between the loxP sequences. If the marker gene is integrated between the loxP sequences, it is removed once transformation has taken place successfully, by expression of the recombinase. Further recombination systems are the HIN/HIX, FLP/FRT and REP/STB system (Tribble et al., J. Biol. Chem., 275, 2000: 22255-22267; Velmurugan et al., J. Cell Biol., 149, 2000: 553-566). A site-specific integration into the plant genome of the nucleic acid sequences according to the invention is possible. Naturally, these methods can also be applied to microorganisms such as yeast, fungi or bacteria. Transgenic/Transgene/Recombinant

For the purposes of the invention, "transgenic", "transgene" or "recombinant" means with regard to, for example, a nucleic acid sequence, an expression cassette, gene construct or a vector comprising the nucleic acid sequence or an organism transformed with the nucleic acid sequences, expression cassettes or vectors according to the invention, all those constructions brought about by recombinant methods in which either

- (a) the nucleic acid sequences encoding proteins useful in the methods of the invention, or
- (b) genetic control sequence(s) which is operably linked with the nucleic acid sequence according to the invention, for example a promoter, or
- (c) a) and b)

are not located in their natural genetic environment or have been modified by recombinant methods, it being possible for the modification to take the form of, for example, a substitution, addition, deletion, inversion or insertion of one or more nucleotide residues. The natural genetic environment is understood as meaning the natural genomic or chromosomal locus in the original plant or the presence in a genomic library. In the case of a genomic library, the natural genetic environment of the nucleic acid sequence is preferably retained, at least in part. The environment flanks the nucleic acid sequence at least on one side and has a sequence length of at least 50 bp, preferably at least 500 bp, especially preferably at least 1000 bp, most preferably at least 5000 bp. A naturally occurring expression cassette—for example the naturally occurring combination of the natural promoter of the nucleic acid sequences with the corresponding nucleic acid sequence encoding a polypeptide useful in the methods of the present invention, as defined above—becomes a transgenic expression cassette when this expression cassette is modified by non-natural, synthetic ("artificial") methods such as, for example, mutagenic treatment. Suitable methods are described, for example, in U.S. Pat. No. 5,565,350 or WO 00/15815.

A transgenic plant for the purposes of the invention is thus understood as meaning, as above, that the nucleic acids used in the method of the invention are not at their natural locus in the genome of said plant, it being possible for the nucleic acids to be expressed homologously or heterologously. However, as mentioned, transgenic also means that, while the

nucleic acids according to the invention or used in the inventive method are at their natural position in the genome of a plant, the sequence has been modified with regard to the natural sequence, and/or that the regulatory sequences of the natural sequences have been modified. Transgenic is preferably understood as meaning the expression of the nucleic acids according to the invention at an unnatural locus in the genome, i.e. homologous or, preferably, heterologous expression of the nucleic acids takes place. Preferred transgenic plants are mentioned herein.

Transformation

The term "introduction" or "transformation" as referred to herein encompasses the transfer of an exogenous polynucleotide into a host cell, irrespective of the method used for transfer. Plant tissue capable of subsequent clonal propagation, whether by organogenesis or embryogenesis, may be transformed with a genetic construct of the present invention and a whole plant regenerated there from. The particular tissue chosen will vary depending on the clonal propagation systems available for, and best suited to, the particular species being transformed. Exemplary tissue targets include leaf disks, pollen, embryos, cotyledons, hypocotyls, megagametophytes, callus tissue, existing meristematic tissue (e.g., apical meristem, axillary buds, and root meristems), and induced meristem tissue (e.g., cotyledon meristem and hypocotyl meristem). The polynucleotide may be transiently or stably introduced into a host cell and may be maintained non-integrated, for example, as a plasmid. Alternatively, it may be integrated into the host genome. The resulting transformed plant cell may then be used to regenerate a transformed plant in a manner known to persons skilled in the art.

The transfer of foreign genes into the genome of a plant is called transformation. Transformation of plant species is now a fairly routine technique. Advantageously, any of several transformation methods may be used to introduce the gene of interest into a suitable ancestor cell. The methods described for the transformation and regeneration of plants from plant tissues or plant cells may be utilized for transient or for stable transformation. Transformation methods include the use of liposomes, electroporation, chemicals that increase free DNA uptake, injection of the DNA directly into the plant, particle gun bombardment, transformation using viruses or pollen and microprojection. Methods may be selected from the calcium/polyethylene glycol method for protoplasts (Krens, F. A. et al., (1982) Nature 296, 72-74; Negrutiu I et al. (1987) Plant Mol Biol 8: 363-373); electroporation of protoplasts (Shillito R. D. et al. (1985) Bio/Technol 3, 1099-1102); microinjection into plant material (Crossway A et al., (1986) Mol. Gen. Genet. 202: 179-185); DNA or RNA-coated particle bombardment (Klein T M et al., (1987) Nature 327: 70) infection with (non-integrative) viruses and the like. Transgenic plants, including transgenic crop plants, are preferably produced via *Agrobacterium*-mediated transformation. An advantageous transformation method is the transformation in planta. To this end, it is possible, for example, to allow the *agrobacteria* to act on plant seeds or to inoculate the plant meristem with *agrobacteria*. It has proved particularly expedient in accordance with the invention to allow a suspension of transformed *agrobacteria* to act on the intact plant or at least on the flower primordia. The plant is subsequently grown on until the seeds of the treated plant are obtained (Clough and Bent, Plant J. (1998) 16, 735-743). Methods for *Agrobacterium*-mediated transformation of rice include well known methods for rice transformation, such as those described in any of the following: European patent application EP 1198985 A1, Aldemita and Hodges (Planta 199: 612-617, 1996); Chan et al. (Plant Mol Biol 22 (3): 491-506,

1993), Hiei et al. (Plant J 6 (2): 271-282, 1994), which disclosures are incorporated by reference herein as if fully set forth. In the case of corn transformation, the preferred method is as described in either Ishida et al. (Nat. Biotechnol 14(6): 745-50, 1996) or Frame et al. (Plant Physiol 129(1): 13-22, 2002), which disclosures are incorporated by reference herein as if fully set forth. Said methods are further described by way of example in B. Jené et al., Techniques for Gene Transfer, in: Transgenic Plants, Vol. 1, Engineering and Utilization, eds. S. D. Kung and R. Wu, Academic Press (1993) 128-143 and in Potrykus Annu. Rev. Plant Physiol. Plant Molec. Biol. 42 (1991) 205-225). The nucleic acids or the construct to be expressed is preferably cloned into a vector, which is suitable for transforming *Agrobacterium tumefaciens*, for example pBin19 (Bevan et al., Nucl. Acids Res. 12 (1984) 8711). *Agrobacterium* transformed by such a vector can then be used in known manner for the transformation of plants, such as plants used as a model, like *Arabidopsis* (*Arabidopsis thaliana* is within the scope of the present invention not considered as a crop plant), or crop plants such as, by way of example, tobacco plants, for example by immersing bruised leaves or chopped leaves in an agrobacterial solution and then culturing them in suitable media. The transformation of plants by means of *Agrobacterium tumefaciens* is described, for example, by Höfgen and Willmitzer in Nucl. Acid Res. (1988) 16, 9877 or is known inter alia from F. F. White, Vectors for Gene Transfer in Higher Plants; in Transgenic Plants, Vol. 1, Engineering and Utilization, eds. S. D. Kung and R. Wu, Academic Press, 1993, pp. 15-38.

In addition to the transformation of somatic cells, which then have to be regenerated into intact plants, it is also possible to transform the cells of plant meristems and in particular those cells which develop into gametes. In this case, the transformed gametes follow the natural plant development, giving rise to transgenic plants. Thus, for example, seeds of *Arabidopsis* are treated with *agrobacteria* and seeds are obtained from the developing plants of which a certain proportion is transformed and thus transgenic [Feldman, K A and Marks M D (1987). Mol Gen Genet. 208:274-289; Feldmann K (1992). In: C Koncz, N-H Chua and J Shell, eds, Methods in *Arabidopsis* Research. Word Scientific, Singapore, pp. 274-289]. Alternative methods are based on the repeated removal of the inflorescences and incubation of the excision site in the center of the rosette with transformed *agrobacteria*, whereby transformed seeds can likewise be obtained at a later point in time (Chang (1994). Plant J. 5: 551-558; Katavik (1994). Mol Gen Genet, 245: 363-370). However, an especially effective method is the vacuum infiltration method with its modifications such as the "floral dip" method. In the case of vacuum infiltration of *Arabidopsis*, intact plants under reduced pressure are treated with an agrobacterial suspension [Bechthold, N (1993). C R Acad Sci Paris Life Sci, 316: 1194-1199], while in the case of the "floral dip" method the developing floral tissue is incubated briefly with a surfactant-treated agrobacterial suspension [Clough, S J and Bent A F (1998) The Plant J. 16, 735-743]. A certain proportion of transgenic seeds are harvested in both cases, and these seeds can be distinguished from non-transgenic seeds by growing under the above-described selective conditions. In addition the stable transformation of plastids is of advantages because plastids are inherited maternally in most crops reducing or eliminating the risk of transgene flow through pollen. The transformation of the chloroplast genome is generally achieved by a process which has been schematically displayed in Klaus et al., 2004 [Nature Biotechnology 22 (2), 225-229]. Briefly the sequences to be transformed are cloned together with a selectable marker gene between flanking

sequences homologous to the chloroplast genome. These homologous flanking sequences direct site specific integration into the plastome. Plastid transformation has been described for many different plant species and an overview is given in Bock (2001) Transgenic plastids in basic research and plant biotechnology. J Mol. Biol. 2001 Sep. 21; 312 (3):425-38 or Maliga, P (2003) Progress towards commercialization of plastid transformation technology. Trends Biotechnol. 21, 20-28. Further biotechnological progress has recently been reported in form of marker free plastid transformants, which can be produced by a transient co-integrated maker gene (Klaus et al., 2004, Nature Biotechnology 22(2), 225-229).

T-DNA Activation Tagging

T-DNA activation tagging (Hayashi et al. Science (1992) 1350-1353), involves insertion of T-DNA, usually containing a promoter (may also be a translation enhancer or an intron), in the genomic region of the gene of interest or 10 kb up- or downstream of the coding region of a gene in a configuration such that the promoter directs expression of the targeted gene. Typically, regulation of expression of the targeted gene by its natural promoter is disrupted and the gene falls under the control of the newly introduced promoter. The promoter is typically embedded in a T-DNA. This T-DNA is randomly inserted into the plant genome, for example, through *Agrobacterium* infection and leads to modified expression of genes near the inserted T-DNA. The resulting transgenic plants show dominant phenotypes due to modified expression of genes close to the introduced promoter.

TILLING

The term "TILLING" is an abbreviation of "Targeted Induced Local Lesions In Genomes" and refers to a mutagenesis technology useful to generate and/or identify nucleic acids encoding proteins with modified expression and/or activity. TILLING also allows selection of plants carrying such mutant variants. These mutant variants may exhibit modified expression, either in strength or in location or in timing (if the mutations affect the promoter for example). These mutant variants may exhibit higher activity than that exhibited by the gene in its natural form. TILLING combines high-density mutagenesis with high-throughput screening methods. The steps typically followed in TILLING are: (a) EMS mutagenesis (Redei G P and Koncz C (1992) In Methods in *Arabidopsis* Research, Koncz C, Chua N H, Schell J, eds. Singapore, World Scientific Publishing Co, pp. 16-82; Feldmann et al., (1994) In Meyerowitz E M, Somerville C R, eds, *Arabidopsis*. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., pp 137-172; Lightner J and Caspar T (1998) In J Martinez-Zapater, J Salinas, eds, Methods on Molecular Biology, Vol. 82. Humana Press, Totowa, N.J., pp 91-104); (b) DNA preparation and pooling of individuals; (c) PCR amplification of a region of interest; (d) denaturation and annealing to allow formation of heteroduplexes; (e) DHPLC, where the presence of a heteroduplex in a pool is detected as an extra peak in the chromatogram; (f) identification of the mutant individual; and (g) sequencing of the mutant PCR product. Methods for TILLING are well known in the art (McCallum et al., (2000) Nat Biotechnol 18: 455-457; reviewed by Stemple (2004) Nat Rev Genet. 5(2): 145-50).

Homologous Recombination

Homologous recombination allows introduction in a genome of a selected nucleic acid at a defined selected position. Homologous recombination is a standard technology used routinely in biological sciences for lower organisms such as yeast or the moss *Physcomitrella*. Methods for performing homologous recombination in plants have been described not only for model plants (Offringa et al. (1990)

EMBO J. 9(10): 3077-84) but also for crop plants, for example rice (Terada et al. (2002) Nat Biotech 20(10): 1030-4; Iida and Terada (2004) Curr Opin Biotech 15(2): 132-8), and approaches exist that are generally applicable regardless of the target organism (Miller et al, Nature Biotechnol. 25, 778-785, 2007).

Yield

The term "yield" in general means a measurable produce of economic value, typically related to a specified crop, to an area, and to a period of time. Individual plant parts directly contribute to yield based on their number, size and/or weight, or the actual yield is the yield per square meter for a crop and year, which is determined by dividing total production (includes both harvested and appraised production) by planted square meters. The term "yield" of a plant may relate to vegetative biomass (root and/or shoot biomass), to reproductive organs, and/or to propagules (such as seeds) of that plant.

Early Vigour

"Early vigour" refers to active healthy well-balanced growth especially during early stages of plant growth, and may result from increased plant fitness due to, for example, the plants being better adapted to their environment (i.e. optimizing the use of energy resources and partitioning between shoot and root). Plants having early vigour also show increased seedling survival and a better establishment of the crop, which often results in highly uniform fields (with the crop growing in uniform manner, i.e. with the majority of plants reaching the various stages of development at substantially the same time), and often better and higher yield. Therefore, early vigour may be determined by measuring various factors, such as thousand kernel weight, percentage germination, percentage emergence, seedling growth, seedling height, root length, root and shoot biomass and many more.

Increase/Improve/Enhance

The terms "increase", "improve" or "enhance" are interchangeable and shall mean in the sense of the application at least a 3%, 4%, 5%, 6%, 7%, 8%, 9% or 10%, preferably at least 15% or 20%, more preferably 25%, 30%, 35% or 40% more yield and/or growth in comparison to control plants as defined herein.

Seed Yield

Increased seed yield may manifest itself as one or more of the following: a) an increase in seed biomass (total seed weight) which may be on an individual seed basis and/or per plant and/or per square meter; b) increased number of flowers per plant; c) increased number of (filled) seeds; d) increased seed filling rate (which is expressed as the ratio between the number of filled seeds divided by the total number of seeds); e) increased harvest index, which is expressed as a ratio of the yield of harvestable parts, such as seeds, divided by the total biomass; and f) increased thousand kernel weight (TKW), and g) increased number of primary panicles, which is extrapolated from the number of filled seeds counted and their total weight. An increased TKW may result from an increased seed size and/or seed weight, and may also result from an increase in embryo and/or endosperm size.

An increase in seed yield may also be manifested as an increase in seed size and/or seed volume. Furthermore, an increase in seed yield may also manifest itself as an increase in seed area and/or seed length and/or seed width and/or seed perimeter. Increased seed yield may also result in modified architecture, or may occur because of modified architecture.

Greenness Index

The "greenness index" as used herein is calculated from digital images of plants. For each pixel belonging to the plant object on the image, the ratio of the green value versus the red value (in the RGB model for encoding color) is calculated.

The greenness index is expressed as the percentage of pixels for which the green-to-red ratio exceeds a given threshold. Under normal growth conditions, under salt stress growth conditions, and under reduced nutrient availability growth conditions, the greenness index of plants is measured in the last imaging before flowering. In contrast, under drought stress growth conditions, the greenness index of plants is measured in the first imaging after drought.

Plant

The term "plant" as used herein encompasses whole plants, ancestors and progeny of the plants and plant parts, including seeds, shoots, stems, leaves, roots (including tubers), flowers, and tissues and organs, wherein each of the aforementioned comprise the gene/nucleic acid of interest. The term "plant" also encompasses plant cells, suspension cultures, callus tissue, embryos, meristematic regions, gametophytes, sporophytes, pollen and microspores, again wherein each of the aforementioned comprises the gene/nucleic acid of interest.

Plants that are particularly useful in the methods of the invention include all plants which belong to the superfamily Viridiplantae, in particular monocotyledonous and dicotyledonous plants including fodder or forage legumes, ornamental plants, food crops, trees or shrubs selected from the list comprising *Acer* spp., *Actinidia* spp., *Abelmoschus* spp., *Agave sisalana*, *Agropyron* spp., *Agrostis stolonifera*, *Allium* spp., *Amaranthus* spp., *Ammophila arenaria*, *Ananas comosus*, *Annona* spp., *Apium graveolens*, *Arachis* spp., *Artocarpus* spp., *Asparagus officinalis*, *Avena* spp. (e.g. *Avena sativa*, *Avena fatua*, *Avena byzantina*, *Avena fatua* var. *sativa*, *Avena hybrida*), *Averrhoa carambola*, *Bambusa* sp., *Benincasa hispida*, *Bertholletia excelsa*, *Beta vulgaris*, *Brassica* spp. (e.g. *Brassica napus*, *Brassica rapa* ssp. [canola, oilseed rape, turnip rape]), *Cadaba farinosa*, *Camellia sinensis*, *Canna indica*, *Cannabis sativa*, *Capsicum* spp., *Carex elata*, *Carica papaya*, *Carissa macrocarpa*, *Carya* spp., *Carthamus tinctorius*, *Castanea* spp., *Ceiba pentandra*, *Cichorium endivia*, *Cinnamomum* spp., *Citrullus lanatus*, *Citrus* spp., *Cocos* spp., *Coffea* spp., *Colocasia esculenta*, *Cola* spp., *Corchorus* spp., *Coriandrum sativum*, *Corylus* spp., *Crataegus* spp., *Crocus sativus*, *Cucurbita* spp., *Cucumis* spp., *Cynara* spp., *Daucus carota*, *Desmodium* spp., *Dimocarpus longan*, *Dioscorea* spp., *Diospyros* spp., *Echinochloa* spp., *Elaeis* (e.g. *Elaeis guineensis*, *Elaeis oleifera*), *Eleusine coracana*, *Eragrostis tef*, *Erianthus* sp., *Eriobotrya japonica*, *Eucalyptus* sp., *Eugenia uniflora*, *Fagopyrum* spp., *Fagus* spp., *Festuca arundinacea*, *Ficus carica*, *Fortunella* spp., *Fragaria* spp., *Ginkgo biloba*, *Glycine* spp. (e.g. *Glycine max*, *Soja hispida* or *Soja max*), *Gossypium hirsutum*, *Helianthus* spp. (e.g. *Helianthus annuus*), *Hemerocallis fulva*, *Hibiscus* spp., *Hordeum* spp. (e.g. *Hordeum vulgare*), *Ipomoea batatas*, *Juglans* spp., *Lactuca sativa*, *Lathyrus* spp., *Lens culinaris*, *Linum usitatissimum*, *Litchi chinensis*, *Lotus* spp., *Luffa acutangula*, *Lupinus* spp., *Luzula sylvatica*, *Lycopersicon* spp. (e.g. *Lycopersicon esculentum*, *Lycopersicon lycopersicum*, *Lycopersicon pyriforme*), *Macrotyloma* spp., *Malus* spp., *Malpighia emarginata*, *Mammea americana*, *Mangifera indica*, *Manihot* spp., *Manilkara zapota*, *Medicago sativa*, *Melilotus* spp., *Mentha* spp., *Miscanthus sinensis*, *Momordica* spp., *Morus nigra*, *Musa* spp., *Nicotiana* spp., *Olea* spp., *Opuntia* spp., *Ornithopus* spp., *Oryza* spp. (e.g. *Oryza sativa*, *Oryza latifolia*), *Panicum miliaceum*, *Panicum virgatum*, *Passiflora edulis*, *Pastinaca sativa*, *Pennisetum* sp., *Persea* spp., *Petroselinum crispum*, *Phalaris arundinacea*, *Phaseolus* spp., *Phleum pratense*, *Phoenix* spp., *Phragmites australis*, *Physalis* spp., *Pinus* spp., *Pistacia vera*, *Pisum* spp., *Poa* spp., *Populus* spp., *Prosopis* spp., *Prunus* spp., *Psidium* spp., *Punica granatum*, *Pyrus communis*, *Quercus* spp., *Raphanus sativus*, *Rheum*

rhabarbarum, *Ribes* spp., *Ricinus communis*, *Rubus* spp., *Saccharum* spp., *Salix* sp., *Sambucus* spp., *Secale cereale*, *Sesamum* spp., *Sinapis* sp., *Solanum* spp. (e.g. *Solanum tuberosum*, *Solanum integrifolium* or *Solanum lycopersicum*), *Sorghum bicolor*, *Spinacia* spp., *Syzygium* spp., *Tagetes* spp., *Tamarindus indica*, *Theobroma cacao*, *Trifolium* spp., *Tripsacum dactyloides*, *Triticale* sp., *Triticosecale rimpaii*, *Triticum* spp. (e.g. *Triticum aestivum*, *Triticum durum*, *Triticum turgidum*, *Triticum hybernum*, *Triticum macha*, *Triticum sativum*, *Triticum monococcum* or *Triticum vulgare*), *Tropaeolum minus*, *Tropaeolum majus*, *Vaccinium* spp., *Vicia* spp., *Vigna* spp., *Viola odorata*, *Vitis* spp., *Zea mays*, *Zizania palustris*, *Ziziphus* spp., amongst others.

DETAILED DESCRIPTION OF THE INVENTION

Surprisingly, it has now been found that modulating expression in a plant of a nucleic acid encoding an ASPAT polypeptide gives plants having enhanced yield-related traits relative to control plants. According to a first embodiment, the present invention provides a method for enhancing yield-related traits in plants relative to control plants, comprising modulating expression in a plant of a nucleic acid encoding an ASPAT polypeptide and optionally selecting for plants having enhanced yield-related traits.

Furthermore surprisingly, it has now been found that increasing expression in a plant of a nucleic acid sequence encoding an MYB91 polypeptide as defined herein, gives plants having increased yield-related traits relative to control plants. According to a further embodiment, the present invention provides a method for increasing yield-related traits in plants relative to control plants, comprising increasing expression in a plant of a nucleic acid sequence encoding an MYB91 polypeptide.

Even furthermore surprisingly, it has now been found that modulating expression in a plant of a nucleic acid encoding a GASA polypeptide gives plants having enhanced yield-related traits relative to control plants. According to a further embodiment, the present invention provides a method for enhancing yield-related traits in plants relative to control plants, comprising modulating expression in a plant of a nucleic acid encoding a GASA polypeptide.

Yet furthermore surprisingly, it has now been found that modulating expression in a plant of a nucleic acid encoding an AUX/IAA polypeptide gives plants having enhanced yield-related traits relative to control plants. According to a first embodiment, the present invention provides a method for enhancing yield-related traits in plants relative to control plants, comprising modulating expression in a plant of a nucleic acid encoding an AUX/IAA polypeptide and wherein the yield related traits do not encompass increased root growth.

Concerning ASPAT polypeptides, a preferred method for modulating (preferably, increasing) expression of a nucleic acid encoding an ASPAT polypeptide (ASPAT nucleic acid) is by introducing and expressing in a plant a nucleic acid encoding an ASPAT polypeptide. Preferably the increased expression of the ASPAT nucleic acid and/or the of the ASPAT polypeptide and/or ASPAT activity occurs in one or more subcellular compartments selected in increasing order of preference from the cytosol, the chloroplast, the peroxisomes, the glyoxisomes and the mitochondria of a plant cell.

Cytosolic levels of the ASPAT nucleic acid expression levels and/or ASPAT polypeptide and/or ASPAT activity may be increased for example by expressing an ASPAT nucleic acid encoding a cytosolic isoform. Alternatively, ASPAT nucleic acids encoding isoforms naturally expressed in an

organelle of the plant cell may be expressed in the cytosol by removing the specific organelle targeting motifs. Similarly a naturally found cytosolic isoform may be expressed in a preferred organelle by fusing specific acid amino acid motifs encoding known specific subcellular targeting signals of such organelle. Tools and techniques to express a polypeptide in a preferred organelle of a plant cell are well known in the art.

Concerning MYB91 polypeptides, a preferred method for increasing expression in a plant of a nucleic acid sequence encoding an MYB91 polypeptide is by introducing and expressing in a plant a nucleic acid sequence encoding an MYB91 polypeptide.

Concerning GASA polypeptides, a preferred method for modulating (preferably, increasing) expression of a nucleic acid encoding a GASA polypeptide is by introducing and expressing in a plant a nucleic acid encoding a GASA polypeptide.

Concerning AUX/IAA polypeptides, a preferred method for modulating (preferably, increasing) expression of a nucleic acid encoding an AUX/IAA polypeptide is by introducing and expressing in a plant a nucleic acid encoding an AUX/IAA polypeptide.

Concerning ASPAT polypeptides, any reference hereinafter to a "protein useful in the methods of the invention" is taken to mean an ASPAT polypeptide as defined herein. Any reference hereinafter to a "nucleic acid useful in the methods of the invention" is taken to mean a nucleic acid capable of encoding such an ASPAT polypeptide. The nucleic acid to be introduced into a plant (and therefore useful in performing the methods of the invention) is any nucleic acid encoding the type of protein which will now be described, hereafter also named "ASPAT nucleic acid" or "ASPAT gene".

Concerning MYB91 polypeptides, any reference hereinafter to a "protein useful in the methods of the invention" is taken to mean an MYB91 polypeptide as defined herein. Any reference hereinafter to a "nucleic acid sequence useful in the methods of the invention" is taken to mean a nucleic acid sequence capable of encoding such an MYB91 polypeptide. The nucleic acid sequence to be introduced into a plant (and therefore useful in performing the methods of the invention) is any nucleic acid sequence encoding the type of polypeptide, which will now be described, hereafter also named "MYB91 nucleic acid sequence" or "MYB91 gene".

Concerning GASA polypeptides, any reference hereinafter to a "protein useful in the methods of the invention" is taken to mean a GASA polypeptide as defined herein. Any reference hereinafter to a "nucleic acid useful in the methods of the invention" is taken to mean a nucleic acid capable of encoding such a GASA polypeptide. The nucleic acid to be introduced into a plant (and therefore useful in performing the methods of the invention) is any nucleic acid encoding the type of protein which will now be described, hereafter also named "GASA nucleic acid" or "GASA gene".

Concerning AUX/IAA polypeptides, any reference hereinafter to a "protein (or polypeptide) useful in the methods of the invention" is taken to mean an AUX/IAA polypeptide as defined herein. Any reference hereinafter to a "nucleic acid useful in the methods of the invention" is taken to mean a nucleic acid capable of encoding such an AUX/IAA polypeptide. The nucleic acid to be introduced into a plant (and therefore useful in performing the methods of the invention) is any nucleic acid encoding the type of protein which will now be described, hereafter also named "AUX/IAA nucleic acid" or "AUX/IAA gene".

An "ASPAT polypeptide" as defined herein refers to any polypeptide comprising an Aminotransferase, class I and II (Aminotran_1_2) domain (Interpro accession number:

IPR004839; pfam accession number: PF00155), and optionally Aspartate Transaminase activity (EC. 2.6.1.1).

Preferably, an ASPAT polypeptide comprises an Aminotran_1_2 domain having in increasing order of preference at least 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or 100% sequence identity to any of the Aminotran_1_2 domains as set forth in Tables D1, Table D2 and Table D3.

Preferably the ASPAT polypeptide comprises a motif having at least 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or 100% to any one or more of the following motif:

- (i) Motif 1 (SEQ ID NO: 207): NPTG;
- (ii) Motif 2 (SEQ ID NO: 208): IVLLHCAHNPTGVDP;
- (iii) Motif 3 (SEQ ID NO: 209): SRLLLCSPSNPTGSVY;

wherein any amino acid maybe substituted by a conserved amino acid.

Preferably, the homologue of an ASPAT polypeptide has in increasing order of preference at least 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or 100% overall sequence identity to the amino acid of any of the polypeptides of Table A1, preferably to any of the polypeptides in phylogenetic class 1 of Table B1, more preferably to SEQ ID NO: 2, even more preferably to SEQ ID NO: 8, most preferably to SEQ ID NO: 6. In addition the homologue of an ASPAT protein preferably comprises an Aminotran_1_2 domain as described above. The sequence identity is determined using a global alignment algorithm, such as the Needleman Wunsch algorithm in the program GAP (GCG Wisconsin Package, Accelrys), preferably with default parameters and preferably with sequences of mature proteins (i.e. without taking into account secretion signals or transit peptides). Compared to overall sequence identity, the sequence identity will generally be higher when only conserved domains or motifs are considered.

Alternatively, an ASPAT polypeptide useful in the methods of the invention has an amino acid sequence which when used in the construction of a phylogenetic tree, such as the one depicted in FIG. 2 clusters in increasing order of preference with any of the polypeptides of phylogenetic class 1, class 2, class 3 and class 4 as set forth in table B1.

A "MYB91 polypeptide" as defined herein refers to any polypeptide comprising (i) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a MYB DNA binding domain with an InterPro accession number IPR014778, as represented by SEQ ID NO: 269; and (ii) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a MYB DNA binding domain

with an InterPro accession number IPR014778, as represented by SEQ ID NO: 270; and (iii) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a Conserved Domain as represented by SEQ ID NO: 271.

Alternatively or additionally, a "MYB91 polypeptide" as defined herein refers to any polypeptide sequence having in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a polypeptide as represented by SEQ ID NO: 221.

Alternatively or additionally, a "MYB91 polypeptide" as defined herein refers to any polypeptide having in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to any of the polypeptide sequences given in Table A2 herein.

Alternatively or additionally, a "MYB91 polypeptide" as defined herein refers to any polypeptide sequence which when used in the construction of a phylogenetic tree of MYB polypeptides, such as the one depicted in FIG. 4, clusters with the MYB91 group of polypeptides rather than with any other group.

A "GASA polypeptide" as defined herein refers to polypeptides comprising in their native form a secretion signal, the GASA domain PF02704 (Interpro IPR003854) and the following three motifs:

Motif 4, (SEQ ID NO: 277) comprising 4 conserved Cys residues:
CXXCCXXCX

Wherein X in position 2 can be any amino acid, but preferably one of N, K, M, G, L, I, Q; and wherein X in position 3 can be any amino acid, but preferably one of V, T, S, M, I, L, H, Y, K; and wherein X in position 6 can be any amino acid, but preferably one of Q, A, N, D, L, V, R, H, S, G, K, E, T; and wherein X in position 7 can be any amino acid, but preferably one of R, T, A, D, K, E, Q, S, W, C; and wherein X in position 9 can be any amino acid, but preferably one of N, K, R, H, S, G, A, Q, L, D.

Motif 5 (SEQ ID NO: 278):
CV(P/L)(P/K/Q/A/S/T)GXX(Q/G/A/S)

Wherein X in position 6 can be any amino acid, but preferably one of T, P, S, Y, V, N, F, L; and wherein X in position 7 can be any amino acid, but preferably one of G, Y, F, S, A, L, V.

Motifs 4 and 5 are adjacent to each other or are separated from each other by 1 amino acid.

Motif 6 (SEQ ID NO: 279):
CY(D/A/T/F/R/N)X(M/L/W/K)

Wherein X in position 4 can be any amino acid, but preferably one of Q, R, S, D, E, N, T, H.

However, the term GASA polypeptide as used in the present invention does not encompass GASA4 from *Arabidopsis thaliana* (SEQ ID NO: 295).

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Preferably, the GASA polypeptide useful in the methods of the present invention comprises one or more of the following motifs:

Motif 7 (SEQ ID NO: 280):
(S/L/Y/K/S/A)C(G/K/M/I/N/L)(L/M/I/V/T/S)CCXXC
(N/G/A/K/R/H/S/D)

Wherein X on position 7 can be any amino acid, but preferably one of E, H, G, K, A, Q, S, R, T, N, D, L, V; and wherein X on position 8 can be any amino acid, but preferably one of E, D, K, Q, S, R, A, T, C.

Motif 8 (SEQ ID NO: 281):
CVP(T/S/P/A/K/Q)G(S/P/T)(G/Y/L/A/S/F)(S/A/G/Q)
(T/S/P/N/D)(R/K/T/Y/L/Q/E)(D/S/H/R/E/N)X(C/I)

Wherein X in position 12 can be any amino acid, but preferably one of E, H, T, A, S, L, V, K, M.

Preferably, motif 7 is immediately followed by motif 8 or is separated by 1 amino acid from motif 8.

Motif 9 (SEQ ID NO: 282):
(P/R/K/T)CY(R/D/T/F/A)(D/Q/R/N/S/T/H/E)(M/K/W/L)
(L/V/K/R/T/N/I)

Preferably, motif 8 is immediately followed by motif 9 or is separated by 1 amino acid from motif 9.

Motif 10 (SEQ ID NO: 283):
(K/T)(R/P/V/A)C(L/N/M/I)(F/T)(Y/F/L)C(N/L/Q)
(H/Y/K)CC(G/K/E/N/A/R)(W/R/K/T/S/A)C(Q/L/R)CV
(P/L)(P/S/K/A)G(Y/T/V/N/F/L)(V/Y/F)G

Motif 11 (SEQ ID NO: 284):
(N/H)K(G/D/E/Q/A)(C/E/T/S/F/A/V)(C/W)(S/P)CY(N/R)
(N/D)(W/L/M)(K/T/E)(T/K/E/N)(Q/K)

Motif 12 (SEQ ID NO: 285)
(N/R)(G/C)(S/K)(H/Q/A/N/K/G)(K/T)(G/S/Q/A/K)
(H/Y/F)(K/T/R/H)

Alternatively, the homologue of a GASA protein has in increasing order of preference at least 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% overall sequence identity to the amino acid represented by SEQ ID NO: 276, provided that the homologous protein comprises the conserved motifs as outlined above. The overall sequence identity is determined using a global alignment algorithm, such as the Needleman Wunsch algorithm in the program GAP (GCG Wisconsin Package, Accelrys), preferably with default parameters and preferably with sequences of mature proteins (i.e. without taking into account secretion signals or transit peptides). Compared to overall sequence identity, the sequence identity will generally be higher when only conserved domains or motifs are considered.

Preferably, the polypeptide sequence which when used in the construction of a phylogenetic tree, such as the one depicted in FIG. 9, clusters with the group of GASA polypeptides comprising the amino acid sequence represented by

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SEQ ID NO: 276 (or SEQ ID NO: 291 or SEQ ID NO: 292) rather than with any other group. It should be noted that GASA4 from *Arabidopsis thaliana* (SEQ ID NO: 295) is excluded from the group of GASA proteins as defined in the present invention.

An "AUX/IAA polypeptide" as defined herein refers to any polypeptide comprising an AUX/IAA domain (PFAM accession number PF02309, InterPro entry IPR003311). An "AUX/IAA polypeptide" as defined herein does not comprise the motif represented by SEQ ID NO: 670: (K/N)(I/M/L)F(S/Y)(Q/G)L (IAA2 motif).

AUX/IAA polypeptides of the invention have equivalent amino acid structure and function as the AUX/IAA family of transcription factors and homologues thereof.

The structure and function of AUX/IAA domains are well known in the art. Typically they can be found in AUX/IAA transcription factors of plants. Members of the AUX/IAA family of transcription factors from plant origin are well known in the art. A compilation of AUX/IAA polypeptides as found in the viridiplantae kingdom can be found in dedicated databases such as the so called "plant transcription database (PInTFDB)" maintained by the university of Postdam (Germany) and described by Riano-Pacheco et al. BMC Bioinformatics 2007 8:47.

In the PInTFDB database the members of the AUX/IAA family are identified as polypeptides having a AUX/IAA domain (PFAM accession number: PF02309) and not having an Auxin_resp domain (pfam accession number: PF06507); Auxin_resp domains are typically found in ARF polypeptides and typically absent from AUX/IAA polypeptides.

An Example of an AUX/IAA domain as found between amino acid coordinates 5-171 of SEQ ID NO: 432. AUX/IAA domains having sequence similarity to the domain as present in SEQ ID NO: 432 are present in the polypeptides of Table A4.

In a one embodiment of the invention, to perform the methods of the invention there is provided a preferred an AUX/IAA polypeptide, also referred to as IAA14-like polypeptide, which comprises an AUX/IAA domain having at least 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% sequence identity to the amino acid of the AUX/IAA domain represented by the amino acids 1 to 220 in SEQ ID NO: 738 (FIG. 13).

Preferably the IAA14-like polypeptide comprises at least one, and in increasing order of preference, 2, 3, 4, 5, or all six of the following motifs:

Motif 13, SEQ ID NO: 739:
(K/R/E/D)(A/E/D)TEL(C/R)LG(L/I)(P/G)

Motif 14, SEQ ID NO: 740:
KRGF(S/A)ET

Motif 15, SEQ ID NO: 741:
VGWPP(V/I)R

Motif 16, SEQ ID NO: 742:
GAPYLK(V/I)DLXX(Y/F)

wherein X on position 11 can be any amino acid, preferably X on position 11 is one of K, T, R, N, S, or Q and wherein X on position 12 can be any amino acid, preferably X on position 12 is one of N, L, T, N, V, I, or C.

Motif 17, SEQ ID NO: 743:
(S/N/G)(S/W/T)(E/D/G)(Y/F/H)(V/A/E)(P/L/V/I)
(S/T/A)YEDKD(N/G)D(W/L)M(L/F)(V/I)GDVP

-continued

Motif 18, SEQ ID NO: 744:
(S/T)C(K/R/Q)(R/K)(L/I)R(I/L)(M/I)K(G/S/E)(S/K/T)
(E/D)(A/T)

Preferably motif 15 is:
VGWPPVR

Motif 16 is preferably:
GAPYLRK(V/I)DL(K/T/R/N)(M/L)Y

Motif 17 is preferably:
(S/N/G)(S/W/T)(E/D)YVP(S/T)YEDKNDWM(L/F)VGDPV

Motif 18 is preferably:
(S/T)CK(R/K)(L/I)R(I/L)MK(G/S)(S/K/T)EA

Preferably the AUX/IAA polypeptide of the invention has in increasing order of preference at least 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% sequence identity to the amino acid of an AUX/IAA domain, preferably to the AUX/IAA domain of any of the polypeptides of Table A4, most preferably to the AUX/IAA domain of SEQ ID NO: 432 as represented by the amino acids located between amino acid coordinates 5 to 171.

Preferably, the IAA14-like polypeptide sequence which when used in the construction of a phylogenetic tree, as depicted in FIG. 1 in Remington et al. (Plant Physiol. 135, 1738-1752, 2004), clusters with group A of the IAA14-like polypeptides, which comprises the amino acid sequence represented by SEQ ID NO: 738, rather than with any other group (see also FIG. 15).

Alternatively, the homologue of an AUX/IAA protein has in increasing order of preference at least 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% overall sequence identity to the amino acid represented by any of the polypeptides of Table A4 or Table A5 preferably by SEQ ID NO: 432 or SEQ ID NO: 738, provided that the homologous protein comprises one or more of the conserved motifs as outlined above. The overall sequence identity is determined using a global alignment algorithm, such as the Needleman Wunsch algorithm in the program GAP (GCG Wisconsin Package, Accelrys), preferably with default parameters and preferably with sequences of mature proteins (i.e. without taking into account secretion signals or transit peptides). Compared to overall sequence identity, the sequence identity will generally be higher when only conserved domains or motifs are considered.

In a preferred embodiment, the polypeptide sequence which when used in the construction of a phylogenetic tree, as depicted in FIG. 1 in Remington et al. (Plant Physiol. 135, 1738-1752, 2004), clusters with group A of the IAA14-like polypeptides, which comprises the amino acid sequence represented by SEQ ID NO: 738, rather than with any other group (see also FIG. 15).

The terms "domain", "signature" and "motif" are defined in the "definitions" section herein. Specialist databases exist for the identification of domains, for example, SMART

(Schultz et al. (1998) Proc. Natl. Acad. Sci. USA 95, 5857-5864; Letunic et al. (2002) Nucleic Acids Res 30, 242-244), InterPro (Mulder et al., (2003) Nucl. Acids. Res. 31, 315-318), Prosite (Bucher and Bairoch (1994), A generalized profile syntax for biomolecular sequences motifs and its function in automatic sequence interpretation. (In) ISMB-94; Proceedings 2nd International Conference on Intelligent Systems for Molecular Biology. Altman R., Brutlag D., Karp P., Lathrop R., Searls D., Eds., pp 53-61, AAAI Press, Menlo Park; Hulo et al., Nucl. Acids. Res. 32:D134-D137, (2004)), or Pfam (Bateman et al., Nucleic Acids Research 30(1): 276-280 (2002)). A set of tools for in silico analysis of protein sequences is available on the ExPASy proteomics server (Swiss Institute of Bioinformatics (Gasteiger et al., ExPASy: the proteomics server for in-depth protein knowledge and analysis, Nucleic Acids Res. 31:3784-3788 (2003))). Domains or motifs may also be identified using routine techniques, such as by sequence alignment.

Concerning MYB91 polypeptides, an alignment of the polypeptides of Table A2 herein, is shown in FIG. 5. Such alignments are useful for identifying the most conserved domains or motifs between the MYB91 polypeptides as defined herein. Examples of such domains are (i) a MYB DNA binding domain with an InterPro accession number IPR014778, as represented by SEQ ID NO: 269 and/or by SEQ ID NO: 270 (marked by X's in FIG. 5); and (ii) a MYB DNA transcription factor with an InterPro entry IPR015495 (also marked by X's in FIG. 2). Another such domain is a C-terminal Conserved Domain as represented by SEQ ID NO: 271, also marked by X's in FIG. 5.

Methods for the alignment of sequences for comparison are well known in the art, such methods include GAP, BEST-FIT, BLAST, FASTA and TFASTA. GAP uses the algorithm of Needleman and Wunsch ((1970) J Mol Biol 48: 443-453) to find the global (i.e. spanning the complete sequences) alignment of two sequences that maximizes the number of matches and minimizes the number of gaps. The BLAST algorithm (Altschul et al. (1990) J Mol Biol 215: 403-10) calculates percent sequence identity and performs a statistical analysis of the similarity between the two sequences. The software for performing BLAST analysis is publicly available through the National Centre for Biotechnology Information (NCBI). Homologues may readily be identified using, for example, the ClustalW multiple sequence alignment algorithm (version 1.83), with the default pairwise alignment parameters, and a scoring method in percentage. Global percentages of similarity and identity may also be determined using one of the methods available in the MatGAT software package (Campanella et al., BMC Bioinformatics. 2003 Jul. 10; 4:29. MatGAT: an application that generates similarity/identity matrices using protein or DNA sequences.). Minor manual editing may be performed to optimise alignment between conserved motifs, as would be apparent to a person skilled in the art. Furthermore, instead of using full-length sequences for the identification of homologues, specific domains may also be used. The sequence identity values may be determined over the entire nucleic acid or amino acid sequence or over selected domains or conserved motif(s), using the programs mentioned above using the default parameters. For local alignments, the Smith-Waterman algorithm is particularly useful (Smith T F, Waterman M S (1981) J. Mol. Biol. 147(1); 195-7).

Concerning MYB91 polypeptides, example 3 herein describes in Table B the percentage identity between the MYB91 polypeptide as represented by SEQ ID NO: 221 and the MYB91 polypeptides listed in Table A2, which can be as low as 52% amino acid sequence identity. In some instances,

the default parameters may be adjusted to modify the stringency of the search. For example using BLAST, the statistical significance threshold (called "expect" value) for reporting matches against database sequences may be increased to show less stringent matches. This way, short nearly exact matches may be identified.

Concerning GASA polypeptides, an alignment can for example be made from the mature protein sequences, that is, without secretion signal peptide. Methods for identifying signal peptides are well known in the art, see for example Bendtsen et al., *J. Mol. Biol.*, 340:783-795 (2004).

The task of protein subcellular localisation prediction is important and well studied. Knowing a protein's localisation helps elucidate its function. Experimental methods for protein localization range from immunolocalization to tagging of proteins using green fluorescent protein (GFP) or beta-glucuronidase (GUS). Such methods are accurate although labor-intensive compared with computational methods. Recently much progress has been made in computational prediction of protein localisation from sequence data. Among algorithms well known to a person skilled in the art are available at the ExPASy Proteomics tools hosted by the Swiss Institute for Bioinformatics, for example, PSort, TargetP, ChloroP, LocTree, Predotar, LipoP, MITOPROT, PATS, PTS1, SignalP, TMHMM, and others. By applying the PSort algorithm to an MYB91 polypeptide as represented by SEQ ID NO: 221, a predicted nuclear subcellular localisation is obtained.

Furthermore, ASPAT polypeptides typically have Aspartate Transaminase also called Aspartate Transferase activity. Tools and techniques for measuring Aspartate Transaminase activity are well known in the art. Aspartate Transaminase activity may be for example assayed *in vivo* by complementation of *E. coli* strains defective in the activity as described by De la Torre et al. 2006. Alternatively, a biochemical determination of Aspartate Transferase activity may be carried out as for example described in De la Torre et al. 2006.

In addition, ASPAT polypeptides, when expressed in rice according to the methods of the present invention as outlined in the Examples section, give plants having increased yield related traits, in particular increased seed yield.

GASA polypeptides, when expressed in rice according to the methods of the present invention as outlined in the examples section, give plants having increased yield related traits, in particular increased total weight of seeds and/or increased number of filled seeds, and/or increased harvest index.

Furthermore, transgenic plants expressing GASA polypeptides (at least in their native form) may have enhanced tolerance to heat stress (Ko et al, 2007). Tools and techniques for measuring resistance of plants to heat stress are well known in the art, see for example the methods described in Ko et al., 2007.

Furthermore, AUX/IAA polypeptides (at least in their native form) typically have protein binding activity: AUX/IAA polypeptides bind to ARF (Auxin Response Factor) polypeptides. Tools and techniques for measuring protein binding activity are well known in the art and include for example, immuno precipitation of protein complexes and yeast two hybrid. Tools and techniques for measuring the association of AUX/IAA and ARF polypeptide are well known in the art., and include for example yeast two hybrid analysis (see for example Fukaki et al. (*Plant J.* 44, 382-395, 2005).

Typically AUX/IAA polypeptides of the invention comprise an EAR domain (Ohata et al; *Plant Cell.* 2001 13(8): 1959-68), which is a well known protein domain that typi-

cally confers repression activity to the transcription factors that comprising such domain. The AUX/IAA polypeptides of the invention have preferably transcription repression activity.

Concerning IAA14-like polypeptides, they (at least in their native form) typically associate with ARF7 or ARF19 proteins. Tools and techniques for measuring this association are well known in the art., and include for example yeast two hybrid analysis (see for example Fukaki et al. (*Plant J.* 44, 382-395, 2005) Further details are provided in the Examples section.

In addition, AUX/IAA polypeptides, when expressed in rice according to the methods of the present invention as outlined in the Examples section, give plants having increased yield related traits selected from increased harvest index, increased root biomass, increased green biomass and increased seed yield.

In addition, AUX/IAA polypeptides, when expressed in rice according to the methods of the present invention as outlined in the Examples section, give plants having increased yield related traits such as increase seed fill rate and increased harvest index.

In addition, IAA14-like polypeptides, when expressed in rice according to the methods of the present invention as outlined in the Examples section, give plants having increased yield related traits, preferably increased seed yield.

Additionally, AUX/IAA polypeptides may display a preferred subcellular localization, typically one or more of nuclear, cytoplasmic, chloroplastic, or mitochondrial. The task of protein subcellular localisation prediction is important and well studied. Knowing a protein's localisation helps elucidate its function. Experimental methods for protein localization range from immunolocalization to tagging of proteins using green fluorescent protein (GFP) or beta-glucuronidase (GUS). Such methods are accurate although labor-intensive compared with computational methods. Recently much progress has been made in computational prediction of protein localisation from sequence data. Among algorithms well known to a person skilled in the art are available at the ExPASy Proteomics tools hosted by the Swiss Institute for Bioinformatics, for example, PSort, TargetP, ChloroP, LocTree, Predotar, LipoP, MITOPROT, PATS, PTS1, SignalP, TMHMM, and others.

Concerning ASPAT polypeptides, the present invention is illustrated by transforming plants with the nucleic acid sequence represented by SEQ ID NO: 1, encoding the polypeptide sequence of SEQ ID NO: 2. However, performance of the invention is not restricted to these sequences; the methods of the invention may advantageously be performed using any ASPAT-encoding nucleic acid or ASPAT polypeptide as defined herein.

Examples of nucleic acids encoding ASPAT polypeptides are given in Table A1 of The Examples section herein. Such nucleic acids are useful in performing the methods of the invention. The amino acid sequences given in Table A1 of The Examples section are example sequences of orthologues and paralogues of the ASPAT polypeptide represented by SEQ ID NO: 2, the terms "orthologues" and "paralogues" being as defined herein. Further orthologues and paralogues may readily be identified by performing a so-called reciprocal blast search. Typically, this involves a first BLAST involving BLASTing a query sequence (for example using any of the sequences listed in Table A1 of The Examples section) against any sequence database, such as the publicly available NCBI database. BLASTN or TBLASTX (using standard default values) are generally used when starting from a nucleotide sequence, and BLASTP or TBLASTN (using standard

default values) when starting from a protein sequence. The BLAST results may optionally be filtered. The full-length sequences of either the filtered results or non-filtered results are then BLASTed back (second BLAST) against sequences from the organism from which the query sequence is derived (where the query sequence is SEQ ID NO: 1 or SEQ ID NO: 2, the second BLAST would therefore be against rice sequences). The results of the first and second BLASTs are then compared. A paralogue is identified if a high-ranking hit from the first blast is from the same species as from which the query sequence is derived, a BLAST back then ideally results in the query sequence amongst the highest hits; an orthologue is identified if a high-ranking hit in the first BLAST is not from the same species as from which the query sequence is derived, and preferably results upon BLAST back in the query sequence being among the highest hits.

Concerning MYB91 polypeptides, the present invention is illustrated by transforming plants with the nucleic acid sequence represented by SEQ ID NO: 220, encoding the MYB91 polypeptide sequence of SEQ ID NO: 221. However, performance of the invention is not restricted to these sequences; the methods of the invention may advantageously be performed using any nucleic acid sequence encoding an MYB91 polypeptide as defined herein.

Examples of nucleic acid sequences encoding MYB91 polypeptides are given in Table A2 of Example 1 herein. Such nucleic acid sequences are useful in performing the methods of the invention. The polypeptide sequences given in Table A2 of Example 1 are example sequences of orthologues and paralogues of the MYB91 polypeptide represented by SEQ ID NO: 221, the terms "orthologues" and "paralogues" being as defined herein. Further orthologues and paralogues may readily be identified by performing a so-called reciprocal blast search. Typically, this involves a first BLAST involving BLASTing a query sequence (for example using any of the sequences listed in Table A1 of Example 1) against any sequence database, such as the publicly available NCBI database. BLASTN or TBLASTX (using standard default values) are generally used when starting from a nucleotide sequence, and BLASTP or TBLASTN (using standard default values) when starting from a protein sequence. The BLAST results may optionally be filtered. The full-length sequences of either the filtered results or non-filtered results are then BLASTed back (second BLAST) against sequences from the organism from which the query sequence is derived (where the query sequence is SEQ ID NO: 220 or SEQ ID NO: 221, the second BLAST would therefore be against *Populus trichocarpa* sequences). The results of the first and second BLASTs are then compared. A paralogue is identified if a high-ranking hit from the first blast is from the same species as from which the query sequence is derived, a BLAST back then ideally results in the query sequence amongst the highest hits; an orthologue is identified if a high-ranking hit in the first BLAST is not from the same species as from which the query sequence is derived, and preferably results upon BLAST back in the query sequence being among the highest hits.

Concerning GASA polypeptides, the present invention is illustrated by transforming plants with the nucleic acid sequence represented by SEQ ID NO: 275, encoding the polypeptide sequence of SEQ ID NO: 276; and with SEQ ID NO: 361, encoding SEQ ID NO: 291. However, performance of the invention is not restricted to these sequences; the methods of the invention may advantageously be performed using any GASA-encoding nucleic acid or GASA polypeptide as defined herein. In a preferred embodiment, the nucleic acid encoding the GASA polypeptide, when expressed in a plant, is a heterologous nucleic acid, the heterologous nucleic acid

being sufficiently different from the endogenous GASA nucleic acid such that gene silencing is avoided.

Examples of nucleic acids encoding GASA polypeptides are given in Table A3 of the Examples section herein. Such nucleic acids are useful in performing the methods of the invention. The amino acid sequences given in Table A3 of the Examples section are example sequences of orthologues and paralogues of the GASA polypeptide represented by SEQ ID NO: 276, the terms "orthologues" and "paralogues" being as defined herein. Further orthologues and paralogues may readily be identified by performing a so-called reciprocal blast search. Typically, this involves a first BLAST involving BLASTing a query sequence (for example using any of the sequences listed in Table A3 of the Examples section) against any sequence database, such as the publicly available NCBI database. BLASTN or TBLASTX (using standard default values) are generally used when starting from a nucleotide sequence, and BLASTP or TBLASTN (using standard default values) when starting from a protein sequence. The BLAST results may optionally be filtered. The full-length sequences of either the filtered results or non-filtered results are then BLASTed back (second BLAST) against sequences from the organism from which the query sequence is derived (where the query sequence is SEQ ID NO: 275 or SEQ ID NO: 276, the second BLAST would therefore be against tomato (*Solanum lycopersicum*) sequences; where the query sequence is SEQ ID NO: 361 or SEQ ID NO: 291, the second BLAST would therefore be against poplar sequences). The results of the first and second BLASTs are then compared. A paralogue is identified if a high-ranking hit from the first blast is from the same species as from which the query sequence is derived, a BLAST back then ideally results in the query sequence amongst the highest hits; an orthologue is identified if a high-ranking hit in the first BLAST is not from the same species as from which the query sequence is derived, and preferably results upon BLAST back in the query sequence being among the highest hits.

Concerning AUX/IAA polypeptides, the present invention is illustrated by transforming plants with the nucleic acid sequence represented by SEQ ID NO: 431 or by SEQ ID NO: 737, encoding the polypeptide sequence of SEQ ID NO: 432 or by SEQ ID NO: 738.

However, performance of the invention is not restricted to these sequences; the methods of the invention may advantageously be performed using any AUX/IAA-encoding nucleic acid or IAA14-like polypeptide as defined herein.

Examples of nucleic acids encoding AUX/IAA polypeptides are given in Table A4 and in Table A5 of the Examples section herein. Such nucleic acids are useful in performing the methods of the invention. The amino acid sequences given in Table A4 and in Table A5 of the Examples section are example sequences of orthologues and paralogues of the AUX/IAA polypeptide represented by SEQ ID NO: 432 or by SEQ ID NO: 738, the terms "orthologues" and "paralogues" being as defined herein. Further orthologues and paralogues may readily be identified by performing a so-called reciprocal blast search. Typically, this involves a first BLAST involving BLASTing a query sequence (for example using any of the sequences listed in Table A4 or Table A5 of the Examples section) against any sequence database, such as the publicly available NCBI database. BLASTN or TBLASTX (using standard default values) are generally used when starting from a nucleotide sequence, and BLASTP or TBLASTN (using standard default values) when starting from a protein sequence. The BLAST results may optionally be filtered. The full-length sequences of either the filtered results or non-filtered results are then BLASTed back (second BLAST)

against sequences from the organism from which the query sequence is derived (where the query sequence is SEQ ID NO: 431 or SEQ ID NO: 432, the second BLAST would therefore be against *Arabidopsis* sequences). The results of the first and second BLASTs are then compared. A paralogue is identified if a high-ranking hit from the first blast is from the same species as from which the query sequence is derived, a BLAST back then ideally results in the query sequence amongst the highest hits; an orthologue is identified if a high-ranking hit in the first BLAST is not from the same species as from which the query sequence is derived, and preferably results upon BLAST back in the query sequence being among the highest hits.

High-ranking hits are those having a low E-value. The lower the E-value, the more significant the score (or in other words the lower the chance that the hit was found by chance). Computation of the E-value is well known in the art. In addition to E-values, comparisons are also scored by percentage identity. Percentage identity refers to the number of identical nucleotides (or amino acids) between the two compared nucleic acid (or polypeptide) sequences over a particular length. In the case of large families, ClustalW may be used, followed by a neighbour joining tree, to help visualize clustering of related genes and to identify orthologues and paralogues.

Nucleic acid variants may also be useful in practising the methods of the invention. Examples of such variants include nucleic acids encoding homologues and derivatives of any one of the amino acid sequences given in Table A1 to A5 of The Examples section, the terms "homologue" and "derivative" being as defined herein. Also useful in the methods of the invention are nucleic acids encoding homologues and derivatives of orthologues or paralogues of any one of the amino acid sequences given in Table A1 to A5 of The Examples section. Homologues and derivatives useful in the methods of the present invention have substantially the same biological and functional activity as the unmodified protein from which they are derived. Also included are nucleic acids variants in which codon usage is optimised or in which miRNA target sites are removed.

Further nucleic acid variants useful in practising the methods of the invention include portions of nucleic acids encoding ASPAT polypeptides, or MYB91 polypeptides, or GASA polypeptides, or AUX/IAA polypeptides, nucleic acids hybridising to nucleic acids encoding ASPAT polypeptides, or MYB91 polypeptides, or GASA polypeptides, or AUX/IAA polypeptides, splice variants of nucleic acids encoding ASPAT polypeptides, or MYB91 polypeptides, or GASA polypeptides, or AUX/IAA polypeptides, allelic variants of nucleic acids encoding ASPAT polypeptides, or MYB91 polypeptides, or GASA polypeptides, or AUX/IAA polypeptides, and variants of nucleic acids encoding ASPAT polypeptides, or MYB91 polypeptides, or GASA polypeptides, or AUX/IAA polypeptides, obtained by gene shuffling. The terms hybridising sequence, splice variant, allelic variant and gene shuffling are as described herein.

Nucleic acids encoding ASPAT polypeptides, or MYB91 polypeptides, or GASA polypeptides, or AUX/IAA polypeptides, need not be full-length nucleic acids, since performance of the methods of the invention does not rely on the use of full-length nucleic acid sequences. According to the present invention, there is provided a method for enhancing yield-related traits in plants, comprising introducing and expressing in a plant a portion of any one of the nucleic acid sequences given in Table A1 to A5 of The Examples section, or a portion

of a nucleic acid encoding an orthologue, paralogue or homologue of any of the amino acid sequences given in Table A1 to A5 of The Examples section.

A portion of a nucleic acid may be prepared, for example, by making one or more deletions to the nucleic acid. The portions may be used in isolated form or they may be fused to other coding (or non-coding) sequences in order to, for example, produce a protein that combines several activities. When fused to other coding sequences, the resultant polypeptide produced upon translation may be bigger than that predicted for the protein portion.

Concerning ASPAT polypeptides, portions useful in the methods of the invention, encode an ASPAT polypeptide as defined herein, and have substantially the same biological activity as the amino acid sequences given in Table A1 of The Examples section. Preferably, the portion is a portion of any one of the nucleic acids given in Table A1 of The Examples section, or is a portion of a nucleic acid encoding an orthologue or paralogue of any one of the amino acid sequences given in Table A1 of The Examples section.

Preferably the portion is at least 100, 200, 300, 400, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000 consecutive nucleotides in length, the consecutive nucleotides being of any one of the nucleic acid sequences given in Table A1 of The Examples section, or of a nucleic acid encoding an orthologue or paralogue of any one of the amino acid sequences given in Table A1 of The Examples section. Even more preferably the portion is a portion of the nucleic acid of SEQ ID NO: 1, most preferably is the nucleic acid of SEQ ID NO: 3. Preferably, the portion encodes a fragment of an amino acid sequence which, when used in the construction of a phylogenetic tree, such as the one depicted in FIG. 2 clusters in increasing order of preference with any of the polypeptides in phylogenetic class 1, class 2, class 3 and class 4 as set forth in Table B1. Most preferably the portion encodes the amino acid fragment as represented by SEQ ID NO: 4.

Concerning MYB91 polypeptides, portions useful in the methods of the invention, encode an MYB91 polypeptide as defined herein, and have substantially the same biological activity as the polypeptide sequences given in Table A2 of Example 1. Preferably, the portion is a portion of any one of the nucleic acid sequences given in Table A2 of Example 1, or is a portion of a nucleic acid sequence encoding an orthologue or paralogue of any one of the polypeptide sequences given in Table A2 of Example 1. Preferably the portion is, in increasing order of preference at least 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000, 1050 or more consecutive nucleotides in length, the consecutive nucleotides being of any one of the nucleic acid sequences given in Table A2 of Example 1, or of a nucleic acid sequence encoding an orthologue or paralogue of any one of the polypeptide sequences given in Table A2 of Example 1. Preferably, the portion is a portion of a nucleic sequence encoding a polypeptide sequence comprising (i) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a MYB DNA binding domain with an InterPro accession number IPR014778, as represented by SEQ ID NO: 269; and (ii) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a MYB DNA binding domain with an InterPro accession number IPR014778, as represented by SEQ ID NO: 270; and (iii) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a Conserved Domain as represented by SEQ ID NO: 271. More preferably, the portion is a portion of a nucleic sequence encoding a

polypeptide sequence having in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to the MYB91 polypeptide as represented by SEQ ID NO: 221 or to any of the polypeptide sequences given in Table A2 herein. Most preferably, the portion is a portion of the nucleic acid sequence of SEQ ID NO: 220.

Concerning GASA polypeptides, portions useful in the methods of the invention, encode a GASA polypeptide as defined herein, and have substantially the same biological activity as the amino acid sequences given in Table A3 of the Examples section. Preferably, the portion is a portion of any one of the nucleic acids given in Table A3 of the Examples section, or is a portion of a nucleic acid encoding an orthologue or paralogue of any one of the amino acid sequences given in Table A3 of the Examples section. Preferably the portion is at least 200, 300, 400, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000 consecutive nucleotides in length, the consecutive nucleotides being of any one of the nucleic acid sequences given in Table A3 of the Examples section, or of a nucleic acid encoding an orthologue or paralogue of any one of the amino acid sequences given in Table A3 of the Examples section. Most preferably the portion is a portion of the nucleic acid of SEQ ID NO: 275. Preferably, the portion encodes a fragment of an amino acid sequence which, when used in the construction of a phylogenetic tree, such as the one depicted in FIG. 9, clusters with the group of GASA polypeptides comprising the amino acid sequence represented by SEQ ID NO: 276 (or SEQ ID NO: 291 or SEQ ID NO: 292) rather than with any other group.

Concerning AUX/IAA polypeptides, portions useful in the methods of the invention, encode an AUX/IAA polypeptide as defined herein, and have substantially the same biological activity as the amino acid sequences given in Table A4 or in Table A5 of the Examples section. Preferably, the portion is a portion of any one of the nucleic acids given in Table A4 or in Table A5 of the Examples section, or is a portion of a nucleic acid encoding an orthologue or paralogue of any one of the amino acid sequences given in Table A4 or in Table A5 of the Examples section. Preferably the portion is at least 100, 200, 300, 400, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000 consecutive nucleotides in length, the consecutive nucleotides being of any one of the nucleic acid sequences given in Table A4 or in Table A5 of the Examples section, or of a nucleic acid encoding an orthologue or paralogue of any one of the amino acid sequences given in Table A4 or in Table A5 of the Examples section. Most preferably the portion is a portion of the nucleic acid of SEQ ID NO: 431 or of SEQ ID NO: 737. Preferably, the portion encodes a fragment of an amino acid sequence comprising an AUX/IAA domain (PFAM accession number PF2309, InterPro entry IPR003311).

In the case of an IAA14-like polypeptide, preferably, the portion encodes a fragment of an amino acid sequence which, when used in the construction of a phylogenetic tree, as depicted in FIG. 1 in Remington et al. (Plant Physiol. 135, 1738-1752, 2004), clusters with group A of the IAA14-like polypeptides, which comprises the amino acid sequence represented by SEQ ID NO: 738, rather than with any other group (see also FIG. 13).

Another nucleic acid variant useful in the methods of the invention is a nucleic acid capable of hybridising, under reduced stringency conditions, preferably under stringent conditions, with a nucleic acid encoding an ASPAT polypeptide, or an MYB91 polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide, as defined herein, or with a portion as defined herein.

According to the present invention, there is provided a method for enhancing yield-related traits in plants, comprising introducing and expressing in a plant a nucleic acid capable of hybridizing to any one of the nucleic acids given in Table A1 to A5 of The Examples section, or comprising introducing and expressing in a plant a nucleic acid capable of hybridising to a nucleic acid encoding an orthologue, paralogue or homologue of any of the nucleic acid sequences given in Table A1 to A5 of The Examples section.

Concerning ASPAT polypeptides, hybridising sequences useful in the methods of the invention encode an ASPAT polypeptide as defined herein, having substantially the same biological activity as the amino acid sequences given in Table A1 of The Examples section. Preferably, the hybridising sequence is capable of hybridising to the complement of any one of the nucleic acids given in Table A1 of The Examples section, or to a portion of any of these sequences, a portion being as defined above, or the hybridising sequence is capable of hybridising to the complement of a nucleic acid encoding an orthologue or paralogue of any one of the amino acid sequences given in Table A1 of The Examples section. Even more preferably, the hybridising sequence is capable of hybridising to the complement of a nucleic acid as represented by SEQ ID NO: 1 or to a portion thereof. Most preferably the hybridising sequence is as represented by SEQ ID NO: 3.

Preferably, the hybridising sequence encodes a polypeptide with an amino acid sequence which, when full-length and used in the construction of a phylogenetic tree, such as the one depicted in FIG. 2 clusters in increasing order of preference with any of the polypeptides in phylogenetic class 1, class 2, class 3 and class 4 as set forth in Table B1.

Concerning MYB91 polypeptides, hybridising sequences useful in the methods of the invention encode an MYB91 polypeptide as defined herein, and have substantially the same biological activity as the polypeptide sequences given in Table A2 of Example 1. Preferably, the hybridising sequence is capable of hybridising to any one of the nucleic acid sequences given in Table A2 of Example 1, or to a complement thereof, or to a portion of any of these sequences, a portion being as defined above, or wherein the hybridising sequence is capable of hybridising to a nucleic acid sequence encoding an orthologue or paralogue of any one of the polypeptide sequences given in Table A2 of Example 1, or to a complement thereof. Preferably, the hybridising sequence is capable of hybridising to a nucleic acid sequence encoding a polypeptide sequence comprising (i) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a MYB DNA binding domain with an InterPro accession number IPR014778, as represented by SEQ ID NO: 269; and (ii) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a MYB DNA binding domain with an InterPro accession number IPR014778, as represented by SEQ ID NO: 270; and (iii) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a Conserved Domain as represented by SEQ ID NO: 271. More preferably, the hybridising sequence is capable of hybridising to a nucleic acid sequence encoding a polypeptide sequence having in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to the MYB91 polypeptide as represented by SEQ ID NO: 221 or to any of the polypeptide sequences given in Table A2 herein. Most preferably, the hybridising sequence is

capable of hybridising to a nucleic acid sequence as represented by SEQ ID NO: 220 or to a portion thereof.

Concerning GASA polypeptides, hybridising sequences useful in the methods of the invention encode a GASA polypeptide as defined herein, having substantially the same biological activity as the amino acid sequences given in Table A3 of the Examples section. Preferably, the hybridising sequence is capable of hybridising to the complement of any one of the nucleic acids given in Table A3 of the Examples section, or to a portion of any of these sequences, a portion being as defined above, or the hybridising sequence is capable of hybridising to the complement of a nucleic acid encoding an orthologue or paralogue of any one of the amino acid sequences given in Table A3 of the Examples section. Most preferably, the hybridising sequence is capable of hybridising to the complement of a nucleic acid as represented by SEQ ID NO: 275 or to a portion thereof.

Preferably, the hybridising sequence encodes a polypeptide with an amino acid sequence which, when full-length and used in the construction of a phylogenetic tree, such as the one depicted in FIG. 9, clusters with the group of GASA polypeptides comprising the amino acid sequence represented by SEQ ID NO: 276 (or SEQ ID NO: 291 or SEQ ID NO: 292) rather than with any other group.

Concerning AUX/IAA polypeptides, hybridising sequences useful in the methods of the invention encode an AUX/IAA polypeptide as defined herein, having substantially the same biological activity as the amino acid sequences given in Table A4 or in Table A5 of the Examples section. Preferably, the hybridising sequence is capable of hybridising to the complement of any one of the nucleic acids given in Table A4 or in Table A5 of the Examples section, or to a portion of any of these sequences, a portion being as defined above, or the hybridising sequence is capable of hybridising to the complement of a nucleic acid encoding an orthologue or paralogue of any one of the amino acid sequences given in Table A4 or in Table A5 of the Examples section. Most preferably, the hybridising sequence is capable of hybridising to the complement of a nucleic acid as represented by SEQ ID NO: 431 or of SEQ ID NO: 737 or to a portion thereof.

Preferably, the hybridising sequence or its complementary sequence encodes a polypeptide with an amino acid sequence comprising an AUX/IAA domain (PFAM accession number PF2309, InterPro entry IPR003311).

In the case IAA14-like polypeptides, preferably, the hybridising sequence encodes a polypeptide with an amino acid sequence which, when full-length and used in the construction of a phylogenetic tree, as depicted in FIG. 1 in Remington et al. (Plant Physiol. 135, 1738-1752, 2004), clusters with group A of the IAA14-like polypeptides, which comprises the amino acid sequence represented by SEQ ID NO: 738, rather than with any other group (see also FIG. 15).

Another nucleic acid variant useful in the methods of the invention is a splice variant encoding an ASPAT polypeptide, or an MYB91 polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide, as defined hereinabove, a splice variant being as defined herein.

According to the present invention, there is provided a method for enhancing yield-related traits in plants, comprising introducing and expressing in a plant a splice variant of any one of the nucleic acid sequences given in Table A1 to A5 of The Examples section, or a splice variant of a nucleic acid encoding an orthologue, paralogue or homologue of any of the amino acid sequences given in Table A1 to A5 of The Examples section.

Concerning ASPAT polypeptides, preferred splice variants are splice variants of a nucleic acid represented by SEQ ID

NO: 1, or a splice variant of a nucleic acid encoding an orthologue or paralogue of SEQ ID NO: 2. Preferably, the amino acid sequence encoded by the splice variant, when used in the construction of a phylogenetic tree, such as the one depicted in FIG. 2 clusters in increasing order of preference with any of the polypeptides in phylogenetic class 1, class 2, class 3 and class 4 as set forth in Table B1.

Concerning MYB91 polypeptides, preferred splice variants are splice variants of a nucleic acid sequence represented by SEQ ID NO: 220, or a splice variant of a nucleic acid sequence encoding an orthologue or paralogue of SEQ ID NO: 221. Preferably, the splice variant is a splice variant of a nucleic acid sequence encoding a polypeptide sequence comprising (i) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a MYB DNA binding domain with an InterPro accession number IPR014778, as represented by SEQ ID NO: 269; and (ii) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a MYB DNA binding domain with an InterPro accession number IPR014778, as represented by SEQ ID NO: 270; and (iii) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a Conserved Domain as represented by SEQ ID NO: 271. More preferably, the splice variant is a splice variant of a nucleic acid sequence encoding a polypeptide sequence having in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to the MYB91 polypeptide as represented by SEQ ID NO: 221 or to any of the polypeptide sequences given in Table A2 herein. Most preferably, the splice variant is a splice variant of a nucleic acid sequence as represented by SEQ ID NO: 220, or of a nucleic acid sequence encoding a polypeptide sequence as represented by SEQ ID NO: 221.

Concerning GASA polypeptides, preferred splice variants are splice variants of a nucleic acid represented by SEQ ID NO: 275, or a splice variant of a nucleic acid encoding an orthologue or paralogue of SEQ ID NO: 276. Preferably, the amino acid sequence encoded by the splice variant, when used in the construction of a phylogenetic tree, such as the one depicted in FIG. 93, clusters with the group of GASA polypeptides comprising the amino acid sequence represented by SEQ ID NO: 276 (or SEQ ID NO: 291 or SEQ ID NO: 292) rather than with any other group.

Concerning AUX/IAA polypeptides, preferred splice variants are splice variants of a nucleic acid represented by SEQ ID NO: 431 or of SEQ ID NO: 737, or a splice variant of a nucleic acid encoding an orthologue or paralogue of SEQ ID NO: 432 or of SEQ ID NO: 738.

Preferably, the amino acid sequence encoded by the splice variant comprises an AUX/IAA domain (PFAM accession number PF2309, InterPro entry IPR003311).

In the case of IAA14-like polypeptides, preferably, the amino acid sequence encoded by the splice variant, when used in the construction of a phylogenetic tree, as depicted in FIG. 1 in Remington et al. (Plant Physiol. 135, 1738-1752, 2004), clusters with group A of the IAA14-like polypeptides, which comprises the amino acid sequence represented by SEQ ID NO: 738, rather than with any other group (see also FIG. 15).

Another nucleic acid variant useful in performing the methods of the invention is an allelic variant of a nucleic acid encoding an ASPAT polypeptide, or an MYB91 polypeptide,

or a GASA polypeptide, or an AUX/IAA polypeptide, as defined hereinabove, an allelic variant being as defined herein.

According to the present invention, there is provided a method for enhancing yield-related traits in plants, comprising introducing and expressing in a plant an allelic variant of any one of the nucleic acids given in Table A1 to A5 of The Examples section, or comprising introducing and expressing in a plant an allelic variant of a nucleic acid encoding an orthologue, paralogue or homologue of any of the amino acid sequences given in Table A1 to A5 of The Examples section.

Concerning ASPAT polypeptides, the polypeptides encoded by allelic variants useful in the methods of the present invention have substantially the same biological activity as the ASPAT polypeptide of SEQ ID NO: 2 and any of the amino acids depicted in Table A1 of The Examples section. Allelic variants exist in nature, and encompassed within the methods of the present invention is the use of these natural alleles. Preferably, the allelic variant is an allelic variant of SEQ ID NO: 1 or an allelic variant of a nucleic acid encoding an orthologue or paralogue of SEQ ID NO: 2. Preferably, the amino acid sequence encoded by the allelic variant, when used in the construction of a phylogenetic tree, such as the one depicted in FIG. 2 clusters in increasing order of preference with any of the polypeptides in phylogenetic class 1, class 2, class 3 and class 4 as set forth in Table B1.

Concerning MYB91 polypeptides, the allelic variants useful in the methods of the present invention have substantially the same biological activity as the MYB91 polypeptide of SEQ ID NO: 221 and any of the polypeptide sequences depicted in Table A2 of Example 1. Allelic variants exist in nature, and encompassed within the methods of the present invention is the use of these natural alleles. Preferably, the allelic variant is an allelic variant of a polypeptide sequence comprising (i) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a MYB DNA binding domain with an InterPro accession number IPR014778, as represented by SEQ ID NO: 269; and (ii) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a MYB DNA binding domain with an InterPro accession number IPR014778, as represented by SEQ ID NO: 270; and (iii) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a Conserved Domain as represented by SEQ ID NO: 271. More preferably the allelic variant is an allelic variant encoding a polypeptide sequence having in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to the MYB91 polypeptide as represented by SEQ ID NO: 221 or to any of the polypeptide sequences given in Table A2 herein. Most preferably, the allelic variant is an allelic variant of SEQ ID NO: 220 or an allelic variant of a nucleic acid sequence encoding an orthologue or paralogue of SEQ ID NO: 221.

Concerning GASA polypeptides, the polypeptides encoded by allelic variants useful in the methods of the present invention have substantially the same biological activity as the GASA polypeptide of SEQ ID NO: 276 and any of the amino acids depicted in Table A3 of the Examples section. Allelic variants exist in nature, and encompassed within the methods of the present invention is the use of these natural alleles. Preferably, the allelic variant is an allelic variant of SEQ ID NO: 275 or an allelic variant of a nucleic acid encoding an orthologue or paralogue of SEQ ID NO: 276.

Preferably, the amino acid sequence encoded by the allelic variant, when used in the construction of a phylogenetic tree, such as the one depicted in FIG. 9, clusters with the group of GASA polypeptides comprising the amino acid sequence represented by SEQ ID NO: 276 (or SEQ ID NO: 291 or SEQ ID NO: 292) rather than with any other group.

Concerning AUX/IAA polypeptides, the polypeptides encoded by allelic variants useful in the methods of the present invention have substantially the same biological activity as the AUX/IAA polypeptide of SEQ ID NO: 432 or of SEQ ID NO: 738 and any of the amino acids depicted in Table A4 or in Table A5 of the Examples section. Allelic variants exist in nature, and encompassed within the methods of the present invention is the use of these natural alleles. Preferably, the allelic variant is an allelic variant of SEQ ID NO: 431 or of SEQ ID NO: 737 or an allelic variant of a nucleic acid encoding an orthologue or paralogue of SEQ ID NO: 432 or of SEQ ID NO: 738. Preferably, the amino acid sequence encoded by the allelic variant comprises an AUX/IAA domain (PFAM accession number PF2309, InterPro entry IPR003311). In the case of IAA14-like, preferably, the amino acid sequence encoded by the allelic variant, when used in the construction of a phylogenetic tree, as depicted in FIG. 1 in Remington et al. (Plant Physiol. 135, 1738-1752, 2004), clusters with group A of the IAA14-like polypeptides, which comprises the amino acid sequence represented by SEQ ID NO: 738, rather than with any other group (see also FIG. 15).

Gene shuffling or directed evolution may also be used to generate variants of nucleic acids encoding ASPAT polypeptides, or MYB91 polypeptides, GASA polypeptides, AUX/IAA polypeptides, or as defined above; the term "gene shuffling" being as defined herein.

According to the present invention, there is provided a method for enhancing yield-related traits in plants, comprising introducing and expressing in a plant a variant of any one of the nucleic acid sequences given in Table A1 to A5 of The Examples section, or comprising introducing and expressing in a plant a variant of a nucleic acid encoding an orthologue, paralogue or homologue of any of the amino acid sequences given in Table A1 to A5 of The Examples section, which variant nucleic acid is obtained by gene shuffling.

Concerning ASPAT polypeptides, preferably, the amino acid sequence encoded by the variant nucleic acid obtained by gene shuffling, when used in the construction of a phylogenetic tree such as the one depicted in FIG. 2 clusters in increasing order of preference with any of the polypeptides in phylogenetic class 1, class 2, class 3 and class 4 as set forth in Table B1.

Concerning MYB91 polypeptides, preferably, the variant nucleic acid sequence obtained by gene shuffling encodes a polypeptide sequence comprising (i) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a MYB DNA binding domain with an InterPro accession number IPR014778, as represented by SEQ ID NO: 269; and (ii) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a MYB DNA binding domain with an InterPro accession number IPR014778, as represented by SEQ ID NO: 270; and (iii) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a Conserved Domain as represented by SEQ ID NO: 271. More preferably, the variant nucleic acid sequence obtained by gene shuffling encodes a polypeptide sequence having in increasing order of preference at least

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50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to the MYB91 polypeptide as represented by SEQ ID NO: 221 or to any of the polypeptide sequences given in Table A1 herein. Most preferably, the nucleic acid sequence obtained by gene shuffling encodes a polypeptide sequence as represented by SEQ ID NO: 221.

Concerning GASA polypeptides, preferably, the amino acid sequence encoded by the variant nucleic acid obtained by gene shuffling, when used in the construction of a phylogenetic tree such as the one depicted in FIG. 9, clusters with the group of GASA polypeptides comprising the amino acid sequence represented by SEQ ID NO: 276 (or SEQ ID NO: 291 or SEQ ID NO: 292) rather than with any other group.

In the case of IAA14-like polypeptides, preferably, the amino acid sequence encoded by the variant nucleic acid obtained by gene shuffling, when used in the construction of a phylogenetic tree, as depicted in FIG. 1 in Remington et al. (Plant Physiol. 135, 1738-1752, 2004), clusters with group A of the IAA14-like polypeptides, which comprises the amino acid sequence represented by SEQ ID NO: 738, rather than with any other group (see also FIG. 15).

Furthermore, nucleic acid variants may also be obtained by site-directed mutagenesis. Several methods are available to achieve site-directed mutagenesis, the most common being PCR based methods (Current Protocols in Molecular Biology. Wiley Eds.).

Nucleic acids encoding ASPAT polypeptides may be derived from any natural or artificial source. The nucleic acid may be modified from its native form in composition and/or genomic environment through deliberate human manipulation. Preferably the ASPAT polypeptide-encoding nucleic acid is from a plant, further preferably from a monocotyledonous plant, more preferably from the family Poaceae, most preferably the nucleic acid is from *Oryza sativa*.

Advantageously, the invention also provides hitherto unknown ASPAT-encoding nucleic acids and ASPAT polypeptides.

According to a further embodiment of the present invention, there is therefore provided an isolated nucleic acid molecule selected from:

- (i) a nucleic acid represented by any one of SEQ ID NO: 81, 147, 153, 183 and 185;
- (ii) the complement of a nucleic acid represented by any one of SEQ ID NO: 81, 147, 153, 183 and 185;
- (iii) a nucleic acid encoding the polypeptide as represented by any one of SEQ ID NO: 82, 148, 154, 184 and 186, preferably as a result of the degeneracy of the genetic code, said isolated nucleic acid can be derived from a polypeptide sequence as represented by any one of SEQ ID NO: 82, 148, 154, 184 and 186 and further preferably confers enhanced yield-related traits relative to control plants;
- (iv) a nucleic acid having, in increasing order of preference at least 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% sequence identity with any of the nucleic acid sequences of Table A1 and further preferably conferring enhanced yield-related traits relative to control plants;
- (v) a nucleic acid molecule which hybridizes with a nucleic acid molecule of (i) to

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(iv) under stringent hybridization conditions and preferably confers enhanced yield-related traits relative to control plants;

(vi) a nucleic acid encoding an ASPAT polypeptide having, in increasing order of preference, at least 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% sequence identity to the amino acid sequence represented by any one of SEQ ID NO: 82, 148, 154, 184 and 186 and any of the other amino acid sequences in Table A1 and preferably conferring enhanced yield-related traits relative to control plants.

According to a further embodiment of the present invention, there is also provided an isolated polypeptide selected from:

- (i) an amino acid sequence represented by any one of SEQ ID NO: 82, 148, 154, 184 and 186;
- (ii) an amino acid sequence having, in increasing order of preference, at least 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% sequence identity to the amino acid sequence represented by any one of SEQ ID NO: 82, 148, 154, 184 and 186, and any of the other amino acid sequences in Table A1 and preferably conferring enhanced yield-related traits relative to control plants.
- (iii) derivatives of any of the amino acid sequences given in (i) or (ii) above.

Nucleic acid sequences encoding MYB91 polypeptides may be derived from any natural or artificial source. The nucleic acid sequence may be modified from its native form in composition and/or genomic environment through deliberate human manipulation. The nucleic acid sequence encoding an MYB91 polypeptide is from a plant, further preferably from a dicotyledonous plant, more preferably from the family Salicaceae, most preferably the nucleic acid sequence is from *Populus trichocarpa*.

Nucleic acids encoding GASA polypeptides may be derived from any natural or artificial source. The nucleic acid may be modified from its native form in composition and/or genomic environment through deliberate human manipulation. Preferably the GASA polypeptide-encoding nucleic acid is from a plant, further preferably from a dicotyledonous plant, more preferably from the family Solanaceae, most preferably the nucleic acid is from *Solanum lycopersicum*. Alternatively, the GASA polypeptide-encoding nucleic acid is from the family Salicaceae, preferably from *Populus* sp.

Nucleic acids encoding AUX/IAA polypeptides may be derived from any natural or artificial source. The nucleic acid may be modified from its native form in composition and/or genomic environment through deliberate human manipulation. Preferably the IAA14-like polypeptide-encoding nucleic acid is from a plant, further preferably from a monocotyledonous or a dicotyledonous plant, more preferably from the family Poaceae or Brassicaceae, most preferably the nucleic acid is from *Oryza sativa* or from *Arabidopsis thaliana*.

Performance of the methods of the invention gives plants having enhanced yield-related traits. In particular performance of the methods of the invention gives plants having increased yield, especially increased seed yield relative to

control plants. The terms “yield” and “seed yield” are described in more detail in the “definitions” section herein.

Reference herein to enhanced yield-related traits is taken to mean an increase in biomass (weight) of one or more parts of a plant, which may include aboveground (harvestable) parts and/or (harvestable) parts below ground. In particular, such harvestable parts are seeds, and performance of the methods of the invention results in plants having increased seed yield relative to the seed yield of control plants. Concerning GASA polypeptides, It should be noted that the plants with modulated expression of a nucleic acid encoding a GASA polypeptide according to the methods of this invention did not show significant changes in branching properties compared to the control plants.

Taking corn as an example, a yield increase may be manifested as one or more of the following: increase in the number of plants established per square meter, an increase in the number of ears per plant, an increase in the number of rows, number of kernels per row, kernel weight, thousand kernel weight, ear length/diameter, increase in the seed filling rate (which is the number of filled seeds divided by the total number of seeds and multiplied by 100), among others. Taking rice as an example, a yield increase may manifest itself as an increase in one or more of the following: number of plants per square meter, number of panicles per plant, number of spikelets per panicle, number of flowers (florets) per panicle (which is expressed as a ratio of the number of filled seeds over the number of primary panicles), increase in the seed filling rate (which is the number of filled seeds divided by the total number of seeds and multiplied by 100), increase in thousand kernel weight, among others.

The present invention provides a method for increasing yield, especially seed yield of plants, relative to control plants, which method comprises modulating expression in a plant of a nucleic acid encoding an ASPAT polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide, as defined herein.

The present invention also provides a method for increasing yield-related traits of plants relative to control plants, which method comprises increasing expression in a plant of a nucleic acid sequence encoding an MYB91 polypeptide as defined herein.

Since the transgenic plants according to the present invention have increased yield and/or increased yield-related traits, it is likely that these plants exhibit an increased growth rate (during at least part of their life cycle), relative to the growth rate of control plants at a corresponding stage in their life cycle.

The increased growth rate may be specific to one or more parts of a plant (including seeds), or may be throughout substantially the whole plant. Plants having an increased growth rate may have a shorter life cycle. The life cycle of a plant may be taken to mean the time needed to grow from a dry mature seed up to the stage where the plant has produced dry mature seeds, similar to the starting material. This life cycle may be influenced by factors such as early vigour, growth rate, greenness index, flowering time and speed of seed maturation. The increase in growth rate may take place at one or more stages in the life cycle of a plant or during substantially the whole plant life cycle. Increased growth rate during the early stages in the life cycle of a plant may reflect increased (early) vigour. The increase in growth rate may alter the harvest cycle of a plant allowing plants to be sown later and/or harvested sooner than would otherwise be possible (a similar effect may be obtained with earlier flowering time; delayed flowering is usually not a desired trait in crops). If the growth rate is sufficiently increased, it may allow for the

further sowing of seeds of the same plant species (for example sowing and harvesting of rice plants followed by sowing and harvesting of further rice plants all within one conventional growing period). Similarly, if the growth rate is sufficiently increased, it may allow for the further sowing of seeds of different plants species (for example the sowing and harvesting of corn plants followed by, for example, the sowing and optional harvesting of soybean, potato or any other suitable plant). Harvesting additional times from the same rootstock in the case of some crop plants may also be possible. Altering the harvest cycle of a plant may lead to an increase in annual biomass production per acre (due to an increase in the number of times (say in a year) that any particular plant may be grown and harvested). An increase in growth rate may also allow for the cultivation of transgenic plants in a wider geographical area than their wild-type counterparts, since the territorial limitations for growing a crop are often determined by adverse environmental conditions either at the time of planting (early season) or at the time of harvesting (late season). Such adverse conditions may be avoided if the harvest cycle is shortened. The growth rate may be determined by deriving various parameters from growth curves, such parameters may be: T-Mid (the time taken for plants to reach 50% of their maximal size) and T-90 (time taken for plants to reach 90% of their maximal size), amongst others.

According to a preferred feature of the present invention, performance of the methods of the invention gives plants having an increased growth rate relative to control plants. Therefore, according to the present invention, there is provided a method for increasing the growth rate of plants, which method comprises modulating expression in a plant of a nucleic acid encoding an ASPAT polypeptide, or an MYB91 polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide, as defined herein.

Increased yield-related traits occur whether the plant is under non-stress conditions or whether the plant is exposed to various stresses compared to control plants grown under comparable conditions. Plants typically respond to exposure to stress by growing more slowly. In conditions of severe stress, the plant may even stop growing altogether. Mild stress on the other hand is defined herein as being any stress to which a plant is exposed which does not result in the plant ceasing to grow altogether without the capacity to resume growth. Mild stress in the sense of the invention leads to a reduction in the growth of the stressed plants of less than 40%, 35% or 30%, preferably less than 25%, 20% or 15%, more preferably less than 14%, 13%, 12%, 11% or 10% or less in comparison to the control plant under non-stress conditions. Due to advances in agricultural practices (irrigation, fertilization, and/or pesticide treatments) severe stresses are not often encountered in cultivated crop plants. As a consequence, the compromised growth induced by mild stress is often an undesirable feature for agriculture. Mild stresses are the everyday biotic and/or abiotic (environmental) stresses to which a plant is exposed. Abiotic stresses may be due to drought or excess water, anaerobic stress, salt stress, chemical toxicity, oxidative stress and hot, cold or freezing temperatures. The abiotic stress may be an osmotic stress caused by a water stress (particularly due to drought), salt stress, oxidative stress or an ionic stress. Biotic stresses are typically those stresses caused by pathogens, such as bacteria, viruses, fungi, nematodes, and insects. The term “non-stress” conditions as used herein are those environmental conditions that allow optimal growth of plants. Persons skilled in the art are aware of normal soil conditions and climatic conditions for a given location.

Performance of the methods of the invention gives plants grown under non-stress conditions or under mild stress conditions having increased yield-related traits, relative to control plants grown under comparable conditions. Therefore, according to the present invention, there is provided a method for increasing yield-related traits in plants grown under non-stress conditions or under mild stress conditions, which method comprises increasing expression in a plant of a nucleic acid sequence encoding an MYB91 polypeptide.

In particular, the methods of the present invention may be performed under non-stress conditions or under conditions of mild drought to give plants having increased yield relative to control plants. As reported in Wang et al. (Planta (2003) 218: 1-14), abiotic stress leads to a series of morphological, physiological, biochemical and molecular changes that adversely affect plant growth and productivity. Drought, salinity, extreme temperatures and oxidative stress are known to be interconnected and may induce growth and cellular damage through similar mechanisms. Rabbani et al. (Plant Physiol (2003) 133: 1755-1767) describes a particularly high degree of "cross talk" between drought stress and high-salinity stress. For example, drought and/or salinisation are manifested primarily as osmotic stress, resulting in the disruption of homeostasis and ion distribution in the cell. Oxidative stress, which frequently accompanies high or low temperature, salinity or drought stress, may cause denaturing of functional and structural proteins. As a consequence, these diverse environmental stresses often activate similar cell signalling pathways and cellular responses, such as the production of stress proteins, up-regulation of anti-oxidants, accumulation of compatible solutes and growth arrest. The term "non-stress" conditions as used herein are those environmental conditions that allow optimal growth of plants. Persons skilled in the art are aware of normal soil conditions and climatic conditions for a given location. Plants with optimal growth conditions, (grown under non-stress conditions) typically yield in increasing order of preference at least 97%, 95%, 92%, 90%, 87%, 85%, 83%, 80%, 77% or 75% of the average production of such plant in a given environment. Average production may be calculated on harvest and/or season basis. Persons skilled in the art are aware of average yield productions of a crop.

Performance of the methods of the invention gives plants grown under non-stress conditions or under mild drought conditions increased yield relative to control plants grown under comparable conditions. Therefore, according to the present invention, there is provided a method for increasing yield in plants grown under non-stress conditions or under mild drought conditions, which method comprises modulating expression in a plant of a nucleic acid encoding an ASPAT polypeptide, or an MYB91 polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide.

The term "abiotic stress" as defined herein is taken to mean any one or more of: water stress (due to drought or excess water), anaerobic stress, salt stress, temperature stress (due to hot, cold or freezing temperatures), chemical toxicity stress and oxidative stress. According to one aspect of the invention, the abiotic stress is an osmotic stress, selected from water stress, salt stress, oxidative stress and ionic stress. Preferably, the water stress is drought stress. The term salt stress is not restricted to common salt (NaCl), but may be any stress caused by one or more of: NaCl, KCl, LiCl, MgCl₂, CaCl₂, amongst others.

Performance of the methods of the invention gives plants grown under conditions of salt stress, increased yield relative to control plants grown under comparable conditions. Therefore, according to the present invention, there is provided a

method for increasing yield in plants grown under conditions of salt stress, which method comprises modulating expression in a plant of a nucleic acid encoding an ASPAT polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide. The term salt stress is not restricted to common salt (NaCl), but may be any one or more of: NaCl, KCl, LiCl, MgCl₂, CaCl₂, amongst others.

Another example of abiotic environmental stress is the reduced availability of one or more nutrients that need to be assimilated by the plants for growth and development. Because of the strong influence of nutrition utilization efficiency on plant yield and product quality, a huge amount of fertilizer is poured onto fields to optimize plant growth and quality. Productivity of plants ordinarily is limited by three primary nutrients, phosphorous, potassium and nitrogen, which is usually the rate-limiting element in plant growth of these three. Therefore the major nutritional element required for plant growth is nitrogen (N). It is a constituent of numerous important compounds found in living cells, including amino acids, proteins (enzymes), nucleic acids, and chlorophyll. 1.5% to 2% of plant dry matter is nitrogen and approximately 16% of total plant protein. Thus, nitrogen availability is a major limiting factor for crop plant growth and production (Frink et al. (1999) Proc Natl Acad Sci USA 96(4): 1175-1180), and has as well a major impact on protein accumulation and amino acid composition. Therefore, of great interest are crop plants with increased yield-related traits, when grown under nitrogen-limiting conditions.

Performance of the methods of the invention gives plants grown under conditions of nutrient deficiency, particularly under conditions of nitrogen deficiency, increased yield relative to control plants grown under comparable conditions. Therefore, according to the present invention, there is provided a method for increasing yield in plants grown under conditions of nutrient deficiency, which method comprises modulating expression in a plant of a nucleic acid encoding an ASPAT polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide. Nutrient deficiency may result from a lack of nutrients such as nitrogen, phosphates and other phosphorous-containing compounds, potassium, calcium, cadmium, magnesium, manganese, iron and boron, amongst others.

Performance of the methods of the invention gives plants grown under conditions of reduced nutrient availability, particularly under conditions of reduced nitrogen availability, having increased yield-related traits relative to control plants grown under comparable conditions. Therefore, according to the present invention, there is provided a method for increasing yield-related traits in plants grown under conditions of reduced nutrient availability, preferably reduced nitrogen availability, which method comprises increasing expression in a plant of a nucleic acid sequence encoding an MYB91 polypeptide. Reduced nutrient availability may result from a deficiency or excess of nutrients such as nitrogen, phosphates and other phosphorous-containing compounds, potassium, calcium, cadmium, magnesium, manganese, iron and boron, amongst others. Preferably, reduced nutrient availability is reduced nitrogen availability.

Performance of the methods of the invention gives plants having increased yield-related traits, under abiotic stress conditions relative to control plants grown in comparable stress conditions. Therefore, according to the present invention, there is provided a method for increasing yield-related traits, in plants grown under abiotic stress conditions, which method comprises increasing expression in a plant of a nucleic acid sequence encoding an MYB91 polypeptide. According to one aspect of the invention, the abiotic stress is an osmotic stress,

selected from one or more of the following: water stress, salt stress, oxidative stress and ionic stress.

The present invention encompasses plants or parts thereof (including seeds) or cells thereof obtainable by the methods according to the present invention. The plants or parts thereof comprise a nucleic acid transgene encoding an ASPAT polypeptide, or an MYB91 polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide, as defined above, operably linked to a promoter functioning in plants.

The invention also provides genetic constructs and vectors to facilitate introduction and/or expression in plants of nucleic acids encoding ASPAT polypeptides, or MYB91 polypeptides, or GASA polypeptides, or AUX/IAA polypeptides, as defined herein. The gene constructs may be inserted into vectors, which may be commercially available, suitable for transforming into plants and suitable for expression of the gene of interest in the transformed cells. The invention also provides use of a gene construct as defined herein in the methods of the invention.

More specifically, the present invention provides a construct comprising:

- (a) a nucleic acid encoding an ASPAT polypeptide, or an MYB91 polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide, as defined above;
- (b) one or more control sequences capable of driving expression of the nucleic acid sequence of (a); and optionally
- (c) a transcription termination sequence.

Preferably, the nucleic acid encoding is an ASPAT polypeptide, or an MYB91 polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide, as defined above. The term "control sequence" and "termination sequence" are as defined herein.

Concerning MYB91 polypeptides, preferably, one of the control sequences of a construct is a constitutive promoter isolated from a plant genome. An example of a constitutive promoter is a GOS2 promoter, preferably a GOS2 promoter from rice, most preferably a GOS2 sequence as represented by SEQ ID NO: 272.

Plants are transformed with a vector comprising any of the nucleic acids described above. The skilled artisan is well aware of the genetic elements that must be present on the vector in order to successfully transform, select and propagate host cells containing the sequence of interest. The sequence of interest is operably linked to one or more control sequences (at least to a promoter).

Advantageously, any type of promoter, whether natural or synthetic, may be used to drive expression of the nucleic acid sequence, but preferably the promoter is of plant origin. A constitutive promoter is particularly useful in the methods. Preferably the constitutive promoter is also a ubiquitous promoter of medium strength. See the "Definitions" section herein for definitions of the various promoter types. Concerning ASPAT polypeptides, also useful in the methods of the invention is a green tissue-specific promoter.

Concerning MYB91 polypeptides, advantageously, any type of promoter, whether natural or synthetic, may be used to increase expression of the nucleic acid sequence. A constitutive promoter is particularly useful in the methods, preferably a constitutive promoter isolated from a plant genome. The plant constitutive promoter drives expression of a coding sequence at a level that is in all instances below that obtained under the control of a 35S CaMV viral promoter. An example of such a promoter is a GOS2 promoter as represented by SEQ ID NO: 272.

Concerning MYB91 polypeptides, organ-specific promoters, for example for preferred expression in leaves, stems,

tubers, meristems, seeds, are useful in performing the methods of the invention. Developmentally-regulated and inducible promoters are also useful in performing the methods of the invention. See the "Definitions" section herein for definitions of the various promoter types.

Concerning ASPAT polypeptides, it should be clear that the applicability of the present invention is not restricted to the ASPAT polypeptide-encoding nucleic acid represented by SEQ ID NO: 1, nor is the applicability of the invention restricted to expression of an ASPAT polypeptide-encoding nucleic acid when driven by a constitutive promoter, or when driven by a green tissue-specific promoter.

The constitutive promoter is preferably a medium strength promoter, more preferably selected from a plant derived promoter, such as a GOS2 promoter, more preferably is the promoter GOS2 promoter from rice. Further preferably the constitutive promoter is represented by a nucleic acid sequence substantially similar to SEQ ID NO: 218, most preferably the constitutive promoter is as represented by SEQ ID NO: 218. See the "Definitions" section herein for further examples of constitutive promoters.

According to another preferred feature of the invention, the nucleic acid encoding an ASPAT polypeptide is operably linked to a green tissue-specific promoter. The green tissue-specific promoter is preferably a promoter of the a Protochlorophyllide reductase (PR) gene, more preferably the PR promoter is from rice, further preferably the PR promoter is represented by a nucleic acid sequence substantially similar to SEQ ID NO: 219, most preferably the promoter is as represented by SEQ ID NO: 219. Examples of other green tissue-specific promoters which may also be used to perform the methods of the invention are shown in Table 3 in the "Definitions" section above.

Concerning MYB91 polypeptides, it should be clear that the applicability of the present invention is not restricted to a nucleic acid sequence encoding the MYB91 polypeptide, as represented by SEQ ID NO: 220, nor is the applicability of the invention restricted to expression of an MYB91 polypeptide-encoding nucleic acid sequence when driven by a constitutive promoter.

Concerning GASA polypeptides, it should be clear that the applicability of the present invention is not restricted to the GASA polypeptide-encoding nucleic acid represented by SEQ ID NO: 275 or SEQ ID NO: 361, nor is the applicability of the invention restricted to expression of a GASA polypeptide-encoding nucleic acid when driven by a constitutive promoter.

The constitutive promoter is preferably a medium strength promoter, more preferably selected from a plant derived promoter, such as a GOS2 promoter, more preferably is the promoter a GOS2 promoter from rice. Further preferably the constitutive promoter is represented by a nucleic acid sequence substantially similar to SEQ ID NO: 290, most preferably the constitutive promoter is as represented by SEQ ID NO: 290. See the "Definitions" section herein for further examples of constitutive promoters.

Optionally, one or more terminator sequences may be used in the construct introduced into a plant. Preferably, the construct comprises an expression cassette comprising a GOS2 promoter and the nucleic acid encoding the GASA polypeptide.

Concerning AUX/IAA polypeptides, it should be clear that the applicability of the present invention is not restricted to the AUX/IAA polypeptide-encoding nucleic acid represented by SEQ ID NO: 431 or by SEQ ID NO: 737, nor is the

applicability of the invention restricted to expression of an AUX/IAA polypeptide-encoding nucleic acid when driven by a constitutive promoter.

The constitutive promoter is preferably a medium strength promoter, more preferably selected from a plant derived promoter, such as a GOS2 promoter, more preferably is the promoter GOS2 promoter from rice. Further preferably the constitutive promoter is represented by a nucleic acid sequence substantially similar to SEQ ID NO: 669, most preferably the constitutive promoter is as represented by SEQ ID NO: 669. See the "Definitions" section herein for further examples of constitutive promoters.

Alternatively, the constitutive promoter is preferably a weak constitutive promoter, more preferably selected from a plant derived promoter, such as a High Mobility Group Protein (HMGP) promoter, more preferably is the promoter HMGP promoter from rice. Further preferably the constitutive promoter is represented by a nucleic acid sequence substantially similar to SEQ ID NO: 747, most preferably the constitutive promoter is as represented by SEQ ID NO: 747. See the "Definitions" section herein for further examples of constitutive promoters.

Optionally, one or more terminator sequences may be used in the construct introduced into a plant. Preferably, the construct comprises an expression cassette comprising a GOS2 or a HMGP promoter, substantially similar to SEQ ID NO: 669 or to SEQ ID NO: 747 respectively, and the nucleic acid encoding the AUX/IAA polypeptide.

Additional regulatory elements may include transcriptional as well as translational enhancers. Those skilled in the art will be aware of terminator and enhancer sequences that may be suitable for use in performing the invention. An intron sequence may also be added to the 5' untranslated region (UTR) or in the coding sequence to increase the amount of the mature message that accumulates in the cytosol, as described in the definitions section. Other control sequences (besides promoter, enhancer, silencer, intron sequences, 3'UTR and/or 5'UTR regions) may be protein and/or RNA stabilizing elements. Such sequences would be known or may readily be obtained by a person skilled in the art.

The genetic constructs of the invention may further include an origin of replication sequence that is required for maintenance and/or replication in a specific cell type. One example is when a genetic construct is required to be maintained in a bacterial cell as an episomal genetic element (e.g. plasmid or cosmid molecule). Preferred origins of replication include, but are not limited to, the fl-ori and colE1.

For the detection of the successful transfer of the nucleic acid sequences as used in the methods of the invention and/or selection of transgenic plants comprising these nucleic acids, it is advantageous to use marker genes (or reporter genes). Therefore, the genetic construct may optionally comprise a selectable marker gene. Selectable markers are described in more detail in the "definitions" section herein. The marker genes may be removed or excised from the transgenic cell once they are no longer needed. Techniques for marker removal are known in the art, useful techniques are described above in the definitions section.

It is known that upon stable or transient integration of nucleic acid sequences into plant cells, only a minority of the cells takes up the foreign DNA and, if desired, integrates it into its genome, depending on the expression vector used and the transfection technique used. To identify and select these integrants, a gene coding for a selectable marker (such as the ones described above) is usually introduced into the host cells together with the gene of interest. These markers can for example be used in mutants in which these genes are not

functional by, for example, deletion by conventional methods. Furthermore, nucleic acid sequence molecules encoding a selectable marker can be introduced into a host cell on the same vector that comprises the sequence encoding the polypeptides of the invention or used in the methods of the invention, or else in a separate vector. Cells which have been stably transfected with the introduced nucleic acid sequence can be identified for example by selection (for example, cells which have integrated the selectable marker survive whereas the other cells die). The marker genes may be removed or excised from the transgenic cell once they are no longer needed. Techniques for marker gene removal are known in the art, useful techniques are described above in the definitions section.

The invention also provides a method for the production of transgenic plants having enhanced yield-related traits relative to control plants, comprising introduction and expression in a plant of any nucleic acid encoding an ASPAT polypeptide, or an MYB91 polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide, as defined hereinabove.

More specifically, the present invention provides a method for the production of transgenic plants having enhanced yield-related traits, particularly increased seed yield, which method comprises:

- (i) introducing and expressing in a plant, plant part, or plant cell a nucleic acid encoding an ASPAT polypeptide, or an MYB91 polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide; and
- (ii) cultivating the plant cell under conditions promoting plant growth and development.

The nucleic acid of (i) may be any of the nucleic acids capable of encoding an ASPAT polypeptide, or an MYB91 polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide, as defined herein.

The nucleic acid may be introduced directly into a plant cell or into the plant itself (including introduction into a tissue, organ or any other part of a plant). According to a preferred feature of the present invention, the nucleic acid is preferably introduced into a plant by transformation. The term "transformation" is described in more detail in the "definitions" section herein.

The nucleic acid may be introduced directly into a plant cell or into the plant itself (including introduction into a tissue, organ or any other part of a plant). According to a preferred feature of the present invention, the nucleic acid is preferably introduced into a plant by transformation. The term "transformation" is described in more detail in the "definitions" section herein.

The genetically modified plant cells can be regenerated via all methods with which the skilled worker is familiar. Suitable methods can be found in the above-mentioned publications by S. D. Kung and R. Wu, Potrykus or Hofgen and Willmitzer.

Generally after transformation, plant cells or cell groupings are selected for the presence of one or more markers which are encoded by plant-expressible genes co-transferred with the gene of interest, following which the transformed material is regenerated into a whole plant. To select transformed plants, the plant material obtained in the transformation is, as a rule, subjected to selective conditions so that transformed plants can be distinguished from untransformed plants. For example, the seeds obtained in the above-described manner can be planted and, after an initial growing period, subjected to a suitable selection by spraying. A further possibility consists in growing the seeds, if appropriate after sterilization, on agar plates using a suitable selection agent so that only the transformed seeds can grow into plants. Alter-

natively, the transformed plants are screened for the presence of a selectable marker such as the ones described above.

Following DNA transfer and regeneration, putatively transformed plants may also be evaluated, for instance using Southern analysis, for the presence of the gene of interest, copy number and/or genomic organisation. Alternatively or additionally, expression levels of the newly introduced DNA may be monitored using Northern and/or Western analysis, both techniques being well known to persons having ordinary skill in the art.

The generated transformed plants may be propagated by a variety of means, such as by clonal propagation or classical breeding techniques. For example, a first generation (or T1) transformed plant may be selfed and homozygous second-generation (or T2) transformants selected, and the T2 plants may then further be propagated through classical breeding techniques. The generated transformed organisms may take a variety of forms. For example, they may be chimeras of transformed cells and non-transformed cells; clonal transformants (e.g., all cells transformed to contain the expression cassette); grafts of transformed and untransformed tissues (e.g., in plants, a transformed rootstock grafted to an untransformed scion).

The present invention clearly extends to any plant cell or plant produced by any of the methods described herein, and to all plant parts and propagules thereof. The present invention extends further to encompass the progeny of a primary transformed or transfected cell, tissue, organ or whole plant that has been produced by any of the aforementioned methods, the only requirement being that progeny exhibit the same genotypic and/or phenotypic characteristic(s) as those produced by the parent in the methods according to the invention.

The invention also includes host cells containing an isolated nucleic acid encoding an ASPAT polypeptide, or an MYB91 polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide, as defined hereinabove. Preferred host cells according to the invention are plant cells. Host plants for the nucleic acids or the vector used in the method according to the invention, the expression cassette or construct or vector are, in principle, advantageously all plants, which are capable of synthesizing the polypeptides used in the inventive method.

The methods of the invention are advantageously applicable to any plant. Plants that are particularly useful in the methods of the invention include all plants which belong to the superfamily Viridiplantae, in particular monocotyledonous and dicotyledonous plants including fodder or forage legumes, ornamental plants, food crops, trees or shrubs. According to a preferred embodiment of the present invention, the plant is a crop plant. Examples of crop plants include soybean, sunflower, canola, alfalfa, rapeseed, linseed, cotton, tomato, potato and tobacco. Further preferably, the plant is a monocotyledonous plant. Examples of monocotyledonous plants include sugarcane. More preferably the plant is a cereal. Examples of cereals include rice, maize, wheat, barley, millet, rye, triticale, sorghum, emmer, spelt, *secale*, einkorn, teff, milo and oats.

The invention also extends to harvestable parts of a plant such as, but not limited to seeds, leaves, fruits, flowers, stems, roots, rhizomes, tubers and bulbs, which harvestable parts comprise a recombinant nucleic acid encoding an ASPAT polypeptide, or an MYB91 polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide. The invention furthermore relates to products derived, preferably directly derived, from a harvestable part of such a plant, such as dry pellets or powders, oil, fat and fatty acids, starch or proteins.

According to a preferred feature of the invention, the modulated expression is increased expression. Methods for

increasing expression of nucleic acids or genes, or gene products, are well documented in the art and examples are provided in the definitions section.

As mentioned above, a preferred method for modulating expression of a nucleic acid encoding an ASPAT polypeptide, or an MYB91 polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide, is by introducing and expressing in a plant a nucleic acid encoding an ASPAT polypeptide, or an MYB91 polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide; however the effects of performing the method, i.e. enhancing yield-related traits may also be achieved using other well known techniques, including but not limited to T-DNA activation tagging, TILLING, homologous recombination. A description of these techniques is provided in the definitions section.

The present invention also encompasses use of nucleic acids encoding ASPAT polypeptides, or GASA polypeptides, or AUX/IAA polypeptides, as described herein and use of these ASPAT polypeptides, or GASA polypeptides, or AUX/IAA polypeptides, in enhancing any of the aforementioned yield-related traits in plants.

The present invention also encompasses use of nucleic acid sequences encoding MYB91 polypeptides as described herein and use of these MYB91 polypeptides in increasing any of the aforementioned yield-related traits in plants, under normal growth conditions, under abiotic stress growth (preferably osmotic stress growth conditions) conditions, and under growth conditions of reduced nutrient availability, preferably under conditions of reduced nitrogen availability.

Nucleic acids encoding an ASPAT polypeptide, or an MYB91 polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide, described herein, or the ASPAT polypeptides, or MYB91 polypeptides, or GASA polypeptides, or AUX/IAA polypeptides, themselves, may find use in breeding programmes in which a DNA marker is identified which may be genetically linked to a gene encoding an ASPAT polypeptide, or an MYB91 polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide. The nucleic acids/genes, or the ASPAT polypeptides themselves may be used to define a molecular marker. This DNA or protein marker may then be used in breeding programmes to select plants having enhanced yield-related traits as defined hereinabove in the methods of the invention.

Allelic variants of a nucleic acid/gene encoding an ASPAT polypeptide, or an MYB91 polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide may also find use in marker-assisted breeding programmes. Such breeding programmes sometimes require introduction of allelic variation by mutagenic treatment of the plants, using for example EMS mutagenesis; alternatively, the programme may start with a collection of allelic variants of so called "natural" origin caused unintentionally. Identification of allelic variants then takes place, for example, by PCR. This is followed by a step for selection of superior allelic variants of the sequence in question and which give increased yield and/or yield-related traits. Selection is typically carried out by monitoring growth performance of plants containing different allelic variants of the sequence in question. Growth performance may be monitored in a greenhouse or in the field. Further optional steps include crossing plants in which the superior allelic variant was identified with another plant. This could be used, for example, to make a combination of interesting phenotypic features.

Nucleic acids encoding ASPAT polypeptides, or MYB91 polypeptides, or GASA polypeptides, or AUX/IAA polypeptides, may also be used as probes for genetically and physically mapping the genes that they are a part of, and as markers

for traits linked to those genes. Such information may be useful in plant breeding in order to develop lines with desired phenotypes. Such use of nucleic acids encoding an ASPAT polypeptide, or an MYB91 polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide, requires only a nucleic acid sequence of at least 15 nucleotides in length. The encoding nucleic acids may be used as restriction fragment length polymorphism (RFLP) markers. Southern blots (Sambrook J, Fritsch E F and Maniatis T (1989) *Molecular Cloning*, A Laboratory Manual) of restriction-digested plant genomic DNA may be probed with the encoding nucleic acids encoding an ASPAT polypeptide, or an MYB91 polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide. The resulting banding patterns may then be subjected to genetic analyses using computer programs such as MapMaker (Lander et al. (1987) *Genomics* 1: 174-181) in order to construct a genetic map. In addition, the nucleic acids may be used to probe Southern blots containing restriction endonuclease-treated genomic DNAs of a set of individuals representing parent and progeny of a defined genetic cross. Segregation of the DNA polymorphisms is noted and used to calculate the position of the nucleic acid encoding an ASPAT polypeptide, or an MYB91 polypeptide, or a GASA polypeptide, or an AUX/IAA polypeptide, in the genetic map previously obtained using this population (Botstein et al. (1980) *Am. J. Hum. Genet.* 32:314-331).

The production and use of plant gene-derived probes for use in genetic mapping is described in Bernatzky and Tanksley (1986) *Plant Mol. Biol. Reporter* 4: 37-41. Numerous publications describe genetic mapping of specific cDNA clones using the methodology outlined above or variations thereof. For example, F2 intercross populations, backcross populations, randomly mated populations, near isogenic lines, and other sets of individuals may be used for mapping. Such methodologies are well known to those skilled in the art.

The nucleic acid probes may also be used for physical mapping (i.e., placement of sequences on physical maps; see Hoheisel et al. In: *Non-mammalian Genomic Analysis: A Practical Guide*, Academic press 1996, pp. 319-346, and references cited therein).

In another embodiment, the nucleic acid probes may be used in direct fluorescence in situ hybridisation (FISH) mapping (Trask (1991) *Trends Genet.* 7:149-154). Although current methods of FISH mapping favour use of large clones (several kb to several hundred kb; see Laan et al. (1995) *Genome Res.* 5:13-20), improvements in sensitivity may allow performance of FISH mapping using shorter probes.

A variety of nucleic acid amplification-based methods for genetic and physical mapping may be carried out using the nucleic acids. Examples include allele-specific amplification (Kazazian (1989) *J. Lab. Clin. Med.* 11:95-96), polymorphism of PCR-amplified fragments (CAPS; Sheffield et al. (1993) *Genomics* 16:325-332), allele-specific ligation (Landegren et al. (1988) *Science* 241:1077-1080), nucleotide extension reactions (Sokolov (1990) *Nucleic Acid Res.* 18:3671), Radiation Hybrid Mapping (Walter et al. (1997) *Nat. Genet.* 7:22-28) and Happy Mapping (Dear and Cook (1989) *Nucleic Acid Res.* 17:6795-6807). For these methods, the sequence of a nucleic acid is used to design and produce primer pairs for use in the amplification reaction or in primer extension reactions. The design of such primers is well known to those skilled in the art. In methods employing PCR-based genetic mapping, it may be necessary to identify DNA sequence differences between the parents of the mapping cross in the region corresponding to the instant nucleic acid sequence. This, however, is generally not necessary for mapping methods.

Concerning ASPAT polypeptides, concerning GASA polypeptides, or an AUX/IAA polypeptide, the methods according to the present invention result in plants having enhanced yield-related traits, as described hereinbefore. These traits may also be combined with other economically advantageous traits, such as further yield-enhancing traits, tolerance to other abiotic and biotic stresses, traits modifying various architectural features and/or biochemical and/or physiological features.

Concerning MYB91 polypeptides, the methods according to the present invention result in plants having increased yield-related traits, as described hereinbefore. These traits may also be combined with other economically advantageous traits, such as further yield-increasing traits, tolerance to abiotic and biotic stresses, tolerance to herbicides, insecticides, traits modifying various architectural features and/or biochemical and/or physiological features.

Items

1. Aspartate AminoTransferase (ASPAT)

1. A method for enhancing yield-related traits in plants relative to control plants, comprising modulating expression in a plant of a nucleic acid encoding an ASPAT (Aspartate Aminotransferase) polypeptide comprising an Aminotransferase class I and II (Aminotran_1_2) domain (Interpro accession number: IPR004839; pfam accession number: PF00155), and optionally selecting plants having enhanced yield-related traits

2. Method according to item 1, wherein said ASPAT polypeptide comprising one or more of the following motifs having at least 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or 100% to any one or more of the following motif:

(i)
Motif 1: NPTG, (SEQ ID NO: 207)

(ii)
Motif 2: IVLLHACAHNPTGVDPT, (SEQ ID NO: 208)

(iii)
Motif 3: SRLILCSPSNPTGSVY (SEQ ID NO: 209)

wherein any amino acid residue may be substituted by a conserved amino acid.

3. Method according to item 1 or 2, wherein said modulated expression is effected by introducing and expressing in a plant a nucleic acid encoding an ASPAT polypeptide.

4. Method according to any one of items 1 to 3, wherein said nucleic acid encoding an ASPAT polypeptide encodes any one of the proteins listed in Table A or is a portion of such a nucleic acid, or a nucleic acid capable of hybridising with such a nucleic acid.

5. Method according to any one of items 1 to 4, wherein said nucleic acid sequence encodes an orthologue or paralogue of any of the proteins given in Table A1.

6. Method according to any preceding item, wherein said enhanced yield-related traits comprise increased yield, preferably increased biomass and/or increased seed yield relative to control plants.

7. Method according to any one of items 1 to 6, wherein said enhanced yield-related traits are obtained under non-stress conditions.

8. Method according to any one of items 1 to 6, wherein said enhanced yield-related traits are obtained under conditions of drought stress, salt stress or nitrogen deficiency.

9. Method according to any one of items 3 to 8, wherein said nucleic acid is operably linked to a constitutive promoter, preferably to a GOS2 promoter, most preferably to a GOS2 promoter from rice.
10. Method according to any one of items 3 to 8, wherein said nucleic acid is operably linked to a green tissue-specific promoter, preferably to a PR promoter, most preferably to a PR promoter from rice.
11. Method according to any one of items 1 to 10, wherein said nucleic acid encoding an ASPAT polypeptide is of plant origin, preferably from a dicotyledonous plant, further preferably from the family poaceae, more preferably from the genus *Oryza*, most preferably from *Oryza sativa*.
12. Plant or part thereof, including seeds, obtainable by a method according to any one of items 1 to 11, wherein said plant or part thereof comprises a recombinant nucleic acid encoding an ASPAT polypeptide.
13. Construct comprising:
 - (i) nucleic acid encoding an ASPAT polypeptide as defined in items 1 or 2;
 - (ii) one or more control sequences capable of driving expression of the nucleic acid sequence of (a); and optionally
 - (iii) a transcription termination sequence.
14. Construct according to item 13, wherein one of said control sequences is a constitutive promoter, preferably a GOS2 promoter, most preferably a GOS2 promoter from rice.
15. Construct according to item 13, wherein one of said control sequences is a green tissue-specific promoter, preferably to a PR promoter, most preferably to a PR promoter from rice.
16. Use of a construct according to item 13 to 15 in a method for making plants having increased yield, particularly increased biomass and/or increased seed yield relative to control plants.
17. Plant, plant part or plant cell transformed with a construct according to item 13 to 15.
18. Method for the production of a transgenic plant having increased yield, particularly increased biomass and/or increased seed yield relative to control plants, comprising:
 - (i) introducing and expressing in a plant a nucleic acid encoding an ASPAT polypeptide as defined in item 1 or 2; and
 - (ii) cultivating the plant cell under conditions promoting plant growth and development.
19. Transgenic plant having increased yield, particularly increased biomass and/or increased seed yield, relative to control plants, resulting from modulated expression of a nucleic acid encoding an ASPAT polypeptide as defined in item 1 or 2, or a transgenic plant cell derived from said transgenic plant.
20. Transgenic plant according to item 11, 17 or 18, or a transgenic plant cell derived thereof, wherein said plant is a crop plant or a monocot or a cereal, such as rice, maize, wheat, barley, millet, rye, triticale, sorghum emmer, spelt, *secale*, einkorn, teff, milo and oats.
21. Harvestable parts of a plant according to item 20, wherein said harvestable parts are preferably shoot biomass and/or seeds.
22. Products derived from a plant according to item 20 and/or from harvestable parts of a plant according to item 21.
23. Use of a nucleic acid encoding an ASPAT polypeptide in increasing yield, particularly in increasing seed yield and/or shoot biomass in plants, relative to control plants.

24. An isolated nucleic acid molecule selected from:
 - (a) a nucleic acid represented by any one of SEQ ID NO: 81, 147, 153, 183 and 185;
 - (b) the complement of a nucleic acid represented by any one of SEQ ID NO: 81, 147, 153, 183 and 185;
 - (c) a nucleic acid encoding the polypeptide as represented by any one of SEQ ID NO: 82, 148, 154, 184 and 186, preferably as a result of the degeneracy of the genetic code, said isolated nucleic acid can be derived from a polypeptide sequence as represented by any one of SEQ ID NO: 82, 148, 154, 184 and 186 and further preferably confers enhanced yield-related traits relative to control plants;
 - (d) a nucleic acid having, in increasing order of preference at least 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% sequence identity with any of the nucleic acid sequences of Table A1 and further preferably conferring enhanced yield-related traits relative to control plants;
 - (e) a nucleic acid molecule which hybridizes with a nucleic acid molecule of (i) to (iv) under stringent hybridization conditions and preferably confers enhanced yield-related traits relative to control plants;
 - (f) a nucleic acid encoding an ASPAT polypeptide having, in increasing order of preference, at least 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% sequence identity to the amino acid sequence represented by any one of SEQ ID NO: 82, 148, 154, 184 and 186 and any of the other amino acid sequences in Table A1 and preferably conferring enhanced yield-related traits relative to control plants.
25. An isolated polypeptide selected from:
 - (i) an amino acid sequence represented by any one of SEQ ID NO: 82, 148, 154, 184 and 186;
 - (ii) an amino acid sequence having, in increasing order of preference, at least 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% sequence identity to the amino acid sequence represented by any one of SEQ ID NO: 82, 148, 154, 184 and 186, and any of the other amino acid sequences in Table A and preferably conferring enhanced yield-related traits relative to control plants.
 - (iii) derivatives of any of the amino acid sequences given in (i) or (ii) above.
2. MYB91 like transcription factor (MYB91)
 1. A method for increasing yield-related traits in plants relative to control plants, comprising increasing expression in a plant of a nucleic acid sequence encoding a MYB91 like transcription factor (MYB91) polypeptide, which MYB91 polypeptide comprises (i) (i) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a MYB DNA binding domain with an InterPro

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- accession number IPR014778, as represented by SEQ ID NO: 269; and (ii) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a MYB DNA binding domain with an InterPro accession number IPR014778, as represented by SEQ ID NO: 270; and (iii) in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a Conserved Domain as represented by SEQ ID NO: 271, and optionally selecting for plants having increased yield-related traits.
2. Method according to item 1, wherein said MYB91 polypeptide comprises in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to a polypeptide as represented by SEQ ID NO: 221.
 3. Method according to item 1, wherein said MYB91 polypeptide comprises in increasing order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity to any of the polypeptide sequences given in Table A2 herein.
 4. Method according to item 1, wherein said MYB91 polypeptide, when used in the construction of a phylogenetic tree of MYB DNA-binding domain polypeptides, such as the one depicted in FIG. 4, clusters with the MYB91 group of polypeptides rather than with any other group.
 5. Method according to any preceding item, wherein said nucleic acid sequence encoding a MYB91 polypeptide is represented by any one of the nucleic acid sequence SEQ ID NOs given in Table A2 or a portion thereof, or a sequence capable of hybridising with any one of the nucleic acid sequences SEQ ID NOs given in Table A2, or to a complement thereof.
 6. Method according to any preceding item, wherein said nucleic acid sequence encodes an orthologue or paralogue of any of the polypeptide sequence SEQ ID NOs given in Table A2.
 7. Method according to any preceding item, wherein said increased expression is effected by any one or more of: T-DNA activation tagging, TILLING, or homologous recombination.
 8. Method according to any preceding item, wherein said increased expression is effected by introducing and expressing in a plant a nucleic acid sequence encoding a MYB91 polypeptide.
 9. Method according to any preceding item, wherein said increased yield-related trait is one or more of: increased plant height, increased harvest index (HI), and/or increased Thousand Kernel Weight (TKW).
 10. Method according to any preceding item, wherein said nucleic acid sequence is operably linked to a constitutive promoter.
 11. Method according to item 10, wherein said constitutive promoter is a GOS2 promoter, preferably a GOS2 promoter from rice, most preferably a GOS2 sequence as represented by SEQ ID NO: 272.
 12. Method according to any preceding item, wherein said nucleic acid sequence encoding a MYB91 polypeptide is from a plant, further preferably from a dicotyledonous plant, more preferably from the family Salicaceae, most preferably the nucleic acid sequence is from *Populus trichocarpa*.

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13. Plants, parts thereof (including seeds), or plant cells obtainable by a method according to any preceding item, wherein said plant, part or cell thereof comprises an isolated nucleic acid transgene encoding a MYB91 polypeptide.
 14. Construct comprising:
 - (a) a nucleic acid sequence encoding a MYB91 polypeptide as defined in any one of items 1 to 6;
 - (b) one or more control sequences capable of driving expression of the nucleic acid sequence of (a); and optionally
 - (c) a transcription termination sequence.
 15. Construct according to item 14 wherein said control sequence is a constitutive promoter.
 16. Construct according to item 15 wherein said constitutive promoter is a GOS2 promoter, preferably a GOS2 promoter from rice, most preferably a GOS2 sequence as represented by SEQ ID NO: 272.
 17. Use of a construct according to any one of items 14 to 16 in a method for making plants having increased yield-related traits relative to control plants, which increased yield-related traits are one or more of: increased plant height, increased harvest index (HI), and increased Thousand Kernel Weight (TKW).
 18. Plant, plant part or plant cell transformed with a construct according to any one of items 14 to 16.
 19. Method for the production of transgenic plants having increased yield-related traits relative to control plants, comprising:
 - (i) introducing and expressing in a plant, plant part, or plant cell, a nucleic acid sequence encoding a MYB91 polypeptide as defined in any one of items 1 to 6; and
 - (ii) cultivating the plant cell, plant part, or plant under conditions promoting plant growth and development.
 20. Transgenic plant having increased yield-related traits relative to control plants, resulting from increased expression of an isolated nucleic acid sequence encoding a MYB91 polypeptide as defined in any one of items 1 to 6, or a transgenic plant cell or transgenic plant part derived from said transgenic plant.
 21. Transgenic plant according to item 13, 18, or 20, wherein said plant is a crop plant or a monocot or a cereal, such as rice, maize, wheat, barley, millet, rye, triticale, sorghum and oats, or a transgenic plant cell derived from said transgenic plant.
 22. Harvestable parts comprising an isolated nucleic acid sequence encoding a MYB91 polypeptide, of a plant according to item 21, wherein said harvestable parts are preferably seeds.
 23. Products derived from a plant according to item 21 and/or from harvestable parts of a plant according to item 22.
 24. Use of a nucleic acid sequence encoding a MYB91 polypeptide as defined in any one of items 1 to 6, in increasing yield-related traits, comprising one or more of: increased plant height, increased harvest index (HI), and increased Thousand Kernel Weight (TKW).
3. Gibberellic Acid-Stimulated *Arabidopsis* (GASA)
1. A method for enhancing yield-related traits in plants relative to control plants, comprising modulating expression in a plant of a nucleic acid encoding a GASA polypeptide, wherein the sequence of said GASA polypeptide comprises a Pfam PF02704 domain, provided that said GASA protein is not GASA4 as represented by SEQ ID NO: 295.

2. Method according to item 1, wherein said GASA polypeptide comprises one or more of the following motifs:

(b) Motif 4, (SEQ ID NO: 277)

(c) Motif 5, (SEQ ID NO: 278)

(d) Motif 6 (SEQ ID NO: 279)

3. Method according to item 1 or 2, wherein said modulated expression is effected by introducing and expressing in a plant a nucleic acid encoding a GASA polypeptide.
4. Method according to any one of items 1 to 3, wherein said nucleic acid encoding a GASA polypeptide encodes any one of the proteins listed in Table A3 or is a portion of such a nucleic acid, or a nucleic acid capable of hybridising with such a nucleic acid.
5. Method according to any one of items 1 to 4, wherein said nucleic acid sequence encodes an orthologue or paralogue of any of the proteins given in Table A3.
6. Method according to any preceding item, wherein said enhanced yield-related traits comprise increased seed yield relative to control plants.
7. Method according to any one of items 1 to 6, wherein said enhanced yield-related traits are obtained under non-stress conditions.
8. Method according to any one of items 1 to 6, wherein said enhanced yield-related traits are obtained under conditions of drought stress, salt stress or nitrogen deficiency.
9. Method according to any one of items 3 to 8, wherein said nucleic acid is operably linked to a constitutive promoter, preferably to a GOS2 promoter, most preferably to a GOS2 promoter from rice.
10. Method according to any one of items 1 to 9, wherein said nucleic acid encoding a GASA polypeptide is of plant origin, preferably from a dicotyledonous plant.
11. Plant or part thereof, including seeds, obtainable by a method according to any one of items 1 to 10, wherein said plant or part thereof comprises a recombinant nucleic acid encoding a GASA polypeptide.
12. Construct comprising:
- nucleic acid encoding a GASA polypeptide as defined in items 1 or 2;
 - one or more control sequences capable of driving expression of the nucleic acid sequence of (a); and optionally
 - a transcription termination sequence.
13. Construct according to item 12, wherein one of said control sequences is a constitutive promoter, preferably a GOS2 promoter, most preferably a GOS2 promoter from rice.
14. Use of a construct according to item 12 or 13 in a method for making plants having increased yield, particularly increased seed yield relative to control plants.
15. Plant, plant part or plant cell transformed with a construct according to item 12 or 13.
16. Method for the production of a transgenic plant having increased yield, particularly increased seed yield relative to control plants, comprising:
- introducing and expressing in a plant a nucleic acid encoding a GASA polypeptide as defined in item 1 or 2; and
 - cultivating the plant cell under conditions promoting plant growth and development.
17. Transgenic plant having increased yield, particularly increased seed yield, relative to control plants, resulting from modulated expression of a nucleic acid encoding

GASA polypeptide as defined in item 1 or 2, or a transgenic plant cell derived from said transgenic plant.

18. Transgenic plant according to item 11, 15 or 17, or a transgenic plant cell derived thereof, wherein said plant is a crop plant or a monocot or a cereal, such as rice, maize, wheat, barley, millet, rye, triticale, sorghum emmer, spelt, *secale*, einkorn, teff, milo and oats.
19. Harvestable parts of a plant according to item 18, wherein said harvestable parts are seeds.
20. Products derived from a plant according to item 18 and/or from harvestable parts of a plant according to item 19.
21. Use of a nucleic acid encoding a GASA polypeptide in increasing yield, particularly in increasing seed yield in plants, relative to control plants.
4. Auxin/Indoleacetic Acid Genes (AUX/IAA)
- A method for enhancing yield-related traits in plants relative to control plants, comprising modulating expression in a plant of a nucleic acid encoding an AUX/IAA polypeptide comprising an AUX/IAA domain.
 - Method according to item 1, wherein said AUX/IAA domain has in increasing order of preference at least 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% sequence identity to the amino acid of an AUX/IAA domain, preferably to the AUX/IAA domain of any of the polypeptides of Table A4, most preferably to the AUX/IAA domain of SEQ ID NO: 432 as represented by the amino acids located between amino acid coordinates 5 to 171.
 - Method according to item 1 wherein said AUX/IAA polypeptide is an IAA14-like polypeptide comprises one or more of the following motifs:
 - Motif 13: SEQ ID NO: 739,
 - Motif 14: SEQ ID NO: 740,
 - Motif 15: SEQ ID NO: 741,
 - Motif 16: SEQ ID NO: 742,
 - Motif 17: SEQ ID NO: 743,
 - Motif 18: SEQ ID NO: 744.
 - Method according to item 1 to 3, wherein said modulated expression is effected by introducing and expressing in a plant a nucleic acid encoding an AUX/IAA polypeptide.
 - Method according to any one of items 1 to 4, wherein said nucleic acid encoding an AUX/IAA polypeptide encodes any one of the proteins listed in Table A4 or in Table A5 or is a portion of such a nucleic acid, or a nucleic acid capable of hybridising with such a nucleic acid.
 - Method according to any one of items 1 to 5, wherein said nucleic acid sequence encodes an orthologue or paralogue of any of the proteins given in Table A4 or in Table A5.
 - Method according to any preceding item, wherein said enhanced yield-related traits comprise increased yield, preferably increased biomass and/or increased seed yield relative to control plants.
 - Method according to any one of items 1 to 7, wherein said enhanced yield-related traits are obtained under non-stress conditions.
 - Method according to any one of items 3 to 8, wherein said nucleic acid is operably linked to a constitutive promoter, preferably to a GOS2 promoter, most preferably to a GOS2 promoter from rice.
 - Method according to any one of items 1 to 9, wherein said nucleic acid encoding an AUX/IAA polypeptide is of plant

- origin, preferably from a monocotyledonous plant, further preferably from the family Poaceae, more preferably from the genus *Oryza*, most preferably from *Oryza sativa*.
11. Plant or part thereof, including seeds, obtainable by a method according to any one of items 1 to 10, wherein said plant or part thereof comprises a recombinant nucleic acid encoding an AUX/IAA polypeptide.
 12. Construct comprising:
 - (i) nucleic acid encoding an AUX/IAA polypeptide as defined in items 1 or 2;
 - (ii) one or more control sequences capable of driving expression of the nucleic acid sequence of (a); and optionally
 - (iii) a transcription termination sequence.
 13. Construct according to item 12, wherein one of said control sequences is a constitutive promoter, preferably a GOS2 promoter, most preferably a GOS2 promoter from rice.
 14. Use of a construct according to item 12 or 13 in a method for making plants having increased yield, particularly increased biomass and/or increased seed yield relative to control plants.
 15. Plant, plant part or plant cell transformed with a construct according to item 12 or 13.
 16. Method for the production of a transgenic plant having increased yield, particularly increased biomass and/or increased seed yield relative to control plants, comprising:
 - (i) introducing and expressing in a plant a nucleic acid encoding an AUX/IAA polypeptide as defined in item 1 or 2; and
 - (ii) cultivating the plant cell under conditions promoting plant growth and development.
 17. Transgenic plant having increased yield, particularly increased biomass and/or increased seed yield, relative to control plants, resulting from modulated expression of a nucleic acid encoding an AUX/IAA polypeptide as defined in item 1 or 2, or a transgenic plant cell derived from said transgenic plant.
 18. Transgenic plant according to item 11, 15 or 17, or a transgenic plant cell derived thereof, wherein said plant is a crop plant or a monocot or a cereal, such as rice, maize, wheat, barley, millet, rye, triticale, sorghum emmer, spelt, *secale*, einkorn, teff, milo and oats.
 19. Harvestable parts of a plant according to item 18, wherein said harvestable parts are preferably shoot biomass and/or seeds.
 20. Products derived from a plant according to item 18 and/or from harvestable parts of a plant according to item 19.
 21. Use of a nucleic acid encoding an AUX/IAA polypeptide in increasing yield, particularly in increasing seed yield and/or shoot biomass in plants, relative to control plants.

DESCRIPTION OF FIGURES

The present invention will now be described with reference to the following figures in which:

FIG. 1 represents a multiple alignment of ASPAT polypeptides. Sequences shown are 100 (SEQ ID NO: 100); 102 (SEQ ID NO: 102); 110 (SEQ ID NO: 110); 76 (SEQ ID NO: 76); 112 (SEQ ID NO: 112); 114 (SEQ ID NO: 114); 118 (SEQ ID NO: 118); 170 (SEQ ID NO: 170); 172 (SEQ ID NO: 172); 174 (SEQ ID NO: 174); 176 (SEQ ID NO: 176); 44 (SEQ ID NO: 44); 2 (SEQ ID NO: 4); 4 (SEQ ID NO: 2); 24 (SEQ ID NO: 24); 6 (SEQ ID NO: 6); 14 (SEQ ID NO: 14); 8 (SEQ ID NO: 8); 50 (SEQ ID NO: 50); 54 (SEQ ID NO: 54); and 62 (SEQ ID NO: 62).

FIG. 2 shows a phylogenetic tree of ASPAT polypeptides.

FIG. 3 represents the binary vector used for increased expression in *Oryza saliva* of an ASPAT-encoding nucleic acid under the control of a rice GOS2 promoter (pGOS2) or of a rice PR promoter.

FIG. 4 represents the phylogenetic relationship among MYB DNA binding domain polypeptides from *Arabidopsis thaliana* and from other plants, based upon amino acid sequence (according to Stracke et al. (2004) Current Opinion in Plant Biology 2001, 4:447-456). The MYB polypeptides were clustered using PHYLIP, and motifs were detected using MEME. Polypeptides useful in performing the methods of the invention cluster with MYB91, circled and marked by a black arrow. Motifs shown are WFKHLESELGLEExDNQQQ (SEQ ID NO: 818); YASSxxNI SE ID NO: 819 SL[F/I]EK-WLF[D/E] (SEQ ID NO: 820); IDxSFw--MxFWFD (SEQ ID NO: 821); DExWRLxxT (SEQ ID NO: 822); KPRPR[S/T]F (SEQ ID NO: 823); WVxxpxFELSxL (SEQ ID NO: 824); GRTxRSxMK (SEQ ID NO: 825); PRLDLLD (SEQ ID NO: 826); IQMExDPxTH (SEQ ID NO: 827); LNL[E/D]L (SEQ ID NO: 828); QxxAAAx (SEQ ID NO: 829); KxQLx-HxMxQ (SEQ ID NO: 830); DDxxSDSxWK (SEQ ID NO: 831); [L/F]LN[K/R]VA (SEQ ID NO: 832); AQWESARxx-AExRLxRES (SEQ ID NO: 833); PxLxFSEW (SEQ ID NO: 834); WxPRL (SEQ ID NO: 835); GLP[L/V]YP (SEQ ID NO: 836); FxDfL (SEQ ID NO: 837); TGLYMSPxSP (SEQ ID NO: 838); GxFMxV (SEQ ID NO: 839); VQEMixx-EVRSYM (SEQ ID NO: 840); LxxYlxx[I/V]N[N/D] (SEQ ID NO: 841); PxLxFSEW (SEQ ID NO: 842); [W]-X(20)-[W]-x(19)-[W]-x(12)-[F]-x(18)-[W]-x(18)-[W] (SEQ ID NO: 843); and [W]-X(19)-[W]-x(21)-[W]x(12)-[L]-x(18)-[W]-x(18)-[W] (SEQ ID NO: 844).

FIG. 5 shows a ClustalW 1.81 multiple sequence alignment of the MYB91 polypeptides from Table A2. Two MYB DNA binding domains with an InterPro accession number IPR014778, a MYB transcription factor with an InterPro accession number IPR015495, and a C-terminal Conserved Domain, are marked with X's below the consensus sequence. Sequences shown are Poptr_MYB91 (SEQ ID NO: 221); Medtr_MYB91_PHAN_ (SEQ ID NO: 251); Pissa_MYB91 (SEQ ID NO: 259); Glyma_MYB91_PHANa_ (SEQ ID NO: 237); Glyma_MYB91_PHANb_ (SEQ ID NO: 239); Lotco_MYB91_PHANb_ (SEQ ID NO: 245); Lotco_MYB91_PHANa_ (SEQ ID NO: 243); Eucgr_MYB91 (SEQ ID NO: 235); Maldo_MYB91 (SEQ ID NO: 249); Lycex_MYB91 (SEQ ID NO: 247); Soltu_MYB91 (SEQ ID NO: 261); Nicta_MYB91 (SEQ ID NO: 255); Vitvi_MYB91 (SEQ ID NO: 263); Goshi_MYB91 (SEQ ID NO: 241); Aqufo_MYB91 (SEQ ID NO: 225); Esc-ca_MYB91 (SEQ ID NO: 233); Arath_AS1_MYB91 (SEQ ID NO: 227); Carhi_MYB91 (SEQ ID NO: 231); Brana_MYB91 (SEQ ID NO: 229); Antma_MYB91 (SEQ ID NO: 223); Orysa_MYB91 (SEQ ID NO: 257); Zeama_MYB91_RS2_ (SEQ ID NO: 265); and Moral_MYB91 (SEQ ID NO: 253).

FIG. 6 shows the binary vector for increased expression in *Oryza sativa* plants of a nucleic acid sequence encoding a MYB91 polypeptide under the control of a promoter functioning in plants.

FIG. 7 represents the domain structure of SEQ ID NO: 276 with the GASA domain PF02704 indicated in bold. The putative secretion signal peptide (amino acid 1-24) is underlined.

FIG. 8 represents a multiple alignment of various GASA proteins. The motifs 4 to 12 or other motifs can be deduced herefrom. Sequences shown are Os05g0432200 (SEQ ID NO: 304); AK110640 (SEQ ID NO: 308); TA53297_4565 (SEQ ID NO: 345); TA52915_4565 (SEQ ID NO: 355);

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scaff_41.75 (SEQ ID NO: 323); TA52374_4081 (SEQ ID NO: 333); TA5035_4679 (SEQ ID NO: 297); Os09g0414900 (SEQ ID NO: 305); GASA6 (SEQ ID NO: 296); scaff_XVII.377 (SEQ ID NO: 316); TA56938_4081 (SEQ ID NO: 336); GASA4 (SEQ ID NO: 295); Os05g0376800 (SEQ ID NO: 300); scaff_VI.397 (SEQ ID NO: 315); scaff_I.1483 (SEQ ID NO: 319); BG128975 (SEQ ID NO: 332); BG130916 (SEQ ID NO: 337); TA52635_4081 (SEQ ID NO: 338); TA5923_4679 (SEQ ID NO: 298); Os06g0266800 (SEQ ID NO: 309); TA100367_4565 (SEQ ID NO: 348); CA725087 (SEQ ID NO: 343); TA77646_4565 (SEQ ID NO: 359); TA92393_4565 (SEQ ID NO: 349); CK153563 (SEQ ID NO: 353); BI208422 (SEQ ID NO: 331); TA37180_4081 (SEQ ID NO: 334); scaff_II.2328 (SEQ ID NO: 325); scaff_II.2330 (SEQ ID NO: 313); GASA5 (SEQ ID NO: 294); GASA12 (SEQ ID NO: 293); Os10g0115550 (SEQ ID NO: 302); TA101332_4565 (SEQ ID NO: 346); TA56201_4081 (SEQ ID NO: 341); AJ785329 (SEQ ID NO: 342); AK105729 (SEQ ID NO: 303); Os03g0760800 (SEQ ID NO: 310); TA66036_4565 (SEQ ID NO: 347); BM136027 (SEQ ID NO: 350); CA705831 (SEQ ID NO: 351); CA593033 (SEQ ID NO: 352); TA66038_4565 (SEQ ID NO: 354); CD899399 (SEQ ID NO: 358); Os03g0607200 (SEQ ID NO: 306); scaff_IX.735 (SEQ ID NO: 314); scaff_I.2410 (SEQ ID NO: 318); Pop_GASA_ (SEQ ID NO: 291); scaff_40.379 (SEQ ID NO: 322); TA45751_4081 (SEQ ID NO: 328); scaff_205.30 (SEQ ID NO: 311); TA69823_4565 (SEQ ID NO: 344); TA69821_4565 (SEQ ID NO: 356); Os07g0592000 (SEQ ID NO: 307); Os04g0465300 (SEQ ID NO: 301); scaff_II.204 (SEQ ID NO: 312); scaff_II.202 (SEQ ID NO: 317); TA35962_4081 (SEQ ID NO: 330); scaff_II.203 (SEQ ID NO: 326); BE353147 (SEQ ID NO: 335); TA41886_4081 (SEQ ID NO: 339); scaff_XII.704 (SEQ ID NO: 321); scaff_XV.507 (SEQ ID NO: 324); TA48119_4081 (SEQ ID NO: 329); Mt_GASA (SEQ ID NO: 292); scaff_I.1926 (SEQ ID NO: 320); scaff_XIX.758 (SEQ ID NO: 327); TA36295_4081 (SEQ ID NO: 340); TA95153_4565 (SEQ ID NO: 357); and TA51752_4565 (SEQ ID NO: 360).

FIG. 9 shows a phylogenetic tree of *Arabidopsis* GASA proteins (Roxrud et al. 2007). Starting from a multiple alignment with ClustalW (Thompson et al., Nucleic Acids Res. 22, 4673-4680, 1994), a neighbour-joining phylogenetic tree was obtained using the PAUP v.4.0 software (paup.csit.fsu.edu), and statistical confidence was calculated by bootstrap analysis with 1,000 resamplings.

FIG. 10 represents the binary vector for increased expression in *Oryza sativa* of a GASA-encoding nucleic acid under the control of a rice GOS2 promoter (pGOS2).

FIG. 11 represents a multiple alignment of AUX/IAA polypeptides.

FIG. 12 represents the binary vector used for increased expression in *Oryza sativa* of an AUX/IAA encoding nucleic acid under the control of a rice GOS2 promoter (pGOS2).

FIG. 13 represents the domain structure of SEQ ID NO: 738 with the AUX/IAA domain in bold and the conserved motifs underlined.

FIG. 14 represents a multiple alignment of IAA14-like protein sequences. Sequences shown are AT3G23050.1 (SEQ ID NO: 748); AT3G23050.2 (SEQ ID NO: 749); AT4G14550.1 (SEQ ID NO: 738); Mt_TA20354 (SEQ ID NO: 752); Pt_566151 (SEQ ID NO: 750); Pt_720961 (SEQ ID NO: 751); SI_TA40922 (SEQ ID NO: 753); AT1G04250.1 (SEQ ID NO: 754); Mt_TA27011 (SEQ ID NO: 760); Mt_TA22814 (SEQ ID NO: 761); Pt_643213 (SEQ ID NO: 762); SI_TA48108 (SEQ ID NO: 759); Os_CB657009 (SEQ ID NO: 755); Os_TA41733 (SEQ ID NO: 756);

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AT3G04730.1 (SEQ ID NO: 758); Mt_TA20951 (SEQ ID NO: 757); Mt_TA25400 (SEQ ID NO: 779); Pt_584053 (SEQ ID NO: 781); Pt_711734 (SEQ ID NO: 780); AT4G29080.1 (SEQ ID NO: 778); Mt_TA23062 (SEQ ID NO: 782); AT3G23030.1 (SEQ ID NO: 763); AT4G14560.1 (SEQ ID NO: 764); SI_TA38817 (SEQ ID NO: 766); SI_TA43058 (SEQ ID NO: 767); Pt_726443 (SEQ ID NO: 768); Pt_564913 (SEQ ID NO: 769); Mt_TA20557 (SEQ ID NO: 772); Pt_831610 (SEQ ID NO: 770); Pt_798526 (SEQ ID NO: 771); Mt_TA31746 (SEQ ID NO: 776); Pt_823671 (SEQ ID NO: 774); Pt_595419 (SEQ ID NO: 775); Mt_TA20558 (SEQ ID NO: 773); AT1G04240.1 (SEQ ID NO: 765); and SI_TA42190 (SEQ ID NO: 777).

FIG. 15 shows a neighbour-joining tree of *Arabidopsis* IAA proteins (Remington et al., 2004). SEQ ID NO: 738 is represented by IAA14 in Group A and IAA14-like proteins preferably cluster in this Group A.

FIG. 16 represents the binary vector used for increased expression in *Oryza sativa* of an IAA14-like-encoding nucleic acid under the control of a rice HMGP promoter (pHMGP).

EXAMPLES

The present invention will now be described with reference to the following examples, which are by way of illustration alone. The following examples are not intended to completely define or otherwise limit the scope of the invention.

DNA manipulation: unless otherwise stated, recombinant DNA techniques are performed according to standard protocols described in (Sambrook (2001) Molecular Cloning: a laboratory manual, 3rd Edition Cold Spring Harbor Laboratory Press, CSH, New York) or in Volumes 1 and 2 of Ausubel et al. (1994), Current Protocols in Molecular Biology, Current Protocols. Standard materials and methods for plant molecular work are described in Plant Molecular Biology Labfax (1993) by R. D. D. Croy, published by BIOS Scientific Publications Ltd (UK) and Blackwell Scientific Publications (UK).

Example 1

Identification of Sequences Related to the Nucleic Acid Sequence Used in the Methods of the Invention

Sequences (full length cDNA, ESTs or genomic) related to the nucleic acid sequence used in the methods of the present invention were identified amongst those maintained in the Entrez Nucleotides database at the National Center for Biotechnology Information (NCBI) using database sequence search tools, such as the Basic Local Alignment Tool (BLAST) (Altschul et al. (1990) J. Mol. Biol. 215:403-410; and Altschul et al. (1997) Nucleic Acids Res. 25:3389-3402). The program is used to find regions of local similarity between sequences by comparing nucleic acid or polypeptide sequences to sequence databases and by calculating the statistical significance of matches. For example, the polypeptide encoded by the nucleic acid used in the present invention was used for the TBLASTN algorithm, with default settings and the filter to ignore low complexity sequences set off. The output of the analysis was viewed by pairwise comparison, and ranked according to the probability score (E-value), where the score reflect the probability that a particular alignment occurs by chance (the lower the E-value, the more significant the hit). In addition to E-values, comparisons were also scored by percentage identity. Percentage identity refers to the number of identical nucleotides (or amino acids)

between the two compared nucleic acid (or polypeptide) sequences over a particular length. In some instances, the default parameters may be adjusted to modify the stringency of the search. For example the E-value may be increased to show less stringent matches. This way, short nearly exact matches may be identified.

1.1. Aspartate AminoTransferase (ASPAT)

Table A1 provides a list of nucleic acid sequences related to the nucleic acid sequence used in the methods of the present invention.

TABLE A1

Examples of ASPAT polypeptides:			
Reference number	Name	Nucleic acid SEQ ID NO:	Amino acid SEQ ID NO:
1	O. sativa_Os01g0760600	1	2
1	O. sativa_Os01g0760600-truncated	3	4
1	A. thaliana_AT5G19550	5	6
1	A. thaliana_AT5G11520	7	8
1	A. thaliana_AT4G31990	9	10
6	A. thaliana_AT1G62800	11	12
7	B. napus_TA23207	13	14
8	B. napus_TA23768	15	16
9	C. sinensis_TA12564	17	18
10	C. solstitialis_TA659	19	20
11	G. hirsutum_TA23799	21	22
12	G. max_AF034210	23	24
13	G. raimondii_TA9413	25	26
14	H. annuus_TA8926	27	28
15	H. paradoxus_TA2606	29	30
16	J. regia_TA762	31	32
17	L. japonicus_TA1537	33	34
18	L. perennis_TA512	35	36
19	L. perennis_TA605	37	38
20	N. tabacum_TA13125	39	40
21	P. glauca_TA15326	41	42
22	P. patens_I36815	43	44
23	P. persica_TA3273	45	46
24	P. sitchensis_TA22265	47	48
25	P. trichocarpa_819551	49	50
26	P. trifoliata_TA8305	51	52
27	S. lycopersicum_TA38054	53	54
28	S. officinarum_TA26595	55	56
29	T. aestivum_TA52678	57	58
30	V. carteri_82929	59	60
31	V. vinifera_GSVIVT00016723001	61	62
32	V. vinifera_GSVIVT00032463001	63	64
33	Z. mays_TA9042	65	66
34	C. reinhardtii_186959	67	68
35	C. solstitialis_TA2275	69	70
36	C. tinctorius_TA12	71	72
37	G. hirsutum_TA24406	73	74
38	G. max_TA61768	75	76
39	G. raimondii_TA9928	77	78
40	H. exilis_TA1663	79	80
41	H. vulgare_BPS_7992	81	82
42	L. japonicus_TA1466	83	84
43	M. polymorpha_TA825	85	86
44	N. tabacum_TA13015	87	88
45	O. sativa_Os02g0797500	89	90
46	P. glauca_TA14780	91	92
47	P. patens_I02134	93	94
48	P. sitchensis_TA20968	95	96
49	P. taeda_TA6616	97	98
50	P. trichocarpa_654206	99	100
51	P. trichocarpa_835828	101	102
52	P. vulgaris_TA4043	103	104
53	S. tuberosum_TA23192	105	106
54	V. carteri_81153	107	108
55	V. vinifera_GSVIVT00032723001	109	110
56	Z. mays_TA10886	111	112
57	A. thaliana_AT2G30970	113	114
58	C. sinensis_TA15250	115	116

TABLE A1-continued

Examples of ASPAT polypeptides:			
Reference number	Name	Nucleic acid SEQ ID NO:	Amino acid SEQ ID NO:
59	G. max_TA50178	117	118
60	G. raimondii_TA9985	119	120
61	H. vulgare_TA32835	121	122
62	H. vulgare_TA36301	123	124
63	O. lucimarinus_31597	125	126
64	O. sativa_Os02g0236000	127	128
65	O. sativa_Os06g0548000	129	130
66	O. taurii_32764	131	132
67	P. patens_169868	133	134
68	P. sitchensis_TA23007	135	136
69	P. taeda_TA7145	137	138
70	V. vinifera_GSVIVT00018772001	139	140
71	V. vinifera_GSVIVT00037462001	141	142
72	A. anophagefferens_21970	143	144
73	A. thaliana_AT2G22250.2	145	146
74	B. napus_BPS_9867	147	148
75	C. reinhardtii_118364	149	150
76	G. hirsutum_TA27281	151	152
77	G. max_BPS_36342	153	154
78	H. vulgare_TA28738	155	156
79	M. domestica_TA26867	157	158
80	N. tabacum_TA15308	159	160
81	O. basilicum_TA1043	161	162
82	O. sativa_Os01g0871300	163	164
83	P. patens_127152	165	166
84	P. pinaster_TA3616_71647	167	168
85	P. trichocarpa_scaff_V.183	169	170
86	P. trichocarpa_scaff_VII.574	171	172
87	S. lycopersicum_TA37592	173	174
88	S. tuberosum_TA27739	175	176
89	T. aestivum_TA71539	177	178
90	V. carteri_103084	179	180
91	V. vinifera_GSVIVT00019453001	181	182
92	Z. mays_BPS_26636	183	184
93	Z. mays_BPS_4233	185	186

In some instances, related sequences are tentatively been assembled and publicly disclosed by research institutions, such as The Institute for Genomic Research (TIGR; beginning with TA). The Eukaryotic Gene Orthologs (EGO) database may be used to identify such related sequences, either by keyword search or by using the BLAST algorithm with the nucleic acid sequence or polypeptide sequence of interest. On other instances, special nucleic acid sequence databases have been from particular organisms, such as those maintained by the Joint Genome Institute, like the poplar genome sequences have been screened.

Further, access to proprietary databases, has allowed the identification of other nucleic acid and polypeptide sequences using the Blast algorithm as described above.

1.2. MYB91 like transcription factor (MYB91)

Table A2 provides a list of nucleic acid sequences related to the nucleic acid sequence used in the methods of the present invention.

TABLE A2

Examples of MYB91 polypeptide sequences, and encoding nucleic acid sequences			
Name	Public database accession number	Nucleic acid SEQ ID NO:	Polypeptide SEQ ID NO:
60			
65			
60	Poptr_MYB91	NA	220
65	Antma_MYB91 (PHAN)	AJ005586	222
			223

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TABLE A2-continued

Examples of MYB91 polypeptide sequences, and encoding nucleic acid sequences			
Name	Public database accession number	Nucleic acid SEQ ID NO:	Polypeptide SEQ ID NO:
Aqufo_MYB91	DR919410	224	225
	DR919310		
Arath_MYB91 (AS1)	AT2G37630	226	227
Branan_MYB91	BN06MC30974_51405116	228	229
	@30844#1		
Carhi_MYB91	DQ512733	230	231
Escca_MYB91	AY228766	232	233
Eucgr_MYB91	BD376532	234	235
Glyma_MYB91 (PHANa)	AY790252	236	237
Glyma_MYB91 (PHANb)	AY790253	238	239
Goshi_MYB91	DT554770	240	241
	DW499296		
Lotco_MYB91 (PHANa)	AY790244	242	243
Lotco_MYB91 (PHANb)	AY790245	244	245
Lyces_MYB91	AF148934	246	247
Maldo_MYB91	DQ074473	248	249
Medtr_MYB91 PHAN	DQ468322	250	251
Moral_MYB91 PHAN1	EF408927	252	253
Nicta_MYB91	AY559043	254	255
Orysa_MYB91	Os12g0572000	256	257
	NM_001073621		
Pissa_MYB91 (PHAN1)	AF299140.2	258	259
Soltu_MYB91	CK274535	260	261
Vitvi_MYB91	AM474349	262	263
Zeama_MYB91 (RS2)	AF126489	264	265
Horvu_MYB91 partial	BF617675.2	266	267
	BG343686.1		

In some instances, related sequences have tentatively been assembled and publicly disclosed by research institutions, such as The Institute for Genomic Research (TIGR; beginning with TA). The Eukaryotic Gene Orthologs (EGO) database may be used to identify such related sequences, either by keyword search or by using the BLAST algorithm with the nucleic acid sequence or polypeptide sequence of interest. On other instances, special nucleic acid sequence databases have been created for particular organisms, such as by the Joint Genome Institute. Further, access to proprietary databases, has allowed the identification of novel nucleic acid and polypeptide sequences.

1.3. Gibberellic Acid-Stimulated *Arabidopsis* (GASA)

Table A3 provides a list of nucleic acid sequences related to the nucleic acid sequence used in the methods of the present invention.

TABLE A3

Examples of GASA polypeptides:			
Name	Polypeptide SEQ ID NO	Nucleic acid SEQ ID NO	
Le_GASA growth induced	276	275	
Pop_GASA growth regulated	291	361	
Mt_GASA growth regulated	292	362	
GASA12 At2g30810	293	363	
GASA5 At3g02885	294	364	
GASA4 At5g15230	295	365	

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TABLE A3-continued

Examples of GASA polypeptides:			
Name	Polypeptide SEQ ID NO	Nucleic acid SEQ ID NO	
GASA6 At1g74670	296	366	
TA5035_4679#1	297	367	
TA5923_4679#1	298	368	
TA3842_4679#1	299	369	
Os05g0376800#1	300	370	
Os04g0465300#1	301	371	
Os10g0115550#1	302	372	
AK105729#1	303	373	
Os05g0432200#1	304	374	
Os09g0414900#1	305	375	
Os03g0607200#1	306	376	
Os07g0592000#1	307	377	
AK110640#1	308	378	
Os06g0266800#1	309	379	
Os03g0760800#1	310	380	
scaff_205.30#1	311	381	
scaff_IL204#1	312	382	
scaff_IL2330#1	313	383	
scaff_IX.735#1	314	384	
scaff_VI.397#1	315	385	
scaff_XVII.377#1	316	386	
scaff_IL202#1	317	387	
scaff_I.2410#1	318	388	
scaff_I.1483#1	319	389	
scaff_I.1926#1	320	390	
scaff_XII.704#1	321	391	
scaff_40.379#1	322	392	
scaff_41.75#1	323	393	
scaff_XV.507#1	324	394	
scaff_IL2328#1	325	395	
scaff_IL203#1	326	396	
scaff_XIX.758#1	327	397	
TA45751_4081#1	328	398	
TA48119_4081#1	329	399	
TA35962_4081#1	330	400	
BI208422#1	331	401	
BG128975#1	332	402	
TA52374_4081#1	333	403	
TA37180_4081#1	334	404	
BE353147#1	335	405	
TA56938_4081#1	336	406	
BG130916#1	337	407	
TA52635_4081#1	338	408	
TA41886_4081#1	339	409	
TA36295_4081#1	340	410	
TA56201_4081#1	341	411	
AJ785329#1	342	412	
CA725087#1	343	413	
TA69823_4565#1	344	414	
TA53297_4565#1	345	415	
TA101332_4565#1	346	416	
TA66036_4565#1	347	417	
TA100367_4565#1	348	418	
TA92393_4565#1	349	419	
BM136027#1	350	420	
CA705831#1	351	421	
CA593033#1	352	422	
CK153563#1	353	423	
TA66038_4565#1	354	424	
TA52915_4565#1	355	425	
TA69821_4565#1	356	426	
TA95153_4565#1	357	427	
CD899399#1	358	428	
TA77646_4565#1	359	429	
TA51752_4565#1	360	430	

In some instances, related sequences have tentatively been assembled and publicly disclosed by research institutions, such as The Institute for Genomic Research (TIGR). The Eukaryotic Gene Orthologs (EGO) database may be used to identify such related sequences, either by keyword search or by using the BLAST algorithm with the nucleic acid or polypeptide sequence of interest.

1.4. Auxin/Indoleacetic Acid Genes (AUX/IAA)

Nucleic acid name	Nucleic Acid SEQ ID NO:	Polypeptide name	Polypeptide SEQ ID NO:
seqidno01; DNA; <i>Oryza sativa</i>	431	seqidno02; PRT; <i>Oryza sativa</i>	432
seqidno1; DNA; <i>Arabidopsis thaliana</i>	433	seqidno2; PRT; <i>Arabidopsis thaliana</i>	434
seqidno3; DNA; <i>Arabidopsis thaliana</i>	435	seqidno4; PRT; <i>Arabidopsis thaliana</i>	436
seqidno5; DNA; <i>Arabidopsis thaliana</i>	437	seqidno6; PRT; <i>Arabidopsis thaliana</i>	438
seqidno7; DNA; <i>Arabidopsis thaliana</i>	439	seqidno8; PRT; <i>Arabidopsis thaliana</i>	440
seqidno9; DNA; <i>Arabidopsis thaliana</i>	441	seqidno10; PRT; <i>Arabidopsis thaliana</i>	442
seqidno11; DNA; <i>Arabidopsis thaliana</i>	443	seqidno12; PRT; <i>Arabidopsis thaliana</i>	444
seqidno13; DNA; <i>Arabidopsis thaliana</i>	445	seqidno14; PRT; <i>Arabidopsis thaliana</i>	446
seqidno15; DNA; <i>Arabidopsis thaliana</i>	447	seqidno16; PRT; <i>Arabidopsis thaliana</i>	448
seqidno17; DNA; <i>Arabidopsis thaliana</i>	449	seqidno18; PRT; <i>Arabidopsis thaliana</i>	450
seqidno19; DNA; <i>Arabidopsis thaliana</i>	451	seqidno20; PRT; <i>Arabidopsis thaliana</i>	452
seqidno21; DNA; <i>Arabidopsis thaliana</i>	453	seqidno22; PRT; <i>Arabidopsis thaliana</i>	454
seqidno23; DNA; <i>Arabidopsis thaliana</i>	455	seqidno24; PRT; <i>Arabidopsis thaliana</i>	456
seqidno25; DNA; <i>Arabidopsis thaliana</i>	457	seqidno26; PRT; <i>Arabidopsis thaliana</i>	458
seqidno27; DNA; <i>Arabidopsis thaliana</i>	459	seqidno28; PRT; <i>Arabidopsis thaliana</i>	460
seqidno29; DNA; <i>Arabidopsis thaliana</i>	461	seqidno30; PRT; <i>Arabidopsis thaliana</i>	462
seqidno31; DNA; <i>Arabidopsis thaliana</i>	463	seqidno32; PRT; <i>Arabidopsis thaliana</i>	464
seqidno33; DNA; <i>Arabidopsis thaliana</i>	465	seqidno34; PRT; <i>Arabidopsis thaliana</i>	466
seqidno35; DNA; <i>Arabidopsis thaliana</i>	467	seqidno36; PRT; <i>Arabidopsis thaliana</i>	468
seqidno37; DNA; <i>Arabidopsis thaliana</i>	469	seqidno38; PRT; <i>Arabidopsis thaliana</i>	470
seqidno39; DNA; <i>Arabidopsis thaliana</i>	471	seqidno40; PRT; <i>Arabidopsis thaliana</i>	472
seqidno41; DNA; <i>Arabidopsis thaliana</i>	473	seqidno42; PRT; <i>Arabidopsis thaliana</i>	474
seqidno43; DNA; <i>Arabidopsis thaliana</i>	475	seqidno44; PRT; <i>Arabidopsis thaliana</i>	476
seqidno45; DNA; <i>Arabidopsis thaliana</i>	477	seqidno46; PRT; <i>Arabidopsis thaliana</i>	478
seqidno47; DNA; <i>Arabidopsis thaliana</i>	479	seqidno48; PRT; <i>Arabidopsis thaliana</i>	480
seqidno49; DNA; <i>Arabidopsis thaliana</i>	481	seqidno50; PRT; <i>Arabidopsis thaliana</i>	482
seqidno51; DNA; <i>Arabidopsis thaliana</i>	483	seqidno52; PRT; <i>Arabidopsis thaliana</i>	484
seqidno53; DNA; <i>Arabidopsis thaliana</i>	485	seqidno54; PRT; <i>Arabidopsis thaliana</i>	486
seqidno55; DNA; <i>Arabidopsis thaliana</i>	487	seqidno56; PRT; <i>Arabidopsis thaliana</i>	488
seqidno57; DNA; <i>Arabidopsis thaliana</i>	489	seqidno58; PRT; <i>Arabidopsis thaliana</i>	490
seqidno59; DNA; <i>Arabidopsis thaliana</i>	491	seqidno60; PRT; <i>Arabidopsis thaliana</i>	492
seqidno61; DNA; <i>Arabidopsis thaliana</i>	493	seqidno62; PRT; <i>Arabidopsis thaliana</i>	494
seqidno63; DNA; <i>Arabidopsis thaliana</i>	495	seqidno64; PRT; <i>Arabidopsis thaliana</i>	496
seqidno65; DNA; <i>Arabidopsis thaliana</i>	497	seqidno66; PRT; <i>Arabidopsis thaliana</i>	498
seqidno67; DNA; <i>Arabidopsis thaliana</i>	499	seqidno68; PRT; <i>Arabidopsis thaliana</i>	500
seqidno69; DNA; <i>Oryza sativa</i>	501	seqidno70; PRT; <i>Oryza sativa</i>	502
seqidno71; DNA; <i>Oryza sativa</i>	503	seqidno72; PRT; <i>Oryza sativa</i>	504
seqidno73; DNA; <i>Oryza sativa</i>	505	seqidno74; PRT; <i>Oryza sativa</i>	506
seqidno75; DNA; <i>Oryza sativa</i>	507	seqidno76; PRT; <i>Oryza sativa</i>	508
seqidno77; DNA; <i>Oryza sativa</i>	509	seqidno78; PRT; <i>Oryza sativa</i>	510
seqidno79; DNA; <i>Oryza sativa</i>	511	seqidno80; PRT; <i>Oryza sativa</i>	512
seqidno81; DNA; <i>Oryza sativa</i>	513	seqidno82; PRT; <i>Oryza sativa</i>	514
seqidno83; DNA; <i>Oryza sativa</i>	515	seqidno84; PRT; <i>Oryza sativa</i>	516
seqidno85; DNA; <i>Oryza sativa</i>	517	seqidno86; PRT; <i>Oryza sativa</i>	518
seqidno87; DNA; <i>Oryza sativa</i>	519	seqidno88; PRT; <i>Oryza sativa</i>	520
seqidno89; DNA; <i>Oryza sativa</i>	521	seqidno90; PRT; <i>Oryza sativa</i>	522
seqidno91; DNA; <i>Oryza sativa</i>	523	seqidno92; PRT; <i>Oryza sativa</i>	524
seqidno93; DNA; <i>Oryza sativa</i>	525	seqidno94; PRT; <i>Oryza sativa</i>	526
seqidno95; DNA; <i>Oryza sativa</i>	527	seqidno96; PRT; <i>Oryza sativa</i>	528
seqidno97; DNA; <i>Oryza sativa</i>	529	seqidno98; PRT; <i>Oryza sativa</i>	530
seqidno99; DNA; <i>Oryza sativa</i>	531	seqidno100; PRT; <i>Oryza sativa</i>	532
seqidno101; DNA; <i>Oryza sativa</i>	533	seqidno102; PRT; <i>Oryza sativa</i>	534
seqidno103; DNA; <i>Oryza sativa</i>	535	seqidno104; PRT; <i>Oryza sativa</i>	536
seqidno105; DNA; <i>Oryza sativa</i>	537	seqidno106; PRT; <i>Oryza sativa</i>	538
seqidno107; DNA; <i>Oryza sativa</i>	539	seqidno108; PRT; <i>Oryza sativa</i>	540
seqidno109; DNA; <i>Oryza sativa</i>	541	seqidno110; PRT; <i>Oryza sativa</i>	542
seqidno111; DNA; <i>Oryza sativa</i>	543	seqidno112; PRT; <i>Oryza sativa</i>	544
seqidno113; DNA; <i>Oryza sativa</i>	545	seqidno114; PRT; <i>Oryza sativa</i>	546
seqidno115; DNA; <i>Oryza sativa</i>	547	seqidno116; PRT; <i>Oryza sativa</i>	548
seqidno117; DNA; <i>Oryza sativa</i>	549	seqidno118; PRT; <i>Oryza sativa</i>	550
seqidno119; DNA; <i>Oryza sativa</i>	551	seqidno120; PRT; <i>Oryza sativa</i>	552
seqidno121; DNA; <i>Oryza sativa</i>	553	seqidno122; PRT; <i>Oryza sativa</i>	554
seqidno123; DNA; <i>Oryza sativa</i>	555	seqidno124; PRT; <i>Oryza sativa</i>	556
seqidno125; DNA; <i>Oryza sativa</i>	557	seqidno126; PRT; <i>Oryza sativa</i>	558
seqidno127; DNA; <i>Oryza sativa</i>	559	seqidno128; PRT; <i>Oryza sativa</i>	560
seqidno129; DNA; <i>Oryza sativa</i>	561	seqidno130; PRT; <i>Oryza sativa</i>	562
seqidno131; DNA; <i>Oryza sativa</i>	563	seqidno132; PRT; <i>Oryza sativa</i>	564
seqidno133; DNA; <i>Oryza sativa</i>	565	seqidno134; PRT; <i>Oryza sativa</i>	566
seqidno135; DNA; <i>Oryza sativa</i>	567	seqidno136; PRT; <i>Oryza sativa</i>	568
seqidno137; DNA; <i>Oryza sativa</i>	569	seqidno138; PRT; <i>Oryza sativa</i>	570
seqidno139; DNA; <i>Oryza sativa</i>	571	seqidno140; PRT; <i>Oryza sativa</i>	572
seqidno141; DNA; <i>Oryza sativa</i>	573	seqidno142; PRT; <i>Oryza sativa</i>	574
seqidno143; DNA; <i>Oryza sativa</i>	575	seqidno144; PRT; <i>Oryza sativa</i>	576
seqidno145; DNA; <i>Oryza sativa</i>	577	seqidno146; PRT; <i>Oryza sativa</i>	578
seqidno147; DNA; <i>Oryza sativa</i>	579	seqidno148; PRT; <i>Oryza sativa</i>	580
seqidno149; DNA; <i>Oryza sativa</i>	581	seqidno150; PRT; <i>Oryza sativa</i>	582

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Nucleic acid name	Nucleic Acid SEQ ID NO:	Polypeptide name	Polypeptide SEQ ID NO:
seqidno151; DNA; <i>Oryza sativa</i>	583	seqidno152; PRT; <i>Oryza sativa</i>	584
seqidno153; DNA; <i>Oryza sativa</i>	585	seqidno154; PRT; <i>Oryza sativa</i>	586
seqidno155; DNA; <i>Oryza sativa</i>	587	seqidno156; PRT; <i>Oryza sativa</i>	588
seqidno157; DNA; <i>Oryza sativa</i>	589	seqidno158; PRT; <i>Oryza sativa</i>	590
seqidno159; DNA; <i>Oryza sativa</i>	591	seqidno160; PRT; <i>Oryza sativa</i>	592
seqidno161; DNA; <i>Oryza sativa</i>	593	seqidno162; PRT; <i>Oryza sativa</i>	594
seqidno163; DNA; <i>Oryza sativa</i>	595	seqidno164; PRT; <i>Oryza sativa</i>	596
seqidno165; DNA; <i>Oryza sativa</i>	597	seqidno166; PRT; <i>Oryza sativa</i>	598
seqidno167; DNA; <i>Oryza sativa</i>	599	seqidno168; PRT; <i>Oryza sativa</i>	600
seqidno169; DNA; <i>Oryza sativa</i>	601	seqidno170; PRT; <i>Oryza sativa</i>	602
seqidno171; DNA; <i>Oryza sativa</i>	603	seqidno172; PRT; <i>Oryza sativa</i>	604
seqidno173; DNA; <i>Oryza sativa</i>	605	seqidno174; PRT; <i>Oryza sativa</i>	606
seqidno175; DNA; <i>Oryza sativa</i>	607	seqidno176; PRT; <i>Oryza sativa</i>	608
seqidno177; DNA; <i>Oryza sativa</i>	609	seqidno178; PRT; <i>Oryza sativa</i>	610
seqidno179; DNA; <i>Oryza sativa</i>	611	seqidno180; PRT; <i>Oryza sativa</i>	612
seqidno181; DNA; <i>Oryza sativa</i>	613	seqidno182; PRT; <i>Oryza sativa</i>	614
seqidno183; DNA; <i>Oryza sativa</i>	615	seqidno184; PRT; <i>Oryza sativa</i>	616
seqidno185; DNA; <i>Oryza sativa</i>	617	seqidno186; PRT; <i>Oryza sativa</i>	618
seqidno187; DNA; <i>Oryza sativa</i>	619	seqidno188; PRT; <i>Oryza sativa</i>	620
seqidno189; DNA; <i>Oryza sativa</i>	621	seqidno190; PRT; <i>Oryza sativa</i>	622
seqidno191; DNA; <i>Oryza sativa</i>	623	seqidno192; PRT; <i>Oryza sativa</i>	624
seqidno193; DNA; <i>Oryza sativa</i>	625	seqidno194; PRT; <i>Oryza sativa</i>	626
seqidno195; DNA; <i>Zea mays</i>	627	seqidno196; PRT; <i>Zea mays</i>	628
seqidno197; DNA; <i>Zea mays</i>	629	seqidno198; PRT; <i>Zea mays</i>	630
seqidno199; DNA; <i>Zea mays</i>	631	seqidno200; PRT; <i>Zea mays</i>	632
seqidno201; DNA; <i>Zea mays</i>	633	seqidno202; PRT; <i>Zea mays</i>	634
seqidno203; DNA; <i>Zea mays</i>	635	seqidno204; PRT; <i>Zea mays</i>	636
seqidno205; DNA; <i>Zea mays</i>	637	seqidno206; PRT; <i>Zea mays</i>	638
seqidno207; DNA; <i>Zea mays</i>	639	seqidno208; PRT; <i>Zea mays</i>	640
seqidno209; DNA; <i>Zea mays</i>	641	seqidno210; PRT; <i>Zea mays</i>	642
seqidno211; DNA; <i>Zea mays</i>	643	seqidno212; PRT; <i>Zea mays</i>	644
seqidno213; DNA; <i>Zea mays</i>	645	seqidno214; PRT; <i>Zea mays</i>	646
seqidno215; DNA; <i>Zea mays</i>	647	seqidno216; PRT; <i>Zea mays</i>	648
seqidno217; DNA; <i>Zea mays</i>	649	seqidno218; PRT; <i>Zea mays</i>	650
seqidno219; DNA; <i>Zea mays</i>	651	seqidno220; PRT; <i>Zea mays</i>	652
seqidno221; DNA; <i>Zea mays</i>	653	seqidno222; PRT; <i>Zea mays</i>	654
seqidno223; DNA; <i>Zea mays</i>	655	seqidno224; PRT; <i>Zea mays</i>	656
seqidno225; DNA; <i>Zea mays</i>	657	seqidno226; PRT; <i>Zea mays</i>	658
seqidno227; DNA; <i>Zea mays</i>	659	seqidno228; PRT; <i>Zea mays</i>	660
seqidno229; DNA; <i>Zea mays</i>	661	seqidno230; PRT; <i>Zea mays</i>	662
seqidno231; DNA; <i>Zea mays</i>	663	seqidno232; PRT; <i>Zea mays</i>	664
seqidno233; DNA; <i>Zea mays</i>	665	seqidno234; PRT; <i>Zea mays</i>	666
seqidno673; DNA; <i>Populus trichocarpa</i>	673	seqidno674; PRT; <i>Populus trichocarpa</i>	674
seqidno675; DNA; <i>Populus trichocarpa</i>	675	seqidno676; PRT; <i>Populus trichocarpa</i>	676
seqidno677; DNA; <i>Populus trichocarpa</i>	677	seqidno678; PRT; <i>Populus trichocarpa</i>	678
seqidno679; DNA; <i>Populus trichocarpa</i>	679	seqidno680; PRT; <i>Populus trichocarpa</i>	680
seqidno681; DNA; <i>Populus trichocarpa</i>	681	seqidno682; PRT; <i>Populus trichocarpa</i>	682
seqidno683; DNA; <i>Populus trichocarpa</i>	683	seqidno684; PRT; <i>Populus trichocarpa</i>	684
seqidno685; DNA; <i>Populus trichocarpa</i>	685	seqidno686; PRT; <i>Populus trichocarpa</i>	686
seqidno687; DNA; <i>Populus trichocarpa</i>	687	seqidno688; PRT; <i>Populus trichocarpa</i>	688
seqidno689; DNA; <i>Populus trichocarpa</i>	689	seqidno690; PRT; <i>Populus trichocarpa</i>	690
seqidno691; DNA; <i>Populus trichocarpa</i>	691	seqidno692; PRT; <i>Populus trichocarpa</i>	692
seqidno693; DNA; <i>Populus trichocarpa</i>	693	seqidno694; PRT; <i>Populus trichocarpa</i>	694
seqidno695; DNA; <i>Populus trichocarpa</i>	695	seqidno696; PRT; <i>Populus trichocarpa</i>	696
seqidno697; DNA; <i>Populus trichocarpa</i>	697	seqidno698; PRT; <i>Populus trichocarpa</i>	698
seqidno699; DNA; <i>Populus trichocarpa</i>	699	seqidno700; PRT; <i>Populus trichocarpa</i>	700
seqidno701; DNA; <i>Populus trichocarpa</i>	701	seqidno702; PRT; <i>Populus trichocarpa</i>	702
seqidno703; DNA; <i>Populus trichocarpa</i>	703	seqidno704; PRT; <i>Populus trichocarpa</i>	704
seqidno705; DNA; <i>Populus trichocarpa</i>	705	seqidno706; PRT; <i>Populus trichocarpa</i>	706
seqidno707; DNA; <i>Populus trichocarpa</i>	707	seqidno708; PRT; <i>Populus trichocarpa</i>	708
seqidno709; DNA; <i>Populus trichocarpa</i>	709	seqidno710; PRT; <i>Populus trichocarpa</i>	710
seqidno711; DNA; <i>Populus trichocarpa</i>	711	seqidno712; PRT; <i>Populus trichocarpa</i>	712
seqidno713; DNA; <i>Populus trichocarpa</i>	713	seqidno714; PRT; <i>Populus trichocarpa</i>	714
seqidno715; DNA; <i>Populus trichocarpa</i>	715	seqidno716; PRT; <i>Populus trichocarpa</i>	716
seqidno717; DNA; <i>Populus trichocarpa</i>	717	seqidno718; PRT; <i>Populus trichocarpa</i>	718
seqidno719; DNA; <i>Populus trichocarpa</i>	719	seqidno720; PRT; <i>Populus trichocarpa</i>	720
seqidno721; DNA; <i>Populus trichocarpa</i>	721	seqidno722; PRT; <i>Populus trichocarpa</i>	722
seqidno723; DNA; <i>Populus trichocarpa</i>	723	seqidno724; PRT; <i>Populus trichocarpa</i>	724
seqidno725; DNA; <i>Populus trichocarpa</i>	725	seqidno726; PRT; <i>Populus trichocarpa</i>	726
seqidno727; DNA; <i>Populus trichocarpa</i>	727	seqidno728; PRT; <i>Populus trichocarpa</i>	728
seqidno729; DNA; <i>Populus trichocarpa</i>	729	seqidno730; PRT; <i>Populus trichocarpa</i>	730
seqidno731; DNA; <i>Populus trichocarpa</i>	731	seqidno732; PRT; <i>Populus trichocarpa</i>	732
seqidno733; DNA; <i>Populus trichocarpa</i>	733	seqidno734; PRT; <i>Populus trichocarpa</i>	734
seqidno735; DNA; <i>Populus trichocarpa</i>	735	seqidno736; PRT; <i>Populus trichocarpa</i>	736

1.5. IAA14 Polypeptides

Table A5 provides a list of nucleic acid sequences related to the nucleic acid sequence used in the methods of the present invention.

TABLE A5

Examples of IAA14-like polypeptides:			
Plant Source	Name	Polypeptide SEQ ID NO:	Nucleic acid SEQ ID NO:
<i>Arabidopsis thaliana</i>	AT4G14550.1#1	738	737
<i>Arabidopsis thaliana</i>	AT3G23050.1#1	748	783
<i>Arabidopsis thaliana</i>	AT3G23050.2#1	749	784
<i>Populus trichocarpa</i>	566151#1	750	785
<i>Populus trichocarpa</i>	720961#1	751	786
<i>Medicago truncatula</i>	TA20354_3880#1	752	787
<i>Solanum lycopersicum</i>	TA40922_4081#1	753	788
<i>Arabidopsis thaliana</i>	AT1G04250.1#1	754	789
<i>Oryza sativa</i>	CB657009#1	755	790
<i>Oryza sativa</i>	TA41733_4530#1	756	791
<i>Medicago truncatula</i>	TA20951_3880#1	757	792
<i>Arabidopsis thaliana</i>	AT3G04730.1#1	758	793
<i>Solanum lycopersicum</i>	TA48108_4081#1	759	794
<i>Medicago truncatula</i>	TA27011_3880#1	760	795
<i>Medicago truncatula</i>	TA22814_3880#1	761	796
<i>Populus trichocarpa</i>	643213#1	762	797
<i>Arabidopsis thaliana</i>	AT3G23030.1#1	763	798
<i>Arabidopsis thaliana</i>	AT4G14560.1#1	764	799
<i>Arabidopsis thaliana</i>	AT1G04240.1#1	765	800
<i>Solanum lycopersicum</i>	TA38817_4081#1	766	801
<i>Solanum lycopersicum</i>	TA43058_4081#1	767	802
<i>Populus trichocarpa</i>	726443#1	768	803
<i>Populus trichocarpa</i>	564913#1	769	804
<i>Populus trichocarpa</i>	831610#1	770	805
<i>Populus trichocarpa</i>	798526#1	771	806
<i>Medicago truncatula</i>	TA20557_3880#1	772	807
<i>Medicago truncatula</i>	TA20558_3880#1	773	808
<i>Populus trichocarpa</i>	823671#1	774	809
<i>Populus trichocarpa</i>	595419#1	775	810
<i>Medicago truncatula</i>	TA31746_3880#1	776	811
<i>Solanum lycopersicum</i>	TA42190_4081#1	777	812
<i>Arabidopsis thaliana</i>	AT4G29080.1#1	778	813
<i>Medicago truncatula</i>	TA25400_3880#1	779	814
<i>Populus trichocarpa</i>	711734#1	780	815
<i>Populus trichocarpa</i>	584053#1	781	816
<i>Medicago truncatula</i>	TA23062_3880#1	782	817

In some instances, related sequences have tentatively been assembled and publicly disclosed by research institutions, such as The Institute for Genomic Research (TIGR). The Eukaryotic Gene Orthologs (EGO) database may be used to identify such related sequences, either by keyword search or by using the BLAST algorithm with the nucleic acid or polypeptide sequence of interest.

Example 2

Alignment of Sequences Related to the Polypeptide
Sequences Used in the Methods of the Invention

2.1. Aspartate AminoTransferase (ASPAT)

Alignment of polypeptide sequences was performed using the ClustalW 2.0 algorithm of progressive alignment (Thompson et al. (1997) Nucleic Acids Res 25:4876-4882; Chema et al. (2003). Nucleic Acids Res 31:3497-3500) with standard setting (slow alignment, similarity matrix: Gonnet (or Blosum 62 (if polypeptides are aligned), gap opening penalty 10, gap extension penalty: 0.2). Minor manual editing was done to further optimise the alignment. The ASPAT polypeptides are aligned in FIG. 1.

A phylogenetic tree of ASPAT polypeptides (FIG. 2) was constructed using a neighbour-joining clustering algorithm as provided in the AlignX programme from the Vector NTI

(Invitrogen). The polypeptides clustered in five major phylogenetic classes, class 1, class 2, class 3, class 4, and class 5. Table B1 shows the polypeptides found within each of the five classes. The polypeptides of Class 5 were used as an outgroup in the phylogenetic analysis and do not represent ASPAT polypeptides. Therefore polypeptides of Class 5 are not part of the invention herein described. Polypeptides within class 1 and 2 are typically expressed in the cytosol or the chloroplast. Class 5 corresponds to the new class of ASPAT polypeptides defined by De La Torre et al. 2006. Polypeptides within class 4 are typically expressed in the mitochondria.

TABLE B1

Phylogenetic classes of ASPAT polypeptides.				
Name	Nucleic acid SEQ ID NO:	Amino acid SEQ ID NO:	Phylo- genetic class	
O. sativa_Os01g0760600	1	2	1	1
O. sativa_Os01g0760600-truncated	3	4	1	1
A. thaliana_AT5G19550	5	6	1	1
A. thaliana_AT5G11520	7	8	1	1
A. thaliana_AT4G31990	9	10	1	1
A. thaliana_AT1G62800	11	12	1	1
B. napus_TA23207	13	14	1	1
B. napus_TA23768	15	16	1	1
C. sinensis_TA12564	17	18	1	1
C. solstitialis_TA659	19	20	1	1
G. hirsutum_TA23799	21	22	1	1
G. max_AF034210	23	24	1	1
G. raimondii_TA9413	25	26	1	1
H. annuus_TA8926	27	28	1	1
H. paradoxus_TA2606	29	30	1	1
J. regia_TA762	31	32	1	1
L. japonicus_TA1537	33	34	1	1
L. perennis_TA512	35	36	1	1
L. perennis_TA605	37	38	1	1
N. tabacum_TA13125	39	40	1	1
P. glauca_TA15326	41	42	1	1
P. patens_136815	43	44	1	1
P. persica_TA3273	45	46	1	1
P. sitchensis_TA22265	47	48	1	1
P. trichocarpa_819551	49	50	1	1
P. trifoliata_TA8305	51	52	1	1
S. lycopersicum_TA38054	53	54	1	1
S. officinarum_TA26595	55	56	1	1
T. aestivum_TA52678	57	58	1	1
V. carteri_82929	59	60	1	1
V. vinifera_GSVIVT00016723001	61	62	1	1
V. vinifera_GSVIVT00032463001	63	64	1	1
Z. mays_TA9042	65	66	1	1
C. reinhardtii_186959	67	68	2	2
C. solstitialis_TA2275	69	70	2	2
C. tinctorius_TA12	71	72	2	2
G. hirsutum_TA24406	73	74	2	2
G. max_TA61768	75	76	2	2
G. raimondii_TA9928	77	78	2	2
H. exilis_TA1663	79	80	2	2
H. vulgare_BPS_7992	81	82	2	2
L. japonicus_TA1466	83	84	2	2
M. polymorpha_TA825	85	86	2	2
N. tabacum_TA13015	87	88	2	2
O. sativa_Os02g0797500	89	90	2	2
P. glauca_TA14780	91	92	2	2
P. patens_102134	93	94	2	2
P. sitchensis_TA20968	95	96	2	2
P. taeda_TA6616	97	98	2	2
P. trichocarpa_654206	99	100	2	2
P. trichocarpa_835828	101	102	2	2
P. vulgaris_TA4043	103	104	2	2
S. tuberosum_TA23192	105	106	2	2
V. carteri_81153	107	108	2	2
V. vinifera_GSVIVT00032723001	109	110	2	2
Z. mays_TA10886	111	112	2	2
A. thaliana_AT2G30970	113	114	4	4
C. sinensis_TA15250	115	116	4	4

TABLE B1-continued

Phylogenetic classes of ASPAT polypeptides.			
Name	Nucleic acid SEQ ID NO:	Amino acid SEQ ID NO:	Phylo- genetic class
G. max_TA50178	117	118	4
G. raimondii_TA9985	119	120	4
H. vulgare_TA32835	121	122	4
H. vulgare_TA36301	123	124	4
O. lucimarinus_31597	125	126	4
O. sativa_Os02g0236000	127	128	4
O. sativa_Os06g0548000	129	130	4
O. taurii_32764	131	132	4
P. patens_169868	133	134	4
P. sitchensis_TA23007	135	136	4
P. taeda_TA7145	137	138	4
V. vinifera_GSVIVT00018772001	139	140	4
V. vinifera_GSVIVT00037462001	141	142	4
A. anophagefferens_21970	143	144	3
A. thaliana_AT2G22250.2	145	146	3
B. napus_BPS_9867	147	148	3
C. reinhardtii_118364	149	150	3
G. hirsutum_TA27281	151	152	3
G. max_BPS_36342	153	154	3
H. vulgare_TA28738	155	156	3
M. domestica_TA26867	157	158	3
N. tabacum_TA15308	159	160	3
O. basilicum_TA1043	161	162	3
O. sativa_Os01g0871300	163	164	3
P. patens_127152	165	166	3
P. pinaster_TA3616_71647	167	168	3
P. trichocarpa_scaff_V.183	169	170	3
P. trichocarpa_scaff_VII.574	171	172	3
S. lycopersicum_TA37592	173	174	3
S. tuberosum_TA27739	175	176	3
T. aestivum_TA71539	177	178	3
V. carteri_103084	179	180	3
V. vinifera_GSVIVT00019453001	181	182	3
Z. mays_BPS_26636	183	184	3
Z. mays_BPS_4233	185	186	3
A. anophagefferens_21841	187	188	5
A. anophagefferens_27031	189	190	5
A. anophagefferens_27395	191	192	5
A. anophagefferens_58638	193	194	5
E. huxleyi_413787	195	196	5
E. huxleyi_437487	197	198	5
E. huxleyi_467854	199	200	5
P. tricornutum_23059	201	202	5
P. tricornutum_23871	203	204	5
T. pseudonana_269248	205	206	5

Alignment of polypeptide sequences was performed using the ClustalW 2.0 algorithm of progressive alignment (Thompson et al. (1997) Nucleic Acids Res 25:4876-4882; Chema et al. (2003). Nucleic Acids Res 31:3497-3500) with standard setting (slow alignment, similarity matrix: Gonnet, gap opening penalty 10, gap extension penalty: 0.2). Minor manual editing was done to further optimise the alignment.

2.2. MYB91 Like Transcription Factor (MYB91)

Multiple sequence alignment of all the MYB91 polypeptide sequences in Table A2 was performed using the ClustalW 1.81 algorithm. Results of the alignment are shown in FIG. 5 of the present application. Two MYB DNA binding domains with an InterPro accession number IPR014778, a MYB transcription factor with an InterPro accession number IPR015495, and a C-terminal Conserved Domain, are marked with X's below the consensus sequence.

2.3. Gibberellic Acid-Stimulated *Arabidopsis* (GASA)

Alignment of polypeptide sequences was performed using the AlignX programme from the Vector NTI (Invitrogen) which is based on the popular Clustal W algorithm of pro-

gressive alignment (Thompson et al. (1997) Nucleic Acids Res 25:4876-4882; Chema et al. (2003). Nucleic Acids Res 31:3497-3500). Default values are for the gap open penalty of 10, for the gap extension penalty of 0.1 and the selected weight matrix is Blosom 62 (if polypeptides are aligned). Minor manual editing was done to further optimise the alignment. Sequence conservation among GASA polypeptides is essentially in the C-terminal part of the polypeptides, the N-terminal part usually being more variable in sequence length and composition. The GASA polypeptides are aligned in FIG. 8.

2.4. Auxin/Indoleacetic Acid Genes (AUX/IAA)

Alignment of polypeptide sequences was performed using the AlignX programme from the Vector NTI (Invitrogen), which is based on the ClustalW 2.0 algorithm for progressive alignment (Thompson et al. (1997) Nucleic Acids Res 25:4876-4882; Chema et al. (2003). Nucleic Acids Res 31:3497-3500); Alignment was performed with standard settings: gap opening penalty 10, gap extension penalty: 0.2. Minor manual editing was done to further optimise the alignment. The AUX/IAA polypeptides are aligned (FIG. 11).

Highly conserved amino acid residues are indicated in the consensus sequence.

2.5. IAA14 Polypeptides

Alignment of polypeptide sequences was performed using the AlignX programme from the Vector NTI (Invitrogen) which is based on the popular Clustal W algorithm of progressive alignment (Thompson et al. (1997) Nucleic Acids Res 25:4876-4882; Chema et al. (2003). Nucleic Acids Res 31:3497-3500). Default values are for the gap open penalty of 10, for the gap extension penalty of 0.1 and the selected weight matrix is Blosom 62 (if polypeptides are aligned). Minor manual editing was done to further optimise the alignment. Sequence conservation among IAA14-like polypeptides is essentially in the C-terminal half of the polypeptides. The IAA14-like polypeptides are aligned in FIG. 14.

Example 3

Calculation of Global Percentage Identity Between Polypeptide Sequences Useful in Performing the Methods of the Invention

3.1. Aspartate AminoTransferase (ASPAT)

Global percentages of similarity and identity between full length polypeptide sequences useful in performing the methods of the invention are determined using one of the methods available in the art, the MatGAT (Matrix Global Alignment Tool) software (BMC Bioinformatics. 2003 4:29. MatGAT: an application that generates similarity/identity matrices using protein or DNA sequences. Campanella J J, Bitincka L, Smalley J; software hosted by Ledion Bitincka). MatGAT software generates similarity/identity matrices for DNA or protein sequences without needing pre-alignment of the data. The program performs a series of pair-wise alignments using the Myers and Miller global alignment algorithm (with a gap opening penalty of 12, and a gap extension penalty of 2), calculates similarity and identity using for example Blosom 62 (for polypeptides), and then places the results in a distance matrix. Sequence similarity is shown in the bottom half of the dividing line and sequence identity is shown in the top half of the diagonal dividing line.

Parameters used in the comparison were:

Scoring matrix: Blosum62

First Gap: 12

Extending gap: 2

A MATGAT table for local alignment of a specific domain, or data on % identity/similarity between specific domains may also be generated.

3.2. MYB91 Like Transcription Factor (MYB91)

Global percentages of similarity and identity between full length polypeptide sequences useful in performing the methods of the invention were determined using one of the methods available in the art, the MatGAT (Matrix Global Alignment Tool) software (BMC Bioinformatics. 2003 4:29. MatGAT: an application that generates similarity/identity matrices using protein or DNA sequences. Campanella J J, Bitincka L, Smalley J; software hosted by Ledion Bitincka). MatGAT software generates similarity/identity matrices for DNA or protein sequences without needing pre-alignment of the data. The program performs a series of pair-wise alignments using the Myers and Miller global alignment algorithm (with a gap opening penalty of 12, and a gap extension penalty of 2), calculates similarity and identity using for example Blosum 62 (for polypeptides), and then places the results in a distance matrix. Sequence similarity is shown in the bottom half of the dividing line and sequence identity is shown in the top half of the diagonal dividing line.

Parameters used in the comparison were:

Scoring matrix: Blosum62

First Gap: 12

Extending gap: 2

Results of the software analysis are shown in Table C1 for the global similarity and identity over the full length of the polypeptide sequences (excluding the partial polypeptide sequences).

The percentage identity between the full length polypeptide sequences useful in performing the methods of the invention can be as low as 52% amino acid identity compared to SEQ ID NO: 221.

The percentage amino acid identity can be significantly increased if the most conserved region of the polypeptides are compared. For example, when comparing the amino acid sequence of a MYB DNA transcription factor with an InterPro entry IPR015495 as represented by SEQ ID NO: 268, or of a MYB DNA binding domain with an InterPro accession number IPR014778 as represented by SEQ ID NO: 269 and/or 270, or of a C-terminal conserved domain as represented by SEQ ID NO: 271 with the respective corresponding domains of the polypeptides of Table A1, the percentage amino acid identity increases significantly (in order of preference at least 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, 99% or more amino acid sequence identity).

3.3. Gibberellic Acid-Stimulated *Arabidopsis* (GASA)

Global percentages of similarity and identity between full length polypeptide sequences useful in performing the methods of the invention were determined using one of the methods available in the art, the MatGAT (Matrix Global Alignment Tool) software (BMC Bioinformatics. 2003 4:29. MatGAT: an application that generates similarity/identity matrices using protein or DNA sequences. Campanella J J, Bitincka L, Smalley J; software hosted by Ledion Bitincka). MatGAT software generates similarity/identity matrices for DNA or protein sequences without needing pre-alignment of the data. The program performs a series of pair-wise alignments using the Myers and Miller global alignment algorithm (with a gap opening penalty of 12, and a gap extension penalty of 2), calculates similarity and identity using for example Blosum 62 (for polypeptides), and then places the results in a distance matrix. Sequence similarity is shown in the bottom half of the dividing line and sequence identity is shown in the top half of the diagonal dividing line.

Parameters used in the comparison were:

Scoring matrix: Blosum62

First Gap: 12

Extending gap: 2

Results of the software analysis are shown in Table C2 for the global similarity and identity over the full length of the polypeptide sequences. Percentage identity is given above the diagonal and percentage similarity is given below the diagonal.

The percentage identity between the GASA polypeptide sequences useful in performing the methods of the invention can be as low as 22.2% amino acid identity compared to SEQ ID NO: 276.

TABLE C1

MatGAT results for global similarity and identity over the full length of the polypeptide sequences of Table A.																							
	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23
1. Antma_MYB91		72	64	64	63	68	69	70	64	70	66	59	71	73	67	48	73	57	68	71	70	73	58
2. Aqufo_MYB91	84		70	68	69	79	74	73	69	76	71	62	76	77	72	50	76	58	73	78	75	83	58
3. Arath_MYB91	77	80		86	91	64	66	66	61	67	63	59	68	67	66	48	68	52	66	71	67	71	54
4. Brana_MYB91	76	80	92		85	63	67	64	62	66	63	59	67	66	65	47	67	52	65	70	66	69	53
5. Carhi_MYB91	77	81	94	91		64	65	64	60	66	62	58	67	66	65	47	67	50	65	69	67	69	52
6. Escca_MYB91	80	86	79	77	78		70	71	68	72	69	60	72	76	71	50	73	57	71	73	72	79	56
7. Eucgr_MYB91	82	87	80	80	79	83		73	68	72	71	64	74	77	71	50	75	54	72	76	74	79	57
8. Glyma_MYB91(a)	80	84	79	78	78	81	84		77	73	76	67	73	76	89	49	74	55	88	76	72	80	57
9. Glyma_MYB91(b)	77	82	76	77	75	78	82	84		69	73	71	67	70	77	51	68	52	76	72	67	74	55
10. Goshi_MYB91	80	87	81	80	79	83	85	83	82		72	62	73	77	73	49	73	55	74	79	72	82	54
11. Lotco_MYB91(a)	77	83	77	77	77	80	84	84	84	83		69	70	73	76	51	72	55	75	73	71	75	56
12. Lotco_MYB91(b)	72	75	72	71	71	72	78	78	80	75	80		62	65	68	46	64	50	68	65	62	66	51
13. Lyces_MYB91	82	87	81	80	81	86	86	83	80	83	81	75		76	72	49	92	56	73	75	98	80	55
14. Maldo_MYB91	84	87	79	81	78	84	88	85	83	86	84	79	86		74	50	77	58	74	79	76	84	58
15. Medtr_MYB91	78	84	79	78	78	82	83	93	85	83	86	79	83	84		49	72	55	96	76	71	78	57
16. Moral_MYB91	63	64	64	65	64	63	65	63	63	62	63	59	63	62	62		49	46	49	52	49	50	44
17. Nicta_MYB91	83	87	80	80	79	84	86	84	81	83	80	76	95	86	82	63		55	73	76	92	81	56
18. Orysa_MYB91	75	75	71	71	69	74	74	70	71	74	73	71	73	75	71	62	72		56	54	55	57	62
19. Pissa_MYB91	79	85	80	79	78	82	83	92	85	83	85	79	83	84	98	63	83	72		76	72	79	57
20. Poptr_MYB91	81	87	83	82	81	84	87	86	82	87	83	75	87	87	85	65	87	73	85		74	86	56
21. Soltu_MYB91	83	86	80	80	80	85	86	82	80	82	83	75	98	86	82	64	95	72	82	86		80	55
22. Vitvi_MYB91	84	91	83	82	82	88	89	88	84	90	85	77	88	90	87	64	89	75	87	93	88		59
23. Zeama_MYB91	73	71	71	70	69	70	72	73	70	70	71	65	71	71	71	64	72	73	71	73	70	71	

TABLE C2

MatGAT results for global similarity and identity over the full length of the polypeptide sequences.													
	1	2	3	4	5	6	7	8	9	10	11	12	13
1. TA5035_4679		42.0	35.5	27.6	35.0	29.9	35.9	52.1	33.0	28.2	35.9	64.3	36.6
2. TA5923_4679	52.1		48.0	34.2	35.6	32.0	33.3	47.1	31.1	28.1	33.3	40.8	36.1
3. Os05g0376800	40.8	55.9		28.8	26.7	28.4	27.6	38.6	23.2	26.6	27.6	34.2	23.9
4. Os04g0465300	37.1	47.1	40.1		24.2	33.1	37.4	28.8	30.6	35.1	37.4	29.5	33.0
5. Os10g0115550	42.7	49.6	42.1	35.0		30.7	33.3	34.6	23.7	29.4	32.5	36.8	30.5
6. AK105729	34.2	42.0	38.8	45.3	49.6		34.2	32.8	42.0	33.1	34.2	29.1	79.5
7. Os05g0432200	44.6	44.5	34.9	47.6	47.0	48.7		33.3	37.9	34.0	98.9	38.0	42.1
8. Os09g0414900	57.3	57.1	48.7	43.6	55.6	41.9	41.0		29.2	31.1	33.3	47.0	35.0
9. Os03g0607200	41.5	42.0	30.9	38.1	37.6	52.1	52.1	40.2		37.7	37.9	31.9	52.6
10. Os07g0592000	38.2	37.8	33.6	47.6	40.2	42.7	43.1	41.0	53.9		34.0	31.1	40.0
11. AK110640	43.5	44.5	34.9	47.6	46.2	48.7	98.9	41.0	52.1	43.1		38.0	42.1
12. Os06g0266800	73.8	46.2	38.2	36.2	44.4	30.8	44.6	53.0	40.4	36.3	43.5		35.4
13. Os03g0760800	43.0	46.2	31.6	46.7	44.4	79.5	59.1	41.0	64.9	51.0	59.1	40.9	
14. scaff_205.30	41.2	43.7	38.2	49.5	39.3	47.9	46.1	47.9	52.9	53.9	46.1	37.3	55.9
15. scaff_II.204	35.6	45.4	37.5	53.3	42.7	49.6	60.4	42.7	48.5	46.1	59.4	37.6	56.4
16. scaff_II.2330	46.3	52.1	45.4	43.0	52.1	43.0	38.0	48.8	33.9	35.5	38.0	43.0	38.0
17. scaff_VI.397	60.0	62.2	49.3	48.6	49.6	45.3	43.0	54.7	43.0	41.2	42.0	59.0	48.0
18. scf_XVII.377	63.6	55.5	48.0	45.8	50.4	40.2	44.9	64.1	41.1	46.7	43.9	55.1	43.9
19. scaff_II.202	38.9	47.1	32.2	56.2	42.7	51.3	64.2	37.6	49.5	44.1	63.2	37.9	61.1
20. scaff_I.2410	44.8	41.2	30.3	40.0	42.7	47.9	53.3	38.5	53.2	48.0	52.2	47.1	57.0
21. scaff_I.1483	54.9	68.1	55.3	45.1	54.7	41.0	43.4	59.8	39.8	36.3	42.5	54.0	45.1
22. scaff_I.1926	18.4	26.1	30.6	26.1	22.4	22.4	22.0	23.3	21.2	20.0	21.6	18.0	19.2
23. scaff_XII.704	43.6	27.7	22.4	41.9	30.8	38.5	47.8	23.9	39.4	33.3	46.7	36.9	48.4
24. scaff_41.75	49.5	41.2	30.9	48.6	44.4	50.4	73.9	40.2	51.1	45.1	72.8	44.0	62.4
25. scaff_40.379	48.9	43.7	32.2	43.8	45.3	53.0	56.5	44.4	64.9	57.8	55.4	45.5	67.7
26. scaff_XV.507	39.8	39.5	28.3	48.6	37.6	41.9	55.9	36.8	42.6	44.1	54.8	38.7	52.7
27. scaff_II.203	43.6	29.4	24.3	36.2	32.5	38.5	54.3	28.2	40.4	34.3	53.3	41.7	47.3
28. scaff_II.2328	58.9	56.3	43.4	45.7	53.8	47.0	55.8	53.0	45.3	44.1	55.8	56.8	54.7
29. scaff_XIX.758	44.8	39.5	30.9	42.9	41.9	41.0	53.3	38.5	47.9	39.2	52.2	43.7	44.1
30. TA45751_4081	47.4	32.8	23.7	32.4	33.3	41.0	44.6	34.2	46.8	44.1	44.6	45.2	51.6
31. TA48119_4081	25.3	37.7	39.5	41.8	39.0	39.7	37.0	37.7	33.6	32.2	36.3	24.7	37.7
32. TA35962_4081	37.5	47.1	36.2	49.5	44.4	47.0	61.5	42.7	48.1	43.3	60.6	38.5	52.9
33. BI208422	65.4	50.4	40.8	40.0	46.2	36.8	43.5	48.7	40.4	43.1	43.5	63.1	46.2
34. BG128975	51.8	64.7	50.0	50.0	58.1	44.4	44.6	62.4	40.2	35.7	43.8	50.9	43.8
35. TA52374_4081	36.6	46.2	35.5	53.6	47.0	47.9	58.0	46.2	46.4	44.6	57.1	39.3	52.7
36. TA37180_4081	57.3	55.5	45.4	45.7	50.4	43.6	49.0	53.0	42.7	49.0	49.0	56.3	50.0
37. BE353147	39.2	44.5	37.5	59.0	36.8	49.6	59.8	40.2	47.1	41.2	58.8	37.3	52.9
38. TA56938_4081	62.5	60.5	46.7	49.5	47.9	41.9	48.1	60.7	42.3	49.0	47.1	55.8	50.0
39. BG130916	70.5	48.7	38.2	40.0	39.3	36.8	40.2	46.2	36.2	37.3	39.1	59.5	45.2
40. SEQ ID NO: 276	51.8	68.1	50.7	48.2	52.1	44.4	45.6	58.1	44.7	42.1	44.7	50.0	45.6
41. TA41886_4081	37.9	45.4	34.9	59.0	35.9	52.1	58.3	38.5	45.6	39.8	58.3	37.9	56.3
42. TA36295_4081	46.6	45.4	35.5	49.5	47.0	41.0	53.4	41.9	47.6	45.6	52.4	41.7	55.3
43. TA56201_4081	50.0	44.5	36.2	47.6	41.9	41.0	43.6	47.0	44.7	45.1	43.6	43.6	51.1
44. AJ785329	52.6	31.9	24.3	26.7	29.1	23.9	30.4	34.2	26.6	28.4	30.4	47.6	30.1
45. CA725087	49.1	54.6	41.4	37.9	49.6	36.8	41.4	56.4	35.3	42.2	41.4	55.2	39.7
46. TA69823_4565	24.4	30.3	29.9	25.4	29.4	33.3	25.4	27.9	28.4	38.8	25.4	20.4	28.4
47. TA53297_4565	43.5	42.9	32.9	50.5	46.2	47.0	87.0	37.6	48.9	36.3	85.9	46.7	58.1
48. TA101332_4565	50.5	55.5	40.1	47.6	63.2	47.9	55.3	56.4	45.6	47.6	54.4	48.5	50.5
49. TA66036_4565	44.7	44.5	34.9	47.6	39.3	73.5	59.6	40.2	63.8	53.9	59.6	42.6	90.4
50. TA100367_4565	55.3	49.6	45.4	44.7	47.9	37.6	40.4	59.0	36.8	42.1	39.5	62.3	38.6
51. TA92393_4565	60.4	55.5	42.1	43.8	51.3	41.0	48.5	58.1	42.6	48.0	48.5	73.3	44.6
52. BM136027	43.6	45.4	34.2	47.6	40.2	72.6	58.5	40.2	62.8	55.9	58.5	42.6	89.4
53. CA705831	33.6	42.0	32.2	38.1	47.9	65.0	44.2	41.0	48.7	43.4	44.2	35.4	69.0
54. CA593033	29.7	38.3	31.6	35.2	41.4	60.2	40.6	36.7	44.5	40.6	40.6	28.1	61.7
55. CK153563	60.6	53.8	40.8	41.9	49.6	38.5	52.1	57.3	44.7	45.1	52.1	68.1	47.9
56. TA66038_4565	40.8	45.4	33.6	42.9	38.5	70.9	58.2	41.0	63.3	52.0	58.2	41.8	85.7
57. TA52915_4565	43.5	41.2	32.2	51.4	45.3	46.2	85.9	38.5	47.9	36.3	84.8	46.7	58.1
58. TA69821_4565	41.1	41.2	38.2	40.2	47.0	46.2	46.7	44.4	49.5	75.7	46.7	34.6	50.5
59. TA95153_4565	30.8	38.7	34.9	41.0	39.3	40.2	47.0	36.8	39.3	36.8	46.2	35.9	41.9
60. CD899399	39.8	44.5	32.9	42.9	38.5	73.5	57.1	40.2	62.2	52.9	57.1	39.8	88.8
61. TA77646_4565	61.6	57.1	43.4	44.8	51.3	38.5	50.5	59.0	41.4	50.0	50.5	70.7	48.5
62. TA51752_4565	29.5	39.5	37.5	34.9	34.9	42.6	44.2	38.8	38.8	35.7	43.4	31.0	38.0
63. Pop_GASA	49.4	43.7	32.2	42.9	43.6	48.7	53.3	42.7	58.5	52.0	52.2	47.2	60.2
64. Mr_GASA	36.6	43.7	36.8	50.9	47.9	45.3	50.0	42.7	50.0	44.6	49.1	36.6	48.2
65. At2g30810	57.5	61.3	45.4	50.9	55.6	45.3	43.4	60.7	39.6	45.3	42.5	51.9	46.2
66. At3g02885	62.9	58.0	46.1	46.7	54.7	44.4	54.6	57.3	47.4	48.0	53.6	59.8	50.5
67. At5g15230	57.5	53.8	40.8	43.4	52.1	42.7	44.3	55.6	42.5	43.4	43.4	54.7	43.4
68. At1g74670	62.4	60.5	45.4	45.7	49.6	40.2	52.5	57.3	41.6	49.0	51.5	58.4	44.6
	14	15	16	17	18	19	20	21	22	23	24	25	26
1. TA5035_4679	31.1	29.7	38.8	51.0	56.1	31.6	34.5	48.7	12.2	36.7	37.6	37.5	32.3
2. TA5923_4679	34.2	36.1	43.8	55.5	46.2	37.0	28.6	54.6	17.6	22.5	29.4	31.1	27.5
3. Os05g0376800	28.3	28.3	36.8	42.8	38.2	26.3	22.4	49.3	22.0	18.3	23.0	24.3	21.6
4. Os04g0465300	34.3	39.0	32.8	32.4	33.6	42.9	28.6	31.6	21.4	36.2	34.0	32.4	39.0
5. Os10g0115550	27.4	28.6	35.6	39.0	38.3	31.9	32.5	34.9	14.0	27.1	32.5	34.2	32.2

TABLE C2-continued

MatGAT results for global similarity and identity over the full length of the polypeptide sequences.													
6. AK105729	39.3	36.0	31.0	32.5	32.5	41.5	40.2	30.6	16.3	30.5	41.0	47.0	33.9
7. Os05g0432200	34.3	49.0	28.9	34.0	34.6	53.1	38.0	32.7	18.0	43.0	59.8	41.3	48.4
8. Os09g0414900	32.5	29.9	38.8	45.3	52.1	29.1	29.1	48.7	16.3	20.3	30.8	33.3	25.4
9. Os03g0607200	37.1	35.6	25.8	27.1	28.2	37.8	38.3	29.3	14.5	31.6	35.1	47.9	33.3
10. Os07g0592000	39.3	33.0	29.3	30.8	33.6	35.0	35.0	27.0	14.6	24.5	31.1	42.7	31.7
11. AK110640	34.3	48.0	28.9	34.0	34.6	52.1	38.0	32.7	17.6	41.9	58.7	41.3	47.3
12. Os06g0266800	32.4	29.8	38.8	51.0	49.5	32.6	35.6	50.4	13.5	32.9	37.4	39.8	34.4
13. Os03g0760800	44.9	43.3	30.6	35.9	34.2	44.9	47.3	33.6	13.7	38.3	51.1	60.6	39.6
14. scaff_205.30		35.9	28.9	33.0	36.1	37.3	49.0	33.6	16.7	26.2	32.4	54.9	31.1
15. scaff_II.204	44.1		29.8	31.7	31.8	77.2	31.7	37.4	19.6	39.2	43.6	34.7	49.0
16. scaff_II.2330	38.0	37.2		39.7	40.5	28.9	24.8	41.3	17.1	23.8	30.6	30.6	25.4
17. scaff_VI.397	46.1	47.5	47.1		49.5	35.0	32.0	61.4	18.4	27.7	36.3	37.0	26.7
18. scf_XVII.377	53.3	48.6	45.5	60.7		33.6	29.9	54.0	13.5	25.0	34.6	38.9	33.3
19. scaff_II.202	45.1	85.1	32.2	47.0	43.0		33.3	33.6	18.0	46.9	47.4	36.1	53.1
20. scaff_I.2410	55.9	43.6	40.5	44.0	45.8	46.3		31.0	14.3	30.7	36.3	62.5	35.7
21. scaff_I.1483	46.0	48.7	52.1	64.6	62.8	44.2	45.1		15.5	24.6	33.6	36.3	30.7
22. scaff_I.1926	22.4	24.9	22.4	23.3	21.2	21.2	20.4	24.5		19.2	15.9	15.5	21.5
23. scaff_XII.704	31.4	44.6	26.4	34.0	29.9	51.6	40.2	30.1	19.6		41.3	32.6	63.8
24. scaff_41.75	47.1	58.4	39.7	48.0	47.7	61.1	50.5	47.8	21.2	48.4		41.8	45.2
25. scaff_40.379	58.8	44.6	39.7	47.0	50.5	47.4	72.7	45.1	19.6	39.8	50.5		36.2
26. scaff_XV.507	37.3	61.4	38.8	40.0	43.0	67.4	49.5	40.7	23.7	67.7	57.0	46.2	
27. scaff_II.203	33.3	55.4	30.6	37.0	30.8	56.8	39.1	34.5	18.0	67.6	57.1	40.9	52.7
28. scaff_II.2328	49.0	49.5	60.3	62.0	57.9	46.3	50.5	61.9	22.0	34.7	53.7	53.7	47.4
29. scaff_XIX.758	37.3	51.5	40.5	46.0	43.9	56.8	43.7	40.7	23.7	52.9	58.2	50.0	55.9
30. TA45751_4081	51.0	32.7	30.6	40.0	37.4	35.8	63.2	31.9	14.7	51.5	42.9	67.0	36.6
31. TA48119_4081	35.6	45.9	37.0	32.2	31.5	44.5	33.6	37.0	32.2	41.8	37.7	31.5	52.7
32. TA35962_4081	40.4	75.0	37.2	46.2	44.9	75.0	41.3	50.4	24.9	47.1	61.5	43.3	58.7
33. BI208422	45.1	42.6	50.4	58.0	54.2	43.2	46.0	56.6	18.8	37.0	48.4	51.1	44.1
34. BG128975	45.5	49.1	52.9	69.6	59.8	42.9	42.9	78.8	24.9	29.5	49.1	48.2	39.3
35. TA52374_4081	49.1	62.5	38.0	44.6	52.7	59.8	39.3	52.2	25.7	42.0	54.5	42.9	52.7
36. TA37180_4081	48.0	46.5	55.4	63.0	61.7	49.0	47.9	62.8	22.4	31.3	49.0	50.0	46.9
37. BE353147	48.0	61.8	34.7	48.0	43.0	63.7	41.2	47.8	24.1	45.1	56.9	46.1	55.9
38. TA56938_4081	54.8	52.9	51.2	62.5	84.1	47.1	47.1	66.4	24.9	32.7	48.1	50.0	43.3
39. BG130916	38.2	38.6	46.3	60.0	47.7	38.9	47.1	54.0	18.4	44.4	45.1	44.3	38.7
40. SEQ ID NO: 276	47.4	45.6	52.1	71.1	60.5	43.9	43.9	71.9	26.9	27.2	42.1	44.7	38.6
41. TA41886_4081	51.5	61.2	38.8	44.7	41.1	64.1	39.8	46.9	26.1	45.6	58.3	43.7	59.2
42. TA36295_4081	42.7	58.3	39.7	44.7	48.6	54.4	45.6	48.7	24.5	42.7	57.3	49.5	55.3
43. TA56201_4081	50.0	45.5	38.8	48.0	51.4	43.2	53.2	49.6	18.8	29.8	46.8	51.1	38.3
44. AJ785329	29.4	26.7	29.8	36.0	35.5	29.5	35.6	34.5	12.2	43.9	31.9	35.2	33.3
45. CA725087	42.2	39.7	49.6	47.4	57.8	38.8	38.8	60.3	19.2	23.3	42.2	38.8	35.3
46. TA69823_4565	27.9	26.9	27.4	25.9	26.9	23.4	25.9	28.9	26.5	15.9	24.4	28.9	24.4
47. TA53297_4565	41.2	60.4	41.3	42.0	47.7	66.3	47.8	47.8	22.0	47.8	72.8	48.9	58.1
48. TA101332_4565	46.6	49.5	45.5	57.3	57.0	55.3	46.6	56.6	24.5	35.0	52.4	52.4	47.6
49. TA66036_4565	55.9	52.5	39.7	40.0	43.9	56.8	59.6	46.0	20.4	45.7	58.5	61.7	51.1
50. TA100367_4565	43.9	47.4	47.1	55.3	60.5	43.9	40.4	62.3	22.4	26.3	43.0	42.1	36.0
51. TA92393_4565	47.1	48.5	51.2	59.4	67.3	46.5	47.5	64.6	19.6	29.7	50.5	45.5	40.6
52. BM136027	54.9	55.4	38.8	49.0	44.9	57.9	59.6	46.0	20.4	45.7	57.4	60.6	51.1
53. CA705831	50.4	46.0	38.8	38.9	41.6	43.4	45.1	44.2	18.0	31.0	44.2	50.4	38.9
54. CA593033	45.3	39.8	33.6	32.8	33.6	38.3	40.6	37.5	17.1	29.7	39.8	45.3	36.7
55. CK153563	51.0	47.5	52.1	57.0	58.9	48.4	53.2	60.2	19.2	31.9	53.2	51.1	42.6
56. TA66038_4565	56.9	49.5	38.8	45.0	45.8	52.0	57.1	46.9	18.8	41.8	56.1	64.3	50.0
57. TA52915_4565	40.2	61.4	42.1	43.0	48.6	64.2	52.2	46.9	22.0	47.8	72.8	47.8	59.1
58. TA69821_4565	53.3	43.0	38.0	42.1	45.8	43.0	46.7	43.4	22.4	29.0	46.7	53.3	39.3
59. TA95153_4565	37.6	41.9	33.1	37.6	37.6	45.3	39.3	41.0	22.0	31.6	43.6	38.5	43.6
60. CD899399	56.9	49.5	38.8	45.0	44.9	52.0	57.1	46.9	20.0	41.8	58.2	63.3	52.0
61. TA77646_4565	50.0	49.5	52.1	58.0	66.4	48.5	48.5	64.6	20.0	30.3	53.5	48.5	44.4
62. TA51752_4565	38.0	39.5	31.0	35.7	38.8	44.2	38.0	39.5	23.3	30.2	40.3	34.9	41.9
63. Pop_GASA	58.8	43.6	41.3	48.0	46.7	46.3	78.7	47.8	21.2	38.2	49.5	78.7	43.0
64. Mt_GASA	42.0	58.0	44.6	43.8	48.2	51.8	44.6	49.6	26.9	48.2	48.2	50.0	67.0
65. At2g30810	48.1	50.9	51.2	59.4	61.7	48.1	44.3	61.1	26.1	31.1	48.1	43.4	41.5
66. At3g02885	48.0	45.5	56.2	62.0	66.4	45.4	51.5	61.9	21.6	35.1	52.6	56.7	50.5
67. At5g15230	46.2	43.4	46.3	53.8	80.4	39.6	45.3	61.1	23.3	31.1	42.5	47.2	42.5
68. At1g74670	50.0	52.5	48.8	65.3	74.8	50.5	44.6	63.7	23.3	35.6	46.5	48.5	47.5
	27	28	29	30	31	32	33	34	35	36	37	38	39
1. TA5035_4679	35.9	48.4	34.5	41.0	21.2	30.8	56.8	44.6	29.5	49.0	32.4	58.7	56.4
2. TA5923_4679	23.5	47.9	31.9	26.9	28.1	35.5	43.7	54.6	31.9	46.3	32.8	47.9	42.0
3. Os05g0376800	19.1	34.9	22.4	21.1	24.1	25.7	33.6	42.1	27.0	36.8	27.6	39.5	34.9
4. Os04g0465300	33.3	38.1	32.4	25.7	33.1	41.0	32.4	36.0	39.8	36.8	45.5	35.8	29.5
5. Os10g0115550	24.8	42.1	29.1	27.7	31.1	40.2	39.1	31.0	39.3	28.2	35.0	32.5	
6. AK105729	32.5	33.3	29.1	38.5	28.2	37.6	28.2	30.8	36.8	30.5	35.9	35.0	29.1
7. Os05g0432200	45.7	42.1	41.3	34.8	30.8	47.6	33.7	35.7	46.4	40.6	44.7	38.5	33.7
8. Os09g0414900	22.2	43.6	28.2	29.1	28.4	32.5	41.0	48.3	33.3	43.6	29.1	51.3	40.2
9. Os03g0607200	33.0	31.6	33.7	36.2	24.8	37.4	30.9	27.0	32.2	27.3	34.3	29.0	26.6
10. Os07g0592000	27.2	35.0	28.8	34.0	21.6	30.2	29.1	28.9	31.6	31.7	28.8	33.6	27.5
11. AK110640	44.6	42.1	40.2	34.8	30.1	46.7	33.7	35.7	45.5	40.6	43.7	38.5	33.7
12. Os06g0266800	34.5	53.7	37.9	39.3	21.9	28.8	56.0	46.4	33.9	50.0	28.4	51.0	56.5

TABLE C2-continued

MatGAT results for global similarity and identity over the full length of the polypeptide sequences.													
13. Os03g0760800	38.7	40.8	33.3	48.4	28.2	40.2	35.5	33.0	40.0	36.4	35.2	35.5	37.6
14. scaff_205.30	30.4	37.3	29.4	47.1	27.4	33.7	32.4	33.9	38.3	34.0	34.9	39.0	31.4
15. scaff_II.204	48.5	42.6	40.6	29.7	34.2	64.4	32.7	36.6	50.9	36.6	52.9	38.1	29.7
16. scaff_II.2330	25.6	53.7	28.9	24.8	25.9	31.4	43.9	43.0	32.2	45.5	27.3	40.5	40.5
17. scaff_VI.397	27.0	55.0	33.0	32.0	22.6	30.8	53.0	59.8	33.0	57.0	34.0	54.8	54.0
18. scf_XVII.377	25.2	47.7	33.6	29.9	24.7	31.8	45.8	51.8	36.8	52.3	31.8	77.6	47.7
19. scaff_II.202	50.5	37.9	41.1	32.6	36.3	62.5	34.7	36.6	50.0	36.5	51.0	39.4	31.6
20. scaff_I.2410	32.2	38.9	35.6	58.6	24.5	33.3	34.5	31.3	28.6	36.5	29.4	31.7	33.3
21. scaff_I.1483	25.7	51.3	29.2	27.4	25.3	36.3	48.7	65.2	37.0	52.6	33.9	54.9	47.8
22. scaff_I.1926	16.3	16.3	18.0	12.2	27.3	19.5	14.3	17.6	20.4	14.9	19.9	15.9	13.9
23. scaff_XII.704	60.9	30.2	45.5	43.9	39.7	43.8	32.9	24.8	35.4	27.8	40.8	27.6	35.6
24. scaff_41.75	44.0	43.2	42.9	35.2	29.5	47.1	37.4	34.5	45.5	36.5	42.2	35.6	38.5
25. scaff_40.379	34.1	38.9	34.1	63.6	24.7	31.7	40.9	35.7	34.2	39.6	32.4	39.0	37.5
26. scaff_XV.507	47.3	36.1	45.2	32.3	47.3	48.6	36.6	31.9	43.4	36.4	43.7	32.4	30.1
27. scaff_II.203		36.8	48.3	41.2	28.8	49.0	38.3	26.8	38.4	32.3	43.1	27.9	38.9
28. scaff_II.2328	42.1		36.5	32.6	26.7	37.5	66.3	50.9	37.7	69.1	35.9	55.8	49.5
29. scaff_XIX.758	55.2	45.3		31.0	29.5	48.1	41.4	31.3	34.8	37.8	38.2	36.5	34.5
30. TA45751_4081	47.1	40.0	40.2		20.5	27.9	37.0	29.5	31.3	32.3	27.5	30.8	40.3
31. TA48119_4081	31.5	33.6	40.4	26.7		33.6	22.6	27.4	37.7	25.0	36.7	26.0	20.5
32. TA35962_4081	55.8	47.1	54.8	29.8	45.9		29.8	33.3	50.0	36.5	50.0	32.7	26.9
33. BI208422	46.9	75.8	52.9	43.2	28.1	41.3		52.7	33.9	84.4	35.3	49.0	61.7
34. BG128975	33.9	65.2	41.1	34.8	39.7	47.3	59.8		33.6	53.1	35.7	58.0	48.2
35. TA52374_4081	50.0	48.2	46.4	34.8	47.3	64.3	40.2	44.6		33.9	40.2	39.8	28.3
36. TA37180_4081	39.6	82.3	50.0	38.5	32.9	47.1	84.4	65.2	44.6		39.0	53.8	51.0
37. BE353147	48.0	53.9	52.0	33.3	43.8	62.5	45.1	48.2	54.5	50.0		37.7	29.4
38. TA56938_4081	34.6	65.4	48.1	37.5	31.5	41.3	56.7	65.2	51.8	61.5	49.0		50.0
39. BG130916	51.4	55.8	47.1	50.0	24.7	35.6	64.2	54.5	34.8	54.2	39.2	51.9	
40. SEQ ID NO: 276	30.7	57.9	38.6	34.2	34.9	43.0	53.5	70.2	43.0	59.6	43.9	65.8	53.5
41. TA41886_4081	49.5	50.5	48.5	34.0	43.2	63.5	42.7	40.2	54.5	50.5	82.5	42.3	38.8
42. TA36295_4081	46.6	49.5	68.0	35.0	41.1	65.4	48.5	53.6	55.4	48.5	57.3	55.8	43.7
43. TA56201_4081	31.9	55.8	38.3	41.5	28.1	38.5	52.1	44.6	42.9	54.2	45.1	51.9	47.9
44. AJ785329	38.2	41.1	34.5	44.6	19.2	27.9	45.7	33.9	26.8	37.5	23.5	36.5	51.4
45. CA725087	30.2	54.3	33.6	29.3	28.8	41.4	50.0	55.2	44.0	54.3	38.8	56.0	41.4
46. TA69823_4565	17.4	27.9	22.9	22.4	34.3	23.9	20.9	27.9	25.9	24.9	23.9	26.9	21.4
47. TA53297_4565	52.2	52.6	57.6	39.1	38.4	60.6	43.5	45.5	55.4	51.0	62.7	46.2	40.2
48. TA101332_4565	36.9	61.2	46.6	39.8	36.3	51.9	54.4	56.3	49.1	60.2	53.4	56.7	48.5
49. TA66036_4565	46.8	52.6	42.6	52.1	37.0	50.0	44.7	46.4	50.0	51.0	53.9	50.0	41.5
50. TA100367_4565	32.5	53.5	36.8	34.2	32.9	36.8	48.2	57.9	43.0	51.8	43.0	55.3	46.5
51. TA92393_4565	36.6	63.4	40.6	36.6	29.5	46.2	58.4	58.0	48.2	62.4	45.1	69.2	51.5
52. BM136027	44.7	52.6	42.6	51.1	37.0	51.0	44.7	46.4	49.1	50.0	52.0	52.9	41.5
53. CA705831	33.6	45.1	32.7	42.5	30.1	41.6	38.1	46.0	46.0	41.6	42.5	49.6	36.3
54. CA593033	29.7	36.7	28.1	38.3	32.2	39.1	28.9	39.1	41.4	32.0	38.3	40.6	30.5
55. CK153563	40.4	67.4	47.9	39.4	28.8	44.2	62.8	58.0	45.5	66.7	44.1	64.4	53.2
56. TA66038_4565	41.8	49.0	42.9	51.0	37.0	51.0	42.9	41.1	50.0	45.9	47.1	49.0	43.9
57. TA52915_4565	52.2	52.6	54.3	39.1	39.0	61.5	43.5	46.4	56.3	50.0	61.8	47.1	40.2
58. TA69821_4565	30.8	45.8	36.4	41.1	34.9	40.2	40.2	43.8	45.5	45.8	44.9	50.5	34.6
59. TA95153_4565	31.6	37.6	38.5	28.2	36.3	47.9	34.2	40.2	47.9	36.8	44.4	39.3	31.6
60. CD899399	42.9	49.0	43.9	51.0	37.0	51.9	42.9	41.1	50.9	46.9	47.1	49.0	42.9
61. TA77646_4565	38.4	66.7	42.4	37.4	30.1	48.1	61.6	59.8	49.1	66.7	47.1	67.3	51.5
62. TA51752_4565	29.5	34.9	35.7	27.1	38.4	45.7	30.2	36.4	51.2	34.1	41.1	39.5	28.7
63. Pop_GASA	37.1	52.6	50.6	61.8	32.2	41.3	51.7	42.9	42.9	50.0	47.1	49.0	48.3
64. Mt_GASA	41.1	45.5	51.8	34.8	56.2	51.8	42.0	50.9	57.1	44.6	48.2	49.1	36.6
65. At2g30810	38.7	60.4	43.4	35.8	37.7	48.1	56.6	65.2	51.8	59.4	54.7	66.0	53.8
66. At3g02885	38.1	79.4	46.4	43.3	36.3	46.2	69.1	66.1	44.6	75.3	46.1	66.3	55.7
67. At5g15230	34.0	59.4	43.4	34.9	33.6	46.2	49.1	57.1	46.4	55.7	42.5	76.4	46.2
68. At1g74670	36.6	65.3	48.5	37.6	32.9	47.1	59.4	64.3	51.8	65.3	44.1	77.9	52.5
	40	41	42	43	44	45	46	47	48	49	50	51	52
1. TA5035_4679	46.5	31.1	36.9	40.4	44.3	41.4	18.8	34.8	42.7	36.2	45.6	50.5	38.3
2. TA5923_4679	57.9	37.0	36.1	33.3	25.8	37.5	23.3	31.7	43.3	32.8	42.9	45.4	33.6
3. Os05g0376800	44.7	25.0	26.3	27.5	19.6	27.2	22.7	27.0	30.9	27.7	33.6	32.9	27.7
4. Os04g0465300	37.4	41.8	39.3	35.2	22.9	27.0	17.2	38.1	35.5	34.5	29.3	33.3	34.5
5. Os10g0115550	35.4	26.9	35.5	30.5	25.4	42.1	20.0	32.5	56.4	29.9	36.2	45.3	29.9
6. AK105729	33.9	36.8	32.5	35.6	22.0	24.8	27.5	37.6	35.6	69.2	33.1	30.6	68.4
7. Os05g0432200	36.8	48.5	43.7	34.0	28.0	28.4	17.8	70.7	39.8	42.1	31.6	37.6	41.1
8. Os09g0414900	47.9	28.1	31.6	38.1	30.5	41.0	21.3	28.2	43.6	30.8	47.0	50.4	31.7
9. Os03g0607200	29.9	31.1	34.9	35.1	23.2	27.1	21.4	35.8	32.1	51.6	28.2	31.5	50.5
10. Os07g0592000	31.0	31.8	30.5	36.5	21.4	28.1	36.6	31.1	36.2	41.9	33.6	33.6	41.9
11. AK110640	36.8	48.5	42.7	34.0	28.0	28.4	17.8	69.6	38.8	42.1	31.6	37.6	41.1
12. Os06g0266800	45.6	32.0	35.0	38.3	41.2	50.9	17.3	37.6	41.9	38.3	59.6	70.3	37.1
13. Os03g0760800	34.2	38.7	37.7	40.8	27.7	30.0	24.0	44.2	40.6	85.1	32.5	36.5	84.0
14. scaff_205.30	35.1	40.2	32.0	35.9	22.3	32.5	21.8	31.4	35.0	46.3	36.0	38.8	45.4
15. scaff_II.204	36.0	53.4	47.1	32.4	22.5	28.3	18.3	51.5	35.9	41.3	32.5	37.1	44.2
16. scaff_II.2330	42.1	31.4	33.1	30.3	27.0	32.6	19.8	29.8	37.2	31.5	38.2	42.1	31.5
17. scaff_VI.397	61.2	32.7	35.9	38.6	32.7	36.8	21.3	31.0	43.7	37.9	45.6	47.5	37.9
18. scf_XVII.377	51.8	32.7	38.3	37.3	31.5	41.1	20.8	35.5	42.6	31.8	49.1	53.3	32.7
19. scaff_II.202	36.0	56.3	44.7	33.3	27.1	29.3	17.3	55.7	40.0	44.9	32.5	39.6	45.9

TABLE C2-continued

MatGAT results for global similarity and identity over the full length of the polypeptide sequences.													
20. scaff_I.2410	29.8	29.1	34.0	38.3	28.4	28.4	19.3	34.8	34.0	48.9	28.1	35.6	48.9
21. scaff_I.1483	60.5	35.1	33.6	37.7	29.8	43.1	21.3	34.5	42.5	37.9	46.5	53.1	37.9
22. scaff_I.1926	18.8	21.1	18.8	14.7	9.4	14.5	17.8	18.8	19.2	16.1	16.3	16.3	16.1
23. scaff_XII.704	21.7	39.4	37.5	25.5	34.8	19.8	12.8	44.1	29.8	36.8	21.7	26.5	36.8
24. scaff_41.75	32.5	45.6	43.3	38.5	29.3	31.4	18.8	56.5	40.8	47.9	34.2	40.8	46.8
25. scaff_40.379	32.5	33.0	37.9	41.5	31.5	31.0	21.3	40.2	42.7	56.4	34.2	39.6	55.3
26. scaff_XV.507	27.0	44.2	46.2	29.8	23.7	27.6	17.7	48.4	35.6	37.8	27.8	32.4	37.8
27. scaff_II.203	25.4	42.7	40.8	28.7	33.3	21.6	14.4	50.0	31.1	38.3	24.6	29.7	37.2
28. scaff_II.2328	47.4	41.7	39.4	41.4	35.4	43.3	20.8	38.9	48.5	40.8	43.5	52.9	40.8
29. scaff_XIX.758	27.2	39.8	58.3	32.6	29.5	26.7	13.4	42.4	38.5	34.0	26.3	32.7	34.0
30. TA45751_4081	27.2	29.1	30.1	35.1	39.4	26.7	18.3	32.6	35.0	50.0	28.9	34.7	48.9
31. TA48119_4081	23.3	36.1	32.2	21.2	15.1	21.0	22.7	31.5	30.6	28.9	24.0	24.7	28.9
32. TA35962_4081	31.6	50.0	50.5	28.6	24.8	28.9	16.7	48.1	38.1	38.3	29.8	35.2	39.3
33. BI208422	48.2	34.0	40.8	42.6	40.2	38.8	14.9	33.7	47.6	35.8	39.5	47.5	35.8
34. BG128975	63.2	30.4	40.0	33.6	28.3	39.5	21.3	35.7	42.9	34.5	46.5	49.1	35.3
35. TA52374_4081	30.7	38.4	42.1	33.6	23.9	34.1	17.3	43.8	38.4	39.1	33.6	42.9	38.3
36. TA37180_4081	50.0	38.1	40.8	39.4	34.0	38.8	16.3	37.5	48.5	37.4	39.7	47.5	36.4
37. BE353147	33.9	72.8	44.3	33.7	21.4	26.7	18.3	47.6	40.0	37.1	33.0	32.4	36.2
38. TA56938_4081	58.8	37.1	44.3	35.5	31.4	43.8	22.3	35.6	41.3	34.6	49.1	59.0	38.0
39. BG130916	47.4	31.1	34.0	36.2	43.8	33.3	16.4	33.7	38.8	35.1	38.6	46.1	35.1
40. SEQ ID NO: 276		35.3	36.8	38.8	27.8	38.2	22.3	34.2	41.7	32.5	44.7	47.4	32.5
41. TA41886_4081	43.9		42.9	34.3	24.0	31.7	18.3	47.6	38.5	37.7	33.9	40.4	37.7
42. TA36295_4081	48.2	55.3		33.7	26.0	31.7	16.3	43.0	40.0	37.4	36.0	37.9	36.4
43. TA56201_4081	48.2	42.7	49.5		45.7	33.3	22.1	34.0	45.7	38.4	34.2	37.3	38.8
44. AJ785329	35.1	25.2	29.1	50.0		27.4	12.4	24.7	33.7	26.3	28.7	35.3	26.3
45. CA725087	53.4	43.1	42.2	44.0	31.0		16.1	29.7	39.2	28.0	54.2	73.7	28.8
46. TA69823_4565	30.3	24.9	25.4	26.4	15.9	22.4		18.8	21.8	21.6	19.3	17.8	21.1
47. TA53297_4565	43.9	61.2	56.3	44.7	27.2	39.7	21.4		40.8	44.2	29.8	39.8	43.2
48. TA101332_4565	56.1	48.5	49.5	55.3	37.9	48.3	28.4	54.4		39.6	38.6	48.5	39.6
49. TA66036_4565	43.9	52.4	52.4	48.9	28.7	33.6	26.4	57.4	47.6		34.2	34.6	98.9
50. TA100367_4565	57.9	45.6	45.6	46.5	35.1	67.2	25.4	41.2	49.1	43.0		68.4	35.0
51. TA92393_4565	60.5	49.5	48.5	49.5	39.6	76.7	23.4	49.5	58.3	39.6	74.6		35.6
52. BM136027	43.9	51.5	51.5	53.2	28.7	34.5	25.9	56.4	47.6	98.9	42.1	40.6	
53. CA705831	43.0	44.2	41.6	36.3	20.4	42.2	27.4	43.4	43.4	69.0	43.9	40.7	68.1
54. CA593033	37.5	39.8	37.5	33.6	21.1	37.5	28.4	38.3	38.3	61.7	39.1	32.8	60.9
55. CK153563	56.1	49.5	46.6	56.4	41.5	70.7	24.4	53.2	56.3	42.6	63.2	85.1	42.6
56. TA66038_4565	46.5	48.5	48.5	49.0	28.6	35.3	28.9	51.0	51.5	82.7	44.7	47.5	81.6
57. TA52915_4565	44.7	60.2	57.3	45.7	27.2	39.7	21.4	98.9	54.4	56.4	41.2	49.5	55.3
58. TA69821_4565	45.6	39.3	44.9	48.6	29.0	38.8	48.3	39.3	58.9	52.3	44.7	43.9	51.4
59. TA95153_4565	44.4	44.4	43.6	37.6	23.1	35.0	25.4	47.9	41.9	41.0	35.9	38.5	40.2
60. CD899399	45.6	48.5	49.5	49.0	27.6	37.1	27.4	52.0	51.5	87.8	43.9	44.6	86.7
61. TA77646_4565	57.0	51.5	50.5	52.5	40.4	80.2	24.4	51.5	59.2	41.4	71.9	94.1	42.4
62. TA51752_4565	41.9	40.3	40.3	38.0	20.2	31.8	28.9	44.2	41.1	38.8	34.9	34.9	36.4
63. Pop_GASA	45.6	44.7	48.5	53.2	36.0	42.2	27.9	43.5	48.5	58.5	43.0	50.5	58.5
64. Mt_GASA	48.2	49.1	54.5	41.1	28.6	43.1	26.4	49.1	47.3	46.4	41.2	44.6	46.4
65. At2g30810	61.4	48.1	49.1	55.7	35.8	50.0	27.9	47.2	66.0	44.3	57.9	60.4	47.2
66. At3g02885	55.3	46.6	48.5	52.6	36.1	54.3	24.9	51.5	66.0	51.5	55.3	65.3	52.6
67. At5g15230	56.1	43.4	52.8	51.9	37.7	55.2	26.9	43.4	57.5	43.4	56.1	64.2	44.3
68. At1g74670	63.2	43.7	53.4	51.5	37.6	56.0	24.4	47.5	57.3	48.5	54.4	64.4	48.5
	53	54	55	56	57	58	59	60	61	62	63	64	65
1. TA5035_4679	30.1	25.8	51.1	36.7	34.8	30.6	23.9	35.7	52.5	21.7	34.8	26.8	48.6
2. TA5923_4679	29.1	26.3	45.4	35.2	30.3	30.6	29.3	34.4	47.1	29.5	32.8	32.5	47.9
3. Os05g0376800	20.7	19.6	32.2	27.1	26.3	28.6	24.0	25.8	33.6	27.9	23.7	27.5	35.5
4. Os04g0465300	27.6	24.6	33.3	31.5	40.0	31.2	32.5	33.3	33.3	27.9	32.4	38.1	36.7
5. Os10g0115550	32.5	27.7	43.6	29.9	32.5	32.2	25.0	29.1	46.2	22.0	33.1	31.3	43.9
6. AK105729	51.5	47.0	30.8	64.4	36.8	36.1	30.8	68.6	30.6	33.6	43.6	31.4	31.6
7. Os05g0432200	30.7	27.9	38.3	44.9	70.7	33.3	37.3	42.9	38.4	34.9	36.6	38.6	34.9
8. Os09g0414900	25.9	23.4	46.2	32.5	29.1	32.8	24.4	31.7	51.3	29.2	30.8	28.9	50.0
9. Os03g0607200	40.4	37.2	31.7	51.5	34.7	36.6	31.1	51.5	32.4	29.0	40.4	33.0	27.5
10. Os07g0592000	32.3	31.2	32.0	40.2	31.1	70.0	27.7	41.1	34.3	26.9	39.8	28.4	32.4
11. AK110640	30.7	27.9	38.3	44.9	69.6	33.3	36.4	42.9	38.4	34.1	36.6	37.7	34.9
12. Os06g0266800	31.0	24.4	64.9	36.7	37.6	29.6	29.1	34.7	67.7	24.0	37.1	28.6	46.2
13. Os03g0760800	63.7	57.0	39.2	78.6	45.3	41.4	32.8	82.7	37.5	29.8	54.8	34.8	34.9
14. scaff_205.30	39.4	36.6	37.9	42.9	30.4	39.8	29.1	44.6	40.8	29.0	55.3	31.9	33.9
15. scaff_II.204	34.1	29.7	37.6	37.5	50.0	32.4	32.5	39.4	37.9	29.5	33.7	43.4	37.0
16. scaff_II.2330	27.3	22.8	41.5	31.5	29.8	30.9	25.2	31.5	41.5	22.5	29.8	34.7	43.0
17. scaff_VI.397	25.4	22.6	47.0	34.0	32.0	32.4	27.4	33.0	48.0	26.4	32.0	31.3	46.2
18. scf_XVII.377	30.2	24.3	49.5	35.5	36.4	35.5	24.8	34.5	53.3	25.6	32.7	31.6	49.5
19. scaff_II.202	34.2	28.8	39.6	38.8	54.7	33.3	33.3	40.8	39.4	32.6	34.7	40.2	36.1
20. scaff_I.2410	38.1	35.2	40.0	50.0	36.6	32.4	29.7	50.0	36.4	26.9	69.2	31.9	31.2
21. scaff_I.1483	28.1	25.3	51.3	35.3	33.6	32.2	28.6	35.3	54.0	30.2	34.5	35.3	49.6
22. scaff_I.1926	11.6	8.6	15.9	14.5	19.2	17.4	16.7	15.3	16.3	17.5	15.9	22.4	17.1
23. scaff_XII.704	24.6	22.5	28.4	33.3	44.1	22.9	25.4	33.3	27.0	23.8	30.0	42.9	25.2
24. scaff_41.75	35.4	30.5	43.2	46.9	56.5	34.5	32.5	48.0	42.6	30.2	36.3	37.5	36.8
25. scaff_40.379	44.2	40.6	43.6	57.6	39.1	40.7	27.4	54.1	40.4	25.6	71.9	36.6	34.0
26. scaff_XV.507	28.2	25.8	34.7	36.4	48.4	27.5	32.2	38.2	36.0	30.0	30.1	56.3	30.8

TABLE C2-continued

MatGAT results for global similarity and identity over the full length of the polypeptide sequences.													
27. scaff_II.203	27.4	22.7	34.0	34.7	50.0	25.0	25.6	35.7	30.3	22.5	31.5	34.8	30.2
28. scaff_II.2328	33.3	27.3	56.7	38.8	38.9	35.2	29.2	38.8	55.9	27.9	38.9	35.7	52.8
29. scaff_XIX.758	25.7	21.1	37.2	32.7	42.4	24.8	27.4	33.7	34.3	24.0	36.0	42.9	33.0
30. TA45751_4081	40.7	37.5	37.2	49.0	32.6	33.3	23.9	49.0	35.4	22.5	57.3	26.8	29.2
31. TA48119_4081	22.0	20.2	23.3	27.5	32.2	26.4	29.9	27.5	25.3	26.0	22.6	43.0	27.4
32. TA35962_4081	29.4	26.2	33.3	35.5	48.1	30.6	37.6	37.4	36.5	34.1	34.6	40.2	36.1
33. BI208422	30.1	23.4	51.1	34.7	33.7	28.7	24.8	34.7	49.5	21.7	36.0	35.7	47.2
34. BG128975	28.9	25.3	50.0	30.4	36.6	33.3	25.6	30.4	50.0	25.6	27.7	33.6	49.6
35. TA52374_4081	32.8	30.2	37.5	38.3	43.8	32.5	34.2	40.9	43.8	33.3	35.7	40.7	39.1
36. TA37180_4081	31.4	24.8	51.0	34.3	37.5	30.6	25.6	33.3	49.5	24.0	35.4	36.6	49.1
37. BE353147	28.2	24.5	34.3	33.3	49.0	32.1	35.9	35.2	34.0	31.0	32.4	36.6	38.9
38. TA56938_4081	33.1	26.8	54.8	35.5	36.5	36.1	28.2	36.4	57.7	26.4	36.8	33.0	52.8
39. BG130916	28.3	24.2	47.9	33.7	33.7	27.8	23.1	32.7	44.0	23.3	36.0	27.7	43.4
40. SEQ ID NO: 276	27.2	24.5	46.5	35.9	34.2	34.5	32.5	35.0	48.2	31.0	32.5	34.8	50.9
41. TA41886_4081	28.8	25.0	41.0	34.0	47.6	33.3	31.6	34.9	39.8	29.5	35.9	36.2	33.0
42. TA36295_4081	30.4	27.1	38.5	36.9	43.0	30.6	31.6	36.9	39.8	27.9	36.5	44.6	38.7
43. TA56201_4081	28.4	26.7	42.1	40.2	35.1	39.4	28.2	40.4	41.0	30.2	39.4	33.0	38.3
44. AJ785329	19.3	19.4	37.9	26.3	24.7	22.2	18.8	25.3	36.0	16.3	28.9	21.4	31.8
45. CA725087	32.2	29.2	68.1	30.5	29.7	25.8	23.3	30.5	78.4	21.4	31.4	27.1	39.0
46. TA69823_4565	19.3	19.6	17.8	21.4	18.3	46.8	18.7	20.6	19.3	22.3	19.8	19.2	21.8
47. TA53297_4565	30.7	27.1	41.7	42.9	97.8	30.8	34.2	44.9	40.6	33.3	31.5	36.8	34.9
48. TA101332_4565	32.8	28.6	47.6	36.8	40.8	40.2	30.8	37.7	50.5	32.6	36.9	33.6	49.5
49. TA66036_4565	65.5	58.6	37.1	79.6	43.2	40.2	31.1	83.7	35.3	26.7	53.2	35.7	34.9
50. TA100367_4565	30.9	27.8	57.9	35.3	30.7	33.6	27.4	34.5	65.8	27.1	32.5	27.8	49.6
51. TA92393_4565	33.3	26.8	84.2	38.7	39.8	33.3	30.8	37.5	94.1	26.4	38.8	33.9	50.9
52. BM136027	64.6	57.8	37.1	78.6	42.1	39.3	30.3	82.7	36.3	28.2	53.2	35.7	37.6
53. CA705831		81.3	35.3	65.8	31.6	31.0	25.4	68.4	33.9	24.0	45.1	26.9	31.3
54. CA593033	82.8		28.2	60.6	27.9	30.1	23.5	61.4	27.2	22.4	41.4	23.5	24.5
55. CK153563	40.7	32.8		41.6	37.2	32.4	28.2	41.6	87.9	26.4	40.4	31.3	50.0
56. TA66038_4565	71.7	65.6	50.0		42.9	39.3	27.7	94.9	38.2	28.2	52.0	32.2	34.9
57. TA52915_4565	43.4	38.3	52.1	51.0		30.8	34.2	44.9	40.6	32.6	31.5	36.3	35.8
58. TA69821_4565	44.2	40.6	42.1	49.5	39.3		28.8	38.2	33.3	27.7	37.6	27.8	34.2
59. TA95153_4565	41.0	38.3	35.9	38.5	48.7	39.3		28.6	29.9	76.7	31.6	31.4	26.5
60. CD899399	72.6	64.8	49.0	96.9	52.0	47.7	36.8		38.2	29.0	53.1	33.0	36.7
61. TA77646_4565	41.6	33.6	88.9	44.4	51.5	45.8	38.5	46.5		27.1	39.6	33.9	51.9
62. TA51752_4565	38.0	38.8	34.1	38.8	45.0	39.5	82.9	37.2	34.9		30.2	30.0	27.1
63. Pop_GASA	50.4	45.3	52.1	61.2	44.6	49.5	39.3	61.2	52.5	39.5		32.1	40.2
64. Mt_GASA	43.4	39.1	41.1	46.4	48.2	42.0	43.6	47.3	46.4	42.6	46.4		33.0
65. At2g30810	41.6	33.6	58.5	45.3	48.1	45.8	41.0	45.3	60.4	40.3	53.8	48.2	
66. At3g02885	46.9	38.3	64.9	51.0	51.5	45.8	41.0	52.0	66.7	37.2	54.6	47.3	61.3
67. At5g15230	38.1	33.6	62.3	48.1	42.5	43.0	39.3	48.1	65.1	38.8	46.2	49.1	55.7
68. At1g74670	42.5	37.5	64.4	48.5	46.5	45.8	40.2	47.5	68.3	39.5	50.5	47.3	64.2
<hr/>													
	66					67				68			
<hr/>													
1. TA5035_4679	48.0					50.0				59.4			
2. TA5923_4679	47.9					45.0				49.6			
3. Os05g0376800	35.5					32.2				39.5			
4. Os04g0465300	37.6					33.6				34.6			
5. Os10g0115550	42.7					37.5				40.7			
6. AK105729	33.9					31.7				34.2			
7. Os05g0432200	39.2					31.1				37.6			
8. Os09g0414900	47.0					41.9				47.0			
9. Os03g0607200	34.0					30.3				27.9			
10. Os07g0592000	38.8					27.9				33.3			
11. AK110640	39.2					31.1				37.6			
12. Os06g0266800	52.6					49.1				48.5			
13. Os03g0760800	41.0					30.3				35.6			
14. scaff_205.30	35.3					33.0				35.9			
15. scaff_II.204	35.6					32.7				38.8			
16. scaff_II.2330	45.5					36.4				40.5			
17. scaff_VI.397	54.0					44.4				54.5			
18. scf_XVII.377	55.5					67.6				63.6			
19. scaff_II.202	40.2					33.0				38.2			
20. scaff_I.2410	35.1					31.1				31.7			
21. scaff_I.1483	51.3					52.2				53.1			
22. scaff_I.1926	15.9					14.7				14.7			
23. scaff_XII.704	27.6					25.2				29.4			
24. scaff_41.75	43.3					33.0				37.6			
25. scaff_40.379	44.3					38.7				35.6			
26. scaff_XV.507	37.0					30.8				38.2			
27. scaff_II.203	32.0					25.5				30.7			
28. scaff_II.2328	64.9					47.2				54.5			
29. scaff_XIX.758	36.1					34.0				35.6			
30. TA45751_4081	33.0					29.2				31.7			

TABLE C2-continued

MatGAT results for global similarity and identity over the full length of the polypeptide sequences.			
31. TA48119_4081	28.8	24.0	27.4
32. TA35962_4081	35.6	34.0	38.5
33. BI208422	57.7	45.3	54.5
34. BG128975	54.0	47.8	53.6
35. TA52374_4081	33.3	31.3	36.8
36. TA37180_4081	61.0	49.5	58.3
37. BE353147	34.6	31.8	34.3
38. TA56938_4081	55.1	63.2	64.4
39. BG130916	48.5	42.5	48.5
40. SEQ ID NO: 276	46.5	43.9	53.5
41. TA41886_4081	35.9	33.3	34.0
42. TA36295_4081	35.9	40.0	41.3
43. TA56201_4081	42.4	37.6	40.2
44. AJ785329	32.7	32.7	33.3
45. CA725087	45.3	42.7	42.4
46. TA69823_4565	21.8	17.8	21.3
47. TA53297_4565	41.2	35.8	32.7
48. TA101332_4565	51.9	41.5	45.6
49. TA66036_4565	41.0	30.3	35.6
50. TA100367_4565	43.6	45.6	42.1
51. TA92393_4565	53.8	54.2	49.5
52. BM136027	42.0	31.2	35.6
53. CA705831	36.1	23.4	31.7
54. CA593033	29.9	21.0	27.5
55. CK153563	55.7	52.3	51.5
56. TA66038_4565	39.2	33.0	36.5
57. TA52915_4565	41.2	34.9	32.7
58. TA69821_4565	37.0	28.7	33.3
59. TA95153_4565	31.1	29.1	30.8
60. CD899399	41.7	33.0	35.6
61. TA77646_4565	58.6	54.2	53.5
62. TA51752_4565	28.7	28.7	26.4
63. Pop_GASA	38.4	33.0	38.2
64. Mt_GASA	33.0	30.1	33.9
65. At2g30810	50.9	45.3	50.9
66. At3g02885		50.0	54.4
67. At5g15230	61.3		57.5
68. At1g74670	65.3	67.9	

3.4. Auxin/Indoleacetic Acid Genes (AUX/IAA)

Global percentages of similarity and identity between full length polypeptide sequences useful in performing the methods of the invention were determined using one of the methods available in the art, the MatGAT (Matrix Global Alignment Tool) software (BMC Bioinformatics. 2003 4:29. MatGAT: an application that generates similarity/identity matrices using protein or DNA sequences. Campanella J J, Bitincka L, Smalley J; software hosted by Ledion Bitincka). MatGAT software generates similarity/identity matrices for DNA or protein sequences without needing pre-alignment of the data. The program performs a series of pair-wise alignments using the Myers and Miller global alignment algorithm (with a gap opening penalty of 12, and a gap extension penalty of 2), calculates similarity and identity using for example Blossum 62 (for polypeptides), and then places the results in a distance matrix.

Parameters that may be used in the comparison:

Scoring matrix: Blossum62

First Gap: 12

Extending gap: 2

3.5. IAA14 Polypeptides

Global percentages of similarity and identity between full length polypeptide sequences useful in performing the methods of the invention were determined using one of the methods available in the art, the MatGAT (Matrix Global Alignment Tool) software (BMC Bioinformatics. 2003 4:29. MatGAT: an application that generates similarity/identity matrices using protein or DNA sequences. Campanella J J, Bitincka L, Smalley J; software hosted by Ledion Bitincka). MatGAT software generates similarity/identity matrices for DNA or protein sequences without needing pre-alignment of the data. The program performs a series of pair-wise alignments using the Myers and Miller global alignment algorithm (with a gap opening penalty of 12, and a gap extension penalty of 2), calculates similarity and identity using for example Blossum 62 (for polypeptides), and then places the results in a distance matrix.

Parameters used in the comparison were:
Scoring matrix: Blossum62
First Gap: 12
Extending gap: 2
Results of the software analysis are shown in Table C3 for the global similarity and identity over the full length of the polypeptide sequences. Percentage identity is given above the diagonal and percentage similarity is given below the diagonal.

Parameters used in the comparison were:

Scoring matrix: Blossum62

First Gap: 12

Extending gap: 2

Results of the software analysis are shown in Table C3 for the global similarity and identity over the full length of the polypeptide sequences. Percentage identity is given above the diagonal and percentage similarity is given below the diagonal.

The percentage identity between the IAA14-like polypeptide sequences useful in performing the methods of the invention can be as low as 26.3% amino acid identity compared to SEQ ID NO: 738 (*A. thaliana*_AT4G14550.1), but is usually above 35%.

TABLE C3

MatGAT results for global similarity and identity over the full length of the polypeptide sequences.												
	1	2	3	4	5	6	7	8	9	10	11	12
1. A. thaliana_AT4G14550.1#1		80.7	68.6	63.6	70.5	73.7	67.2	66.8	26.3	54.5	61.4	64.2
2. A. thaliana_AT3G23050.1#1	84.8		86.4	63.3	68.1	70.2	63.9	63.5	24.7	55.8	60.2	64.3
3. A. thaliana_AT3G23050.2#1	76.3	86.4		53.6	57.5	58.6	53.5	53.6	12.0	45.0	49.2	53.2
4. P. trichocarpa_566151#1	72.2	72.9	61.4		85.6	66.9	63.3	57.3	21.3	56.6	57.8	54.5
5. P. trichocarpa_720961#1	79.0	81.0	68.5	87.4		74.3	70.9	63.9	23.8	54.2	63.6	59.9
6. M. truncatula_TA20354_3880#1	81.4	79.8	69.1	75.8	83.5		72.3	64.4	26.3	55.8	61.2	63.9
7. S. lycopersicum_TA40922_4081#1	76.3	77.4	66.9	70.4	78.2	83.1		60.5	24.6	55.3	59.4	62.5
8. A. thaliana_AT1G04250.1#1	82.5	77.0	67.7	67.5	75.4	77.1	74.6		22.3	50.2	55.5	59.3
9. O. sativa_CB657009#1	27.2	26.3	15.2	23.1	25.4	26.7	25.8	26.6		23.8	24.1	25.4
10. O. sativa_TA41733_4530#1	64.6	66.8	55.2	71.1	67.9	66.8	63.9	59.9	23.8		55.7	54.2
11. M. truncatula_TA20951_3880#1	71.9	73.1	60.5	69.0	76.3	71.5	69.6	68.4	25.7	67.1		63.5
12. A. thaliana_AT3G04730.1#1	75.8	77.8	66.5	67.9	75.8	78.8	77.1	74.6	27.5	64.6	73.5	
13. S. lycopersicum_TA48108_4081#1	69.7	67.9	63.3	62.1	68.5	72.0	69.5	71.2	29.8	56.0	66.8	71.6
14. M. truncatula_TA27011_3880#1	58.5	59.5	51.2	61.5	60.9	59.2	57.5	54.8	18.7	55.2	58.5	58.2
15. M. truncatula_TA22814_3880#1	71.0	72.2	60.0	65.7	72.2	74.3	71.4	71.4	25.7	59.6	70.4	71.4
16. P. trichocarpa_643213#1	75.9	75.7	64.6	68.2	74.2	79.7	78.9	74.3	27.0	66.4	73.9	76.8
17. A. thaliana_AT3G23030.1#1	51.8	48.1	47.6	44.0	48.8	50.8	49.2	52.8	25.3	44.0	47.8	48.7
18. A. thaliana_AT4G14560.1#1	53.5	48.6	48.6	46.2	50.0	53.0	51.3	54.1	29.8	45.5	50.6	52.1
19. A. thaliana_AT1G04240.1#1	54.8	53.5	53.3	46.9	53.6	54.7	53.0	55.0	26.5	47.7	53.8	54.2
20. S. lycopersicum_TA38817_4081#1	54.8	52.7	52.4	46.2	51.2	49.2	51.3	52.0	23.7	44.8	50.2	50.4
21. S. lycopersicum_TA43058_4081#1	55.3	53.1	53.3	48.4	55.6	53.8	51.3	56.8	23.0	47.3	53.4	55.1
22. P. trichocarpa_726443#1	54.4	53.5	53.8	43.3	48.8	52.1	49.2	55.0	24.0	48.0	48.6	55.5
23. P. trichocarpa_564913#1	57.9	52.7	51.4	48.7	51.6	54.7	53.4	59.8	23.2	51.6	51.4	53.8
24. P. trichocarpa_831610#1	57.9	56.0	55.7	49.8	54.8	56.4	55.5	57.2	25.1	50.2	53.0	58.1
25. P. trichocarpa_798526#1	56.6	55.1	54.8	48.4	54.8	57.6	55.9	57.6	23.6	49.1	53.8	57.6
26. M. truncatula_TA20557_3880#1	55.7	53.9	53.8	44.8	50.4	52.5	51.7	55.0	26.4	47.3	50.6	53.0
27. M. truncatula_TA20558_3880#1	55.3	49.8	49.0	46.9	53.6	51.7	53.8	54.1	26.3	48.0	50.6	55.9
28. P. trichocarpa_823671#1	58.3	53.9	54.3	48.0	54.0	56.4	54.7	57.6	23.2	49.8	53.8	55.5
29. P. trichocarpa_595419#1	57.0	55.1	55.7	47.3	53.6	55.9	54.7	55.9	23.4	48.4	52.6	53.8
30. M. truncatula_TA31746_3880#1	56.6	55.1	54.8	49.5	54.0	53.8	53.8	58.5	25.0	48.4	54.2	55.9
31. S. lycopersicum_TA42190_4081#1	54.4	53.9	52.9	49.5	55.2	55.9	53.8	54.1	25.9	50.9	54.2	55.5
32. A. thaliana_AT4G29080.1#1	53.1	54.4	44.9	57.7	55.1	54.4	50.8	52.1	19.3	57.7	58.0	53.8
33. M. truncatula_TA25400_3880#1	46.5	43.2	35.7	37.2	41.5	41.9	43.6	45.0	45.5	40.4	41.5	44.1
34. P. trichocarpa_711734#1	47.0	49.6	41.0	51.0	48.1	48.7	48.1	48.1	17.8	51.3	53.3	48.7
35. P. trichocarpa_584053#1	51.8	56.7	46.6	53.7	55.4	53.4	55.7	53.1	20.2	57.0	56.0	53.4
36. M. truncatula_TA23062_3880#1	46.4	50.1	41.5	50.4	47.6	46.1	47.0	48.4	17.9	51.6	50.7	47.8
	13	14	15	16	17	18	19	20	21	22	23	24
1. A. thaliana_AT4G14550.1#1	58.5	49.3	62.5	63.2	40.6	42.3	41.8	41.8	42.7	43.5	45.6	45.5
2. A. thaliana_AT3G23050.1#1	57.3	48.4	61.2	62.0	39.8	38.9	43.0	40.2	41.6	41.4	41.5	45.3
3. A. thaliana_AT3G23050.2#1	46.1	41.9	50.4	50.2	35.0	35.3	37.9	34.6	36.5	38.3	36.0	39.7
4. P. trichocarpa_566151#1	54.3	44.3	56.4	56.6	36.6	37.5	38.8	38.1	38.8	36.5	41.2	40.8
5. P. trichocarpa_720961#1	58.3	46.9	60.8	61.2	38.8	39.9	43.8	42.0	44.2	40.3	42.7	43.5
6. M. truncatula_TA20354_3880#1	61.3	50.2	64.7	68.0	42.2	42.8	44.5	42.7	43.5	41.8	44.6	45.8
7. S. lycopersicum_TA40922_4081#1	61.6	45.0	60.7	64.3	39.3	41.3	44.6	40.5	42.7	39.8	41.9	45.0
8. A. thaliana_AT1G04250.1#1	58.6	44.3	58.8	59.3	43.3	42.8	45.9	41.5	45.5	41.6	46.1	45.4
9. O. sativa_CB657009#1	26.9	16.4	24.1	24.9	20.9	22.9	21.3	20.2	20.1	19.0	20.0	21.5
10. O. sativa_TA41733_4530#1	50.0	42.0	49.8	57.0	34.9	36.6	37.9	37.5	38.9	39.9	42.0	43.4
11. M. truncatula_TA20951_3880#1	57.6	47.2	61.2	64.7	37.9	39.9	43.5	39.1	43.5	39.9	40.9	45.5
12. A. thaliana_AT3G04730.1#1	60.2	45.9	57.8	62.5	40.3	41.9	42.5	39.2	43.8	41.1	42.2	45.2
13. S. lycopersicum_TA48108_4081#1		45.2	58.7	60.9	43.9	46.2	47.4	44.1	47.2	43.5	44.9	47.2
14. M. truncatula_TA27011_3880#1	52.8		57.5	55.5	30.1	32.0	34.7	31.9	33.7	32.2	33.0	32.3
15. M. truncatula_TA22814_3880#1	66.9	67.6		67.7	39.6	43.5	43.7	42.0	42.5	40.0	41.9	41.9
16. P. trichocarpa_643213#1	70.0	64.5	78.0		40.1	43.5	41.8	40.4	40.6	41.3	44.4	45.6
17. A. thaliana_AT3G23030.1#1	53.4	39.5	50.6	49.4		75.0	57.5	61.7	62.4	60.7	57.5	60.5
18. A. thaliana_AT4G14560.1#1	56.7	41.1	50.6	52.3	85.1		60.2	60.5	59.7	59.8	57.2	59.0
19. A. thaliana_AT1G04240.1#1	61.5	42.8	50.6	53.2	68.3	69.8		62.6	65.5	59.8	57.1	58.1
20. S. lycopersicum_TA38817_4081#1	56.3	43.8	52.2	52.3	71.6	68.9	75.3		77.6	67.2	65.6	63.4
21. S. lycopersicum_TA43058_4081#1	60.6	43.5	51.4	52.3	68.9	67.9	75.5	84.2		66.3	63.3	64.9
22. P. trichocarpa_726443#1	59.1	41.5	50.6	52.3	69.8	66.7	73.4	80.2	77.0		83.7	68.3
23. P. trichocarpa_564913#1	60.1	41.8	51.8	56.5	65.7	63.8	70.0	73.9	73.4	87.0		66.2
24. P. trichocarpa_831610#1	62.0	42.8	51.8	57.4	69.2	68.2	73.3	74.4	76.5	79.5	74.4	
25. P. trichocarpa_798526#1	61.1	43.8	51.0	57.0	67.3	67.3	70.4	73.9	76.4	77.4	73.9	95.0
26. M. truncatula_TA20557_3880#1	57.2	42.1	50.6	54.9	75.8	74.7	75.1	75.3	74.5	80.7	73.4	77.4
27. M. truncatula_TA20558_3880#1	60.1	42.1	50.2	54.0	67.2	68.8	74.1	77.9	75.0	78.6	74.4	80.0
28. P. trichocarpa_823671#1	62.0	44.8	52.2	56.5	63.5	63.1	71.9	75.4	74.9	75.9	73.9	80.3
29. P. trichocarpa_595419#1	63.0	45.2	53.9	53.2	67.7	64.2	73.1	77.6	74.6	76.6	72.0	81.1
30. M. truncatula_TA31746_3880#1	61.1	42.1	52.7	56.1	63.2	65.7	71.1	70.6	72.1	72.5	71.5	82.8
31. S. lycopersicum_TA42190_4081#1	58.7	44.1	51.4	55.3	68.6	71.4	75.7	72.6	74.0	75.0	67.6	76.4
32. A. thaliana_AT4G29080.1#1	49.8	51.1	54.4	55.1	42.0	41.3	47.9	44.3	44.9	45.6	46.9	48.5
33. M. truncatula_TA25400_3880#1	49.5	33.4	42.4	45.1	44.8	50.0	42.3	41.6	39.8	39.6	40.6	41.5
34. P. trichocarpa_711734#1	45.6	49.9	48.7	49.3	35.5	37.5	38.7	37.2	38.7	39.0	40.4	40.4
35. P. trichocarpa_584053#1	50.2	51.8	53.7	53.7	39.4	41.0	42.7	42.0	43.3	44.0	44.6	44.0
36. M. truncatula_TA23062_3880#1	43.8	47.0	47.3	48.7	35.7	36.3	39.8	38.9	37.8	39.8	38.9	41.5

TABLE C3-continued

MatGAT results for global similarity and identity over the full length of the polypeptide sequences.												
	25	26	27	28	29	30	31	32	33	34	35	36
1. A. thaliana_AT4G14550.1#1	43.1	42.9	44.0	43.2	41.7	44.4	42.4	43.0	36.1	38.4	43.3	36.5
2. A. thaliana_AT3G23050.1#1	41.8	41.9	40.7	42.6	42.4	43.8	42.3	43.1	33.9	39.1	43.2	37.3
3. A. thaliana_AT3G23050.2#1	36.6	37.4	35.0	37.4	37.3	38.2	36.6	35.7	25.0	31.7	35.1	31.5
4. P. trichocarpa_566151#1	38.6	37.9	36.5	39.7	37.9	40.4	38.3	41.8	31.5	40.0	41.7	37.0
5. P. trichocarpa_720961#1	43.5	42.3	41.1	44.8	42.7	43.5	42.3	43.3	34.8	39.5	43.6	36.6
6. M. truncatula_TA20354_3880#1	43.9	42.3	40.8	44.6	42.6	43.3	43.9	43.6	35.3	39.3	41.9	36.7
7. S. lycopersicum_TA40922_4081#1	42.3	39.0	41.9	42.1	42.6	44.1	41.8	43.6	37.4	41.5	45.9	37.6
8. A. thaliana_AT1G04250.1#1	44.6	40.8	43.2	45.9	42.4	44.0	42.4	42.6	35.9	37.4	41.2	36.4
9. O. sativa_CB657009#1	19.6	22.6	22.2	19.4	20.6	22.1	23.2	17.0	37.9	15.5	17.3	14.7
10. O. sativa_TA41733_4530#1	40.3	38.2	39.0	42.3	40.1	40.1	39.5	44.6	33.2	41.3	42.2	40.5
11. M. truncatula_TA20951_3880#1	43.9	40.7	39.5	44.1	43.3	42.9	45.5	48.0	34.9	42.4	45.0	40.9
12. A. thaliana_AT3G04730.1#1	42.9	42.4	42.8	41.9	43.0	41.2	44.9	42.0	35.7	37.5	41.4	36.7
13. S. lycopersicum_TA48108_4081#1	46.8	44.1	46.7	46.6	46.1	45.3	48.1	39.9	41.1	38.3	41.7	35.1
14. M. truncatula_TA27011_3880#1	33.0	33.7	33.3	33.9	33.8	35.3	35.0	33.7	26.6	31.1	33.1	30.4
15. M. truncatula_TA22814_3880#1	42.1	41.5	40.0	42.4	41.5	41.5	42.4	44.1	36.4	39.1	43.1	37.5
16. P. trichocarpa_643213#1	44.4	44.0	45.0	43.0	42.0	44.3	43.0	43.6	38.1	40.7	43.7	38.0
17. A. thaliana_AT3G23030.1#1	57.4	58.0	56.9	54.6	54.2	54.6	55.6	34.1	36.0	28.9	30.9	27.3
18. A. thaliana_AT4G14560.1#1	56.8	58.1	58.3	57.6	57.2	55.9	58.1	33.4	36.6	30.9	33.6	29.1
19. A. thaliana_AT1G04240.1#1	58.4	59.0	59.7	60.5	60.6	56.5	58.5	37.7	31.6	29.8	33.9	30.3
20. S. lycopersicum_TA38817_4081#1	61.9	62.6	64.2	61.8	62.4	61.2	59.3	35.4	31.9	29.5	33.2	30.3
21. S. lycopersicum_TA43058_4081#1	62.4	61.7	61.8	60.9	59.5	62.6	61.0	37.0	32.4	30.7	34.7	30.3
22. P. trichocarpa_726443#1	66.3	69.4	64.9	65.4	62.6	61.8	60.1	38.4	30.5	32.3	36.2	30.3
23. P. trichocarpa_564913#1	63.5	62.3	63.0	62.9	59.7	61.0	55.8	39.0	32.2	33.7	37.5	30.8
24. P. trichocarpa_831610#1	92.0	62.8	66.8	69.1	67.3	70.0	65.2	38.7	31.8	34.1	37.0	33.4
25. P. trichocarpa_798526#1		62.3	64.5	66.8	65.0	69.1	62.2	37.4	31.7	33.2	36.2	33.1
26. M. truncatula_TA20557_3880#1	74.9		69.4	60.9	61.0	58.8	59.0	36.7	33.2	27.8	32.9	29.7
27. M. truncatula_TA20558_3880#1	77.4	81.7		65.0	63.7	61.5	56.3	33.8	33.5	30.1	35.5	31.7
28. P. trichocarpa_823671#1	80.8	72.9	75.4		89.2	63.8	57.8	38.0	31.9	33.2	36.2	32.6
29. P. trichocarpa_595419#1	82.1	74.1	75.6	94.6		62.7	57.8	39.7	31.7	31.5	35.5	32.3
30. M. truncatula_TA31746_3880#1	82.8	73.0	73.5	76.5	77.5		60.1	38.8	31.6	33.5	38.4	34.8
31. S. lycopersicum_TA42190_4081#1	73.9	76.2	75.3	73.9	73.6	71.6		37.7	32.1	30.9	38.1	29.1
32. A. thaliana_AT4G29080.1#1	48.2	43.3	44.9	47.2	47.9	46.9	45.9		32.5	54.6	57.7	45.3
33. M. truncatula_TA25400_3880#1	42.2	44.9	44.1	41.9	40.8	40.2	44.3	36.7		30.7	36.2	28.0
34. P. trichocarpa_711734#1	39.8	36.4	36.4	39.5	39.3	40.1	39.3	66.8	35.2		61.4	49.1
35. P. trichocarpa_584053#1	44.6	43.0	42.0	46.3	45.0	46.9	46.3	69.4	39.1	69.1		47.3
36. M. truncatula_TA23062_3880#1	41.8	38.9	39.8	40.6	40.6	43.2	38.6	58.5	32.0	65.6	59.7	

Example 4

Identification of Domains Comprised in Polypeptide Sequences Useful in Performing the Methods of the Invention

4.1. Aspartate AminoTransferase (ASPAT)

The Integrated Resource of Protein Families, Domains and Sites (InterPro) database is an integrated interface for the commonly used signature databases for text- and sequence-based searches. The InterPro database combines these databases, which use different methodologies and varying degrees of biological information about well-characterized proteins to derive protein signatures. Collaborating databases

include SWISS-PROT, PROSITE, TrEMBL, PRINTS, ProPom and Pfam, Smart and TIGRFAMs. Pfam is a large collection of multiple sequence alignments and hidden Markov models covering many common protein domains and families. Pfam is hosted at the Sanger Institute server in the United Kingdom. Interpro is hosted at the European Bioinformatics Institute in the United Kingdom.

The results of the InterPro scan of the polypeptide sequence as represented by SEQ ID NO: 4, by SEQ ID NO: 2 and by SEQ ID NO: 6 are presented in Table D1, Table D2 and Table D3, respectively.

Tables D1, D2, D3: InterPro scan results (major accession numbers) of the polypeptide sequence as represented by SEQ ID NO: 4, SEQ ID NO: 2 and SEQ ID NO: 6 respectively.

TABLE D1

Database	Accession number	Accession name	Amino Acid Coordinates in on SEQ ID NO: 2, (Start-End)	e-value
InterPro	IPR000796	Aspartate/other aminotransferase		
HMMPanther	PTHR11879	ASPARTATE AMINOTRANSFERASE	[1-204]	2.6e-123
InterPro	IPR004839	Aminotransferase, class I and II	class	
HMMFam	PF00155	Aminotran_1_2	[31-203]	8.3e-61
InterPro	IPR015421	Pyridoxal phosphate-dependent transferase, major region, subdomain	[50-203]	
Gene3D	G3DSA:3.40.640.10	no description	description	7.8e-57
InterPro	IPR015424	Pyridoxal phosphate-dependent transferase,	phosphate-dependent	
superfamily	SSF53383	PLP-dependent transferases	[2-203]	6.2e-56

TABLE D2

Database	Accession number	Aspartate/other aminotransferase	Amino Acid Coordinates in on SEQ ID NO: 6, (Start-End)	e-value
InterPro FPrintScan	IPR000796 PR00799	Aspartate/other aminotransferase TRANSAMINASE	[234-253]; [265-279]; [301-321]; [401-419]; 427-445] [38-460]	5.9E-68 0.0
HMMPanther InterPro	PTHR11879 IPR004838	Asp_trans Aminotransferases, class-I, pyridoxal-phosphate-binding site AA_TRANSFER_CLASS_1	[303-316]	8.0E-5
ProfileScan InterPro HMMPfam InterPro	PS00105 IPR004839 PF00155 IPR015421	Aminotransferase, class I and II Aminotran_1_2 Pyridoxal phosphate-dependent transferase major region, subdomain I	[84-452]	0.0
Gene3D InterPro	G3DSA:3.40.640.10 IPR015424	PyrdxIP-dep_Trfase_major_sub1 Pyridoxal phosphate-dependent transferase major region	[103-375]	3.8E-111
superfamily	SSF53383	PyrdxIP-dep_Trfase_major	[55-460]	6.8E-121

TABLE D3

Database	Accession number	Aspartate/other aminotransferase	Amino Acid Coordinates [Start-End] - Evalule
InterPro FPrintScan	IPR000796 PR00799	Aspartate/other aminotransferase TRANSAMINASE	aminotransferase [179-198]; [210-224]; [246-266]; [278-303]; [346-364]; [372-390]; - 1.6e-70
HMMPanther InterPro	PTHR11879 IPR004838	ASPARTATE AMINOTRANSFERASE Aminotransferases, Class I pyridoxal- phosphate-binding site AA_TRANSFER_CLASS_1	[1-405] - 6.2e-259
ScanRegExp InterPro HMMPfam InterPro	PS00105 IPR004839 PF00155 IPR015421	Aminotransferase, class I and II Aminotran_1_2 Pyridoxal phosphate-dependent transferase, major region subdomain I	[248-261] - 0.00008 [29-397] - 1.4e-140
Gene3D InterPro	G3DSA:3.40.640.10 IPR015424	no description Pyridoxal phosphate transferase major region	[48-320] - 1.7e-107
superfamily	SSF53383	PLP-dependent transferase	[1-405] - 1.3e-119

4.2. MYB91 Like Transcription Factor (MYB91)

The Integrated Resource of Protein Families, Domains and Sites (InterPro) database is an integrated interface for the commonly used signature databases for text- and sequence-based searches. The InterPro database combines these databases, which use different methodologies and varying degrees of biological information about well-characterized proteins to derive protein signatures. Collaborating databases

include SWISS-PROT, PROSITE, TrEMBL, PRINTS, ProPom and Pfam, Smart and TIGRFAMs. Interpro is hosted at the European Bioinformatics Institute in the United Kingdom.

The results of the InterPro scan of the polypeptide sequence as represented by SEQ ID NO: 221 are presented in Table D4.

TABLE D4

InterPro scan results of the polypeptide sequence as represented by SEQ ID NO: 221			
InterPro accession number and name	Integrated database Name	Integrated database accession number	Integrated database accession name
IPR001005 SANT, DNA-binding domain	SMART	SM00717	SANT
IPR009057 homeodomain-like	SUPERFAMILY	SSF46689	Homeodomain-like
IPR012287 Homeodomain-related	GENE3D	G3DSA:1.10.10.60	
IPR014778 Myb, DNA-binding	PFAM	PF00249	Myb_DNA-binding
IPR015495 Myb transcription factor	PANTHER	PTHR10641	MYB-related
No IPR unintegrated	PANTHER	PTHR10641:SF24	Assymetric leaves1 and Rough Sheath2
No IPR unintegrated	PROFILE	PS51294	HTH_MYB

4.3. Gibberellic Acid-Stimulated *Arabidopsis* (GASA)

The Integrated Resource of Protein Families, Domains and Sites (InterPro) database is an integrated interface for the commonly used signature databases for text- and sequence-based searches. The InterPro database combines these databases, which use different methodologies and varying degrees of biological information about well-characterized proteins to derive protein signatures. Collaborating databases include SWISS-PROT, PROSITE, TrEMBL, PRINTS, Pro-

pom and Pfam, Smart and TIGRFAMs. Pfam is a large collection of multiple sequence alignments and hidden Markov models covering many common protein domains and families. Pfam is hosted at the Sanger Institute server in the United Kingdom. Interpro is hosted at the European Bioinformatics Institute in the United Kingdom.

The results of the InterPro scan of the polypeptide sequence as represented by SEQ ID NO: 2 are presented in Table D5.

TABLE D5

InterPro scan results (major accession numbers) of the polypeptide sequence as represented by SEQ ID NO: 276.			
Database	Accession number	Accession name	Amino acid coordinates on SEQ ID NO 2
InterPro	IPR003854	Gibberellin regulated protein GASA	5-114
HMMPfam	PF02704		

4.4. Auxin/Indoleacetic Acid Genes (AUX/IAA)

The presence of conserved protein domains in SEQ ID NO: 432 was determined by searching the pfam database. Pfam is a large collection of multiple sequence alignments and hidden Markov models covering many common protein domains and families. Pfam is hosted at the Sanger Institute server in the United Kingdom.

The results of the search of the Pfam with the query sequence as represented by SEQ ID NO: 432 are presented in Table D6.

TABLE D6

Pfam search results (major accession numbers) of the polypeptide sequence as represented by SEQ ID NO: 432.								
		Entry	Amino acid coordinate of domain PF02309 in SEQ ID NO: 2		HMM	Bits	Alignment	
Pfam-A	Description	type	Start	End	From	To	score	E-value mode
AUX_IAA	AUX/IAA family	Family PF02309	5	171	1	269	70.3	6.9e-18 Is

The Alignment mode use is the so called “Is”. Parameters used in the model are given in Table D7.

TABLE D7

HMM model Is model: hmmbuild -F HMM_Is SEED hmmcalibrate --cpu 1 --seed 0 HMM_Is			
Is			
Parameter	Sequence	Domain	
Gathering cut-off	-83	-83	
Trusted cut-off	-82	-82	
Noise cut-off	-83.5	-83.5	

4.5. IAA14 Polypeptides

The Integrated Resource of Protein Families, Domains and Sites (InterPro) database is an integrated interface for the commonly used signature databases for text- and sequence-based searches. The InterPro database combines these databases, which use different methodologies and varying degrees of biological information about well-characterized proteins to derive protein signatures. Collaborating databases include SWISS-PROT, PROSITE, TrEMBL, PRINTS, Pro-

pom and Pfam, Smart and TIGRFAMs. Pfam is a large collection of multiple sequence alignments and hidden Markov models covering many common protein domains and families. Pfam is hosted at the Sanger Institute server in the United Kingdom. Interpro is hosted at the European Bioinformatics Institute in the United Kingdom.

The results of the InterPro scan of the polypeptide sequence as represented by SEQ ID NO: 738 are presented in Table D8.

TABLE D8

InterPro scan results (major accession numbers) of the polypeptide sequence as represented by SEQ ID NO: 738.			
Database	Accession number	Accession name	Amino acid coordinates on SEQ ID NO 738
InterPro	IPR003311	AUX/IAA protein	
HMMPfam	PF02309	AUX_IAA	1-220
InterPro	IPR011525	Aux/IAA-ARF-dimerisation	
ProfileScan	PS50962	IAA_ARF	111-211
InterPro	NULL	NULL	
superfamily	SSF54277	CAD & PB1 domains	106-209

Example 5

Topology Prediction of the Polypeptide Sequences Useful in Performing the Methods of the Invention

5.1. Aspartate AminoTransferase (ASPAT)

TargetP 1.1 predicts the subcellular location of eukaryotic proteins. The location assignment is based on the predicted presence of any of the N-terminal pre-sequences: chloroplast transit peptide (cTP), mitochondrial targeting peptide (mTP) or secretory pathway signal peptide (SP). Scores on which the final prediction is based are not really probabilities, and they do not necessarily add to one. However, the location with the highest score is the most likely according to TargetP, and the relationship between the scores (the reliability class) may be

an indication of how certain the prediction is. The reliability class (RC) ranges from 1 to 5, where 1 indicates the strongest prediction. TargetP is maintained at the server of the Technical University of Denmark.

For the sequences predicted to contain an N-terminal pre-sequence a potential cleavage site can also be predicted.

A number of parameters were selected, such as organism group (non-plant or plant), cutoff sets (none, predefined set of cutoffs, or user-specified set of cutoffs), and the calculation of prediction of cleavage sites (yes or no).

The protein sequences representing the GRP are used to query TargetP 1.1. The "plant" organism group is selected, no cutoffs defined, and the predicted length of the transit peptide requested.

Many other algorithms can be used to perform such analyses, including:

ChloroP 1.1 hosted on the server of the Technical University of Denmark;

Protein Prowler Subcellular Localisation Predictor version 1.2 hosted on the server of the Institute for Molecular Bioscience, University of Queensland, Brisbane, Australia;

PENCE Proteome Analyst PA-GOSUB 2.5 hosted on the server of the University of Alberta, Edmonton, Alberta, Canada;

TMHMM, hosted on the server of the Technical University of Denmark

5.2. Gibberellic Acid-Stimulated *Arabidopsis* (GASA)

TargetP 1.1 predicts the subcellular location of eukaryotic proteins. The location assignment is based on the predicted presence of any of the N-terminal pre-sequences: chloroplast transit peptide (cTP), mitochondrial targeting peptide (mTP) or secretory pathway signal peptide (SP). Scores on which the final prediction is based are not really probabilities, and they do not necessarily add to one. However, the location with the highest score is the most likely according to TargetP, and the relationship between the scores (the reliability class) may be an indication of how certain the prediction is. The reliability class (RC) ranges from 1 to 5, where 1 indicates the strongest prediction. TargetP is maintained at the server of the Technical University of Denmark.

For the sequences predicted to contain an N-terminal pre-sequence a potential cleavage site can also be predicted.

A number of parameters were selected, such as organism group (non-plant or plant), cutoff sets (none, predefined set of cutoffs, or user-specified set of cutoffs), and the calculation of prediction of cleavage sites (yes or no).

The results of TargetP 1.1 analysis of the polypeptide sequence as represented by SEQ ID NO: 221 are presented Table E1. The "plant" organism group has been selected, no cutoffs defined, and the predicted length of the transit peptide requested. The polypeptide sequence as represented by SEQ ID NO: 221 is predicted to be secreted, with a secretion signal sequence of 24 amino acids.

TABLE E1

TargetP 1.1 analysis of the polypeptide sequence as represented by SEQ ID NO: 221	
Length (AA)	114
Chloroplastic transit peptide	0.022
Mitochondrial transit peptide	0.022
Secretory pathway signal peptide	0.960
Other subcellular targeting	0.023
Predicted Location	S
Reliability class	1
Predicted transit peptide length	24

Many other algorithms can be used to perform such analyses, including:

ChloroP 1.1 hosted on the server of the Technical University of Denmark;

Protein Prowler Subcellular Localisation Predictor version 1.2 hosted on the server of the Institute for Molecular Bioscience, University of Queensland, Brisbane, Australia;

PENCE Proteome Analyst PA-GOSUB 2.5 hosted on the server of the University of Alberta, Edmonton, Alberta, Canada;

TMHMM, hosted on the server of the Technical University of Denmark

PSORT (URL: psort.org)

PLOC (Park and Kanehisa, Bioinformatics, 19, 1656-1663, 2003).

5.3. Auxin/Indoleacetic Acid Genes (AUX/IAA)

TargetP 1.1 predicts the subcellular location of eukaryotic proteins. The location assignment is based on the predicted presence of any of the N-terminal pre-sequences: chloroplast transit peptide (cTP), mitochondrial targeting peptide (mTP) or secretory pathway signal peptide (SP). Scores on which the final prediction is based are not really probabilities, and they do not necessarily add to one. However, the location with the highest score is the most likely according to TargetP, and the relationship between the scores (the reliability class) may be an indication of how certain the prediction is. The reliability class (RC) ranges from 1 to 5, where 1 indicates the strongest prediction. TargetP is maintained at the server of the Technical University of Denmark.

For the sequences predicted to contain an N-terminal pre-sequence a potential cleavage site can also be predicted.

A number of parameters were selected, such as organism group (non-plant or plant), cutoff sets (none, predefined set of cutoffs, or user-specified set of cutoffs), and the calculation of prediction of cleavage sites (yes or no).

Many other algorithms can be used to perform such analyses, including:

ChloroP 1.1 hosted on the server of the Technical University of Denmark;

Protein Prowler Subcellular Localisation Predictor version 1.2 hosted on the server of the Institute for Molecular Bioscience, University of Queensland, Brisbane, Australia;

PENCE Proteome Analyst PA-GOSUB 2.5 hosted on the server of the University of Alberta, Edmonton, Alberta, Canada;

TMHMM, hosted on the server of the Technical University of Denmark

PSORT (URL: psort.org)

PLOC (Park and Kanehisa, Bioinformatics, 19, 1656-1663, 2003).

5.4. IAA14 polypeptides

TargetP 1.1 predicts the subcellular location of eukaryotic proteins. The location assignment is based on the predicted presence of any of the N-terminal pre-sequences: chloroplast transit peptide (cTP), mitochondrial targeting peptide (mTP) or secretory pathway signal peptide (SP). Scores on which the final prediction is based are not really probabilities, and they do not necessarily add to one. However, the location with the highest score is the most likely according to TargetP, and the relationship between the scores (the reliability class) may be an indication of how certain the prediction is. The reliability class (RC) ranges from 1 to 5, where 1 indicates the strongest prediction. TargetP is maintained at the server of the Technical University of Denmark.

For the sequences predicted to contain an N-terminal pre-sequence a potential cleavage site can also be predicted.

A number of parameters were selected, such as organism group (non-plant or plant), cutoff sets (none, predefined set of

cutoffs, or user-specified set of cutoffs), and the calculation of prediction of cleavage sites (yes or no).
The results of TargetP 1.1 analysis of the polypeptide sequence as represented by SEQ ID NO: 738 are presented Table E2. The “plant” organism group has been selected, no cutoffs defined, and the predicted length of the transit peptide requested. The subcellular localization of the polypeptide sequence as represented by SEQ ID NO: 738 may be the cytoplasm or nucleus, no transit peptide is predicted.

TABLE E2

TargetP 1.1 analysis of the polypeptide sequence as represented by SEQ ID NO: 738.								
Name	Len	cTP	mTP	SP	other	Loc	RC	TPlen
AtIAA14	228	0.116	0.087	0.047	0.879	—	2	—
cutoff		0.000	0.000	0.000	0.000			

Abbreviations:
Len, Length;
cTP, Chloroplastic transit peptide;
mTP, Mitochondrial transit peptide,
SP, Secretory pathway signal peptide,
other, Other subcellular targeting,
Loc, Predicted Location;
RC, Reliability class;
TPlen, Predicted transit peptide length.

Many other algorithms can be used to perform such analyses, including:
ChloroP 1.1 hosted on the server of the Technical University of Denmark;
Protein Prowler Subcellular Localisation Predictor version 1.2 hosted on the server of the Institute for Molecular Bioscience, University of Queensland, Brisbane, Australia;
PENCE Proteome Analyst PA-GOSUB 2.5 hosted on the server of the University of Alberta, Edmonton, Alberta, Canada;
TMHMM, hosted on the server of the Technical University of Denmark
PSORT (URL: psort.org)
PLOC (Park and Kanehisa, Bioinformatics, 19, 1656-1663, 2003).
PSORT analysis predicts a nuclear localisation, which is in agreement with the data from the literature (Fukaki et al., 2002).

Example 6

Subcellular Localisation Prediction of the Polypeptide Sequences Useful in Performing the Methods of the Invention

6.1. MYB91 Like Transcription Factor (MYB91)

Experimental methods for protein localization range from immunolocalization to tagging of proteins using green fluorescent protein (GFP) or beta-glucuronidase (GUS). Such methods to identify subcellular compartmentalisation of GRF polypeptides are well known in the art.
A predicted nuclear localisation signal (NLS) can be found by multiple sequence alignment, followed by eye inspection, in the polypeptide sequences of Table A2. An NLS is one or more short sequences of positively charged lysines or arginines.
Computational prediction of protein localisation from sequence data was performed. Among algorithms well known to a person skilled in the art are available at the ExPASy Proteomics tools hosted by the Swiss Institute for Bioinformatics, for example, PSort, TargetP, ChloroP, LocTree, Predotar, LipoP, MITOPROT, PATS, PTS1, SignalP, TMHMM, TMPred, and others.

The PSort algorithm predicts a nuclear subcellular localization for a MYB91 polypeptide as represented by SEQ ID NO: 221, as highest probability (0.088). In addition, two putative NLS are predicted:

Found: pos: 81 (3) KK IAAEVPGRTA KRLGK
Found: pos: 273 (3) RR VELQLESERS CRRRE

Example 7

Assay Related to the Polypeptide Sequences Useful in Performing the Methods of the Invention

7.1. MYB91 Like Transcription Factor (MYB91)
MYB91 polypeptides useful in the methods of the present invention (at least in their native form) typically, but not necessarily, have transcriptional regulatory activity and capacity to interact with other proteins. DNA-binding activity and protein-protein interactions may readily be determined in vitro or in vivo using techniques well known in the art (for example in Current Protocols in Molecular Biology, Volumes 1 and 2, Ausubel et al. (1994), Current Protocols). MYB91 polypeptides contain two Myb DNA-binding domain (InterPro accession IPR014778).
7.2. Gibberellic Acid-Stimulated *Arabidopsis* (GASA)
Transgenic plants expressing GASA polypeptides (at least in their native form) may have enhanced tolerance to heat stress. A thermotolerance assay is described by Ko et al. (2007): to examine the heat stress test response in seed germination, seeds are sown on water-saturated filter paper. They are left to imbibe at room temperature for 18 h, transferred to 50° C., and subjected to 3 h of heat treatment. Thereafter they are transferred to 22° C. Cotyledon emergence is determined after 5 days. Experiments are done in triplicate for each line (30 seeds each). To assess heat tolerance assay, seeds are germinated on normal MS (Murashige & Skoog salt mixture) medium. Seven-day-old seedlings are exposed to 50° C. for 2.5 h, and the surviving plants are scored 10 days after returning to normal growth conditions. Experiments were done in triplicate for each line (40 seeds each). Wild type plants are used as controls.
7.3. IAA14 Polypeptides
IAA14 is reported to interact with ARF7 and ARF19 in a yeast two-hybrid system (Fukaki et al., 2005): The cDNA fragments encoding the C-terminus of *Arabidopsis* ARF5 (amino acids 778-902), ARF7 (amino acids 1031-1164) and ARF19 (amino acids 952-1086) are amplified from a flower cDNA library using the following primer sets: 5'-agaattcAATAGTAAAGGCTCATC ATGGCAG-3' and 5'-agtcgacGTTACATTATGAAACAGAAGTCTTAAGATCG-3' for ARF5, 5'-agtcgacaAGCTCAGACTCAGCGAATGCG-3' and 5'-cagtcgacTCACCGGTTAAACGAA GTGGC-3' for ARF7, and 5'-gagaattcAATCAGACTCAACGAATGCG-3' and 5'-agtcgac CTATCTGTTGAAAGAAGCTGCAGC-3' for ARF19.
The full-length IAA14 open reading frame is amplified using two primers, 5'-cgaattcAT GAACCTTAAGGAGACG-GAGC-3' and 5'-tgtcgacTCATGATCTGTCTTGAAC-TCTCC-3'. PCR products are subcloned into pCR-Blunt II TOPO (Invitrogen, Carlsbad, Calif., USA) and are sequenced before in-frame insertion into pAD-GAL4-2.1 or pBD-GAL4 Cam (Stratagene, Calif., USA) via EcoRI/SalI (IAA14, ARF5 and ARF19) or SalI (ARF7) sites. Constructs are next introduced into *Saccharomyces cerevisiae* Y190 cells, and transformants are subjected to assays for beta-galactosidase activity as previously described (Kaiser et al., Methods in Yeast

Genetics: A Cold Spring Harbor Laboratory Course Manual.
Cold Spring Harbor, N.Y.: Cold Spring Harbor Laboratory
Press, 1994).

Example 8

Cloning of the Nucleic Acid Sequence Used in the Methods of the Invention

8.1. Aspartate AminoTransferase (ASPAT)

The nucleic acid sequence used in the methods of the invention was amplified by PCR using as template a custom-made cDNA library from either *Arabidopsis thaliana* seedlings or from *Oryza sativa* (in pCMV Sport 6.0; Invitrogen, Paisley, UK). PCR was performed using Hifi Taq DNA polymerase in standard conditions, using 200 ng of template in a 50 µl PCR mix. The cDNA library and primers used are given in Table F1.

TABLE F1

ORF in SEQ ID NO:	cDNA library	Primer forward (sense)	Primer reverse (complementary)
SEQ ID NO: 3	<i>Oryza sativa</i>	Ggggacaagttt gtacaaaaaagc aggcttaacaa tggcgtcgtcgt cc	Ggggaccactt tgtacaagaaa gctgggtatgc taccatcattc acttca
SEQ ID NO: 5	<i>Arabidopsis thaliana</i>	Ggggacaagttt gtacaaaaaagc aggcttaacaa tggattccgtct tctctaac	Ggggaccactt tgtacaagaaa gctgggtataaa atgtatggtcg ctagtt
SEQ ID NO: 7	<i>Arabidopsis thaliana</i>	Ggggacaagttt gtacaaaaaagc aggcttaacaa tgaaaactactc atttctcttc	Ggggaccactt tgtacaagaaa gctgggttggt gttcagtttct cagac
SEQ ID NO: 9	<i>Arabidopsis thaliana</i>	Ggggacaagttt gtacaaaaaagc aggcttaacaa tggcttctttaa tggtatct	Ggggaccactt tgtacaagaaa gctgggttggt atctactgaga tggaag

Primers include the AttB sites for Gateway recombination. The amplified PCR fragment was purified also using standard methods. The first step of the Gateway procedure, the BP reaction, was then performed, during which the PCR fragment recombined in vivo with the pDONR201 plasmid to produce, according to the Gateway terminology, an "entry clone", pASPAT. Plasmid pDONR201 was purchased from Invitrogen, as part of the Gateway® technology.

The entry clone comprising SEQ ID NO: 1 was then used in an LR reaction with a destination vector used for *Oryza sativa* transformation. This vector contained as functional elements within the T-DNA borders: a plant selectable marker; a screenable marker expression cassette; and a Gateway cassette intended for LR in vivo recombination with the nucleic acid sequence of interest already cloned in the entry clone. A rice GOS2 promoter (SEQ ID NO: 218) for constitutive specific expression was located upstream of this Gateway cassette.

After the LR recombination step, the resulting expression vector pGOS2::ASPAT (FIG. 3) was transformed into *Agrobacterium* strain LBA4044 according to methods well known in the art.

Similarly, expression vectors were generated comprising the following features (Table F2):

TABLE F2

Vector	Promoter	ASPT nucleic acid (Longest ORF in SEQ ID NO:)
ExprVect1	pPR (SEQ ID NO: 219)	SEQ ID NO: 3
ExprVect2	pGOS2 (SEQ ID NO: 218)	SEQ ID NO: 5
ExprVect3	pPR (SEQ ID NO: 219)	SEQ ID NO: 5
ExprVect4	pGOS2 (SEQ ID NO: 218)	SEQ ID NO: 7
ExprVect5	pGOS2 (SEQ ID NO: 218)	SEQ ID NO: 9

8.2. MYB91 Like Transcription Factor (MYB91)

Unless otherwise stated, recombinant DNA techniques are performed according to standard protocols described in (Sambrook (2001) Molecular Cloning: a laboratory manual, 3rd Edition Cold Spring Harbor Laboratory Press, CSH, New York) or in Volumes 1 and 2 of Ausubel et al. (1994), Current Protocols in Molecular Biology, Current Protocols. Standard materials and methods for plant molecular work are described in Plant Molecular Biology Labfax (1993) by R. D. D. Croy, published by BIOS Scientific Publications Ltd (UK) and Blackwell Scientific Publications (UK).

The *Populus trichocarpa* nucleic acid sequence encoding a MYB91 polypeptide sequence as represented by SEQ ID NO: 221 was amplified by PCR using as template a cDNA bank constructed using RNA from tomato plants at different developmental stages. The following primers, which include the AttB sites for Gateway recombination, were used for PCR amplification:

- 1) prm11884 (SEQ ID NO: 271, sense):
5' -GGGGACAAGTTTGTACAAAAAAGCAGGCTTAACCAATGAAGGAGA
GGCAGCGT-3'
- 2) prm11885 (SEQ ID NO: 272, reverse,
complementary):
5' -GGGGACCACTTTGTACAAAGAAAGCTGGGTGACCTGATACAGCTGG
ACGTA-3'

PCR was performed using Hifi Taq DNA polymerase in standard conditions. A PCR fragment of the expected length (including attB sites) was amplified and purified also using standard methods. The first step of the Gateway procedure, the BP reaction, was then performed, during which the PCR fragment recombined in vivo with the pDONR201 plasmid to produce, according to the Gateway terminology, an "entry clone". Plasmid pDONR201 was purchased from Invitrogen, as part of the Gateway® technology.

The entry clone comprising SEQ ID NO: 220 was subsequently used in an LR reaction with a destination vector used for *Oryza sativa* transformation. This vector contained as functional elements within the T-DNA borders: a plant selectable marker; a screenable marker expression cassette; and a Gateway cassette intended for LR in vivo recombination with the nucleic acid sequence of interest already cloned in the entry clone. A rice GOS2 promoter (SEQ ID NO: 53) for constitutive expression was located upstream of this Gateway cassette.

After the LR recombination step, the resulting expression vector pGOS2::MYB91 (FIG. 6) for constitutive expression, was transformed into *Agrobacterium* strain LBA4044 according to methods well known in the art.

8.3. Gibberellic Acid-Stimulated *Arabidopsis* (GASA)

a) Cloning of Tomato GASA:

The tomato nucleic acid sequence used in the methods of the invention was amplified by PCR using as template a custom-made *Solanum lycopersicum* seedlings cDNA library (in pCMV Sport 6.0; Invitrogen, Paisley, UK). PCR was performed using Hifi Taq DNA polymerase in standard conditions, using 200 ng of template in a 50 µl PCR mix. The primers used were prm10623 (SEQ ID NO: 286; sense, start

codon in bold): 5'-ggggacaagttgtacaaaaaagc aggccttaacaatg-gagaagacacttagctta-3' and prm10624 (SEQ ID NO: 287; reverse, complementary): 5'-ggggaccactttgtacaagaaagctggg-tatatatgattaagggcatttt-3', which include the AttB sites for Gateway recombination. The amplified PCR fragment was purified also using standard methods. The first step of the Gateway procedure, the BP reaction, was then performed, during which the PCR fragment recombined in vivo with the pDONR201 plasmid to produce, according to the Gateway terminology, an "entry clone", pGASA. Plasmid pDONR201 was purchased from Invitrogen, as part of the Gateway® technology.

The entry clone comprising SEQ ID NO: 275 was then used in an LR reaction with a destination vector used for *Oryza sativa* transformation. This vector contained as functional elements within the T-DNA borders: a plant selectable marker; a screenable marker expression cassette; and a Gateway cassette intended for LR in vivo recombination with the nucleic acid sequence of interest already cloned in the entry clone. A rice GOS2 promoter (SEQ ID NO: 290) for constitutive specific expression was located upstream of this Gateway cassette.

After the LR recombination step, the resulting expression vector pGOS2::GASA (FIG. 3) was transformed into *Agrobacterium* strain LBA4044 according to methods well known in the art.

b) Cloning of Poplar GASA

The poplar nucleic acid sequence used in the methods of the invention was amplified by PCR using as template a custom-made poplar seedlings cDNA library (in pCMV Sport 6.0; Invitrogen, Paisley, UK). PCR was performed using Hifi Taq DNA polymerase in standard conditions, using 200 ng of template in a 50 µl PCR mix. The primers used were prm10625 (SEQ ID NO: 288; sense, start codon in bold): 5'-gggacaagttgtacaaaaaagcaggcctt aacaatgaagaagctcttctgtct-3' and prm10626 (SEQ ID NO: 289; reverse, complementary): 5'-ggggaccactttgtacaagaagctggg-tacatgcacatcttgactgtct-3', which include the AttB sites for Gateway recombination. The amplified PCR fragment was purified also using standard methods, and the further cloning procedure was as described above, including use of the rice GOS2 promoter.

8.4. Auxin/Indoleacetic Acid Genes (AUX/IAA)

The nucleic acid sequence used in the methods of the invention was amplified by PCR using as template a custom-made *Oryza sativa* seedlings cDNA library (in pCMV Sport 6.0; Invitrogen, Paisley, UK). PCR was performed using Hifi Taq DNA polymerase in standard conditions, using 200 ng of template in a 50 µl PCR mix with a set of primer complementary to the first and last 20 nucleotides of SEQ ID NO: 431. The sequence of the forward primer used in the PCR can be represented by SEQ ID NO: 667 and the reverse primer by SEQ ID NO: 668. The amplified PCR fragment was purified also using standard methods. The first step of the Gateway procedure, the BP reaction, was then performed, during which the PCR fragment recombined in vivo with the pDONR201 plasmid to produce, according to the Gateway terminology, an "entry clone", p AUX/IAA. Plasmid pDONR201 was purchased from Invitrogen, as part of the Gateway® technology.

The entry clone comprising SEQ ID NO: 431 was then used in an LR reaction with a destination vector used for *Oryza sativa* transformation. This vector contained as functional elements within the T-DNA borders: a plant selectable marker; a screenable marker expression cassette; and a Gateway cassette intended for LR in vivo recombination with the nucleic acid sequence of interest already cloned in the entry clone. A rice GOS2 promoter (SEQ ID NO: 669) for constitutive specific expression was located upstream of this Gateway cassette.

After the LR recombination step, the resulting expression vector pGOS2:: AUX/IAA (FIG. 12) was transformed into *Agrobacterium* strain LBA4044 according to methods well known in the art.

8.5. IAA14 Polypeptides

The nucleic acid sequence used in the methods of the invention was amplified by PCR using as template a custom-made *Arabidopsis thaliana* seedlings cDNA library (in pCMV Sport 6.0; Invitrogen, Paisley, UK). PCR was performed using Hifi Taq DNA polymerase in standard conditions, using 200 ng of template in a 50 µl PCR mix. The primers used were prm07273 (SEQ ID NO: 745; sense, start codon in bold): 5'-ggggacaagttgtacaaaaaagcagg cttacaacat-gaaccttaaggagacggag-3' and prm07274 (SEQ ID NO: 746; reverse, complementary): 5'-ggggaccactttgtacaa-gaaagctgggtcaatgcattgtctctttt-3', which include the AttB sites for Gateway recombination. The amplified PCR fragment was purified also using standard methods. The first step of the Gateway procedure, the BP reaction, was then performed, during which the PCR fragment recombined in vivo with the pDONR201 plasmid to produce, according to the Gateway terminology, an "entry clone", pIAA14-like. Plasmid pDONR201 was purchased from Invitrogen, as part of the Gateway® technology.

The entry clone comprising SEQ ID NO: 737 was then used in an LR reaction with a destination vector used for *Oryza sativa* transformation. This vector contained as functional elements within the T-DNA borders: a plant selectable marker; a screenable marker expression cassette; and a Gateway cassette intended for LR in vivo recombination with the nucleic acid sequence of interest already cloned in the entry clone. A rice HMGP promoter (SEQ ID NO: 747) for weak constitutive expression was located upstream of this Gateway cassette.

After the LR recombination step, the resulting expression vector pHMGP::IAA14-like (FIG. 16) was transformed into *Agrobacterium* strain LBA4044 according to methods well known in the art.

Example 9

Plant Transformation

Rice Transformation

The *Agrobacterium* containing the expression vector was used to transform *Oryza sativa* plants. Mature dry seeds of the rice japonica cultivar Nipponbare were dehusked. Sterilization was carried out by incubating for one minute in 70% ethanol, followed by 30 minutes in 0.2% HgCl₂, followed by a 6 times 15 minutes wash with sterile distilled water. The sterile seeds were then germinated on a medium containing 2,4-D (callus induction medium). After incubation in the dark for four weeks, embryogenic, scutellum-derived calli were excised and propagated on the same medium. After two weeks, the calli were multiplied or propagated by subculture on the same medium for another 2 weeks. Embryogenic callus pieces were sub-cultured on fresh medium 3 days before co-cultivation (to boost cell division activity).

Agrobacterium strain LBA4404 containing the expression vector was used for co-cultivation. *Agrobacterium* was inoculated on AB medium with the appropriate antibiotics and cultured for 3 days at 28° C. The bacteria were then collected and suspended in liquid co-cultivation medium to a density (OD₆₀₀) of about 1. The suspension was then transferred to a Petri dish and the calli immersed in the suspension for 15 minutes. The callus tissues were then blotted dry on a filter paper and transferred to solidified, co-cultivation medium and incubated for 3 days in the dark at 25° C. Co-cultivated calli were grown on 2,4-D-containing medium for 4 weeks in the dark at 28° C. in the presence of a selection agent. During

this period, rapidly growing resistant callus islands developed. After transfer of this material to a regeneration medium and incubation in the light, the embryogenic potential was released and shoots developed in the next four to five weeks. Shoots were excised from the calli and incubated for 2 to 3 weeks on an auxin-containing medium from which they were transferred to soil. Hardened shoots were grown under high humidity and short days in a greenhouse.

Approximately 35 independent T0 rice transformants were generated for one construct. The primary transformants were transferred from a tissue culture chamber to a greenhouse. After a quantitative PCR analysis to verify copy number of the T-DNA insert, only single copy transgenic plants that exhibit tolerance to the selection agent were kept for harvest of T1 seed. Seeds were then harvested three to five months after transplanting. The method yielded single locus transformants at a rate of over 50% (Aldemita and Hodges 1996, Chan et al. 1993, Hiei et al. 1994).

Corn Transformation

Transformation of maize (*Zea mays*) is performed with a modification of the method described by Ishida et al. (1996) Nature Biotech 14(6): 745-50. Transformation is genotype-dependent in corn and only specific genotypes are amenable to transformation and regeneration. The inbred line A188 (University of Minnesota) or hybrids with A188 as a parent are good sources of donor material for transformation, but other genotypes can be used successfully as well. Ears are harvested from corn plant approximately 11 days after pollination (DAP) when the length of the immature embryo is about 1 to 1.2 mm. Immature embryos are cocultivated with *Agrobacterium tumefaciens* containing the expression vector, and transgenic plants are recovered through organogenesis. Excised embryos are grown on callus induction medium, then maize regeneration medium, containing the selection agent (for example imidazolinone but various selection markers can be used). The Petri plates are incubated in the light at 25° C. for 2-3 weeks, or until shoots develop. The green shoots are transferred from each embryo to maize rooting medium and incubated at 25° C. for 2-3 weeks, until roots develop. The rooted shoots are transplanted to soil in the greenhouse. T1 seeds are produced from plants that exhibit tolerance to the selection agent and that contain a single copy of the T-DNA insert.

Wheat Transformation

Transformation of wheat is performed with the method described by Ishida et al. (1996) Nature Biotech 14(6): 745-50. The cultivar Bobwhite (available from CIMMYT, Mexico) is commonly used in transformation. Immature embryos are co-cultivated with *Agrobacterium tumefaciens* containing the expression vector, and transgenic plants are recovered through organogenesis. After incubation with *Agrobacterium*, the embryos are grown in vitro on callus induction medium, then regeneration medium, containing the selection agent (for example imidazolinone but various selection markers can be used). The Petri plates are incubated in the light at 25° C. for 2-3 weeks, or until shoots develop. The green shoots are transferred from each embryo to rooting medium and incubated at 25° C. for 2-3 weeks, until roots develop. The rooted shoots are transplanted to soil in the greenhouse. T1 seeds are produced from plants that exhibit tolerance to the selection agent and that contain a single copy of the T-DNA insert.

Soybean Transformation

Soybean is transformed according to a modification of the method described in the Texas A&M patent U.S. Pat. No. 5,164,310. Several commercial soybean varieties are amenable to transformation by this method. The cultivar Jack (available from the Illinois Seed foundation) is commonly used for transformation. Soybean seeds are sterilised for in vitro sowing. The hypocotyl, the radicle and one cotyledon

are excised from seven-day old young seedlings. The epicotyl and the remaining cotyledon are further grown to develop axillary nodes. These axillary nodes are excised and incubated with *Agrobacterium tumefaciens* containing the expression vector. After the cocultivation treatment, the explants are washed and transferred to selection media. Regenerated shoots are excised and placed on a shoot elongation medium. Shoots no longer than 1 cm are placed on rooting medium until roots develop. The rooted shoots are transplanted to soil in the greenhouse. T1 seeds are produced from plants that exhibit tolerance to the selection agent and that contain a single copy of the T-DNA insert.

Rapeseed/Canola Transformation

Cotyledonary petioles and hypocotyls of 5-6 day old young seedling are used as explants for tissue culture and transformed according to Babic et al. (1998, Plant Cell Rep 17: 183-188). The commercial cultivar Westar (Agriculture Canada) is the standard variety used for transformation, but other varieties can also be used. Canola seeds are surface-sterilized for in vitro sowing. The cotyledon petiole explants with the cotyledon attached are excised from the in vitro seedlings, and inoculated with *Agrobacterium* (containing the expression vector) by dipping the cut end of the petiole explant into the bacterial suspension. The explants are then cultured for 2 days on MSBAP-3 medium containing 3 mg/l BAP, 3% sucrose, 0.7% Phytagar at 23° C., 16 hr light. After two days of co-cultivation with *Agrobacterium*, the petiole explants are transferred to MSBAP-3 medium containing 3 mg/l BAP, cefotaxime, carbenicillin, or timentin (300 mg/l) for 7 days, and then cultured on MSBAP-3 medium with cefotaxime, carbenicillin, or timentin and selection agent until shoot regeneration. When the shoots are 5-10 mm in length, they are cut and transferred to shoot elongation medium (MSBAP-0.5, containing 0.5 mg/l BAP). Shoots of about 2 cm in length are transferred to the rooting medium (MS0) for root induction. The rooted shoots are transplanted to soil in the greenhouse. T1 seeds are produced from plants that exhibit tolerance to the selection agent and that contain a single copy of the T-DNA insert.

Alfalfa Transformation

A regenerating clone of alfalfa (*Medicago sativa*) is transformed using the method of (McKersie et al., 1999 Plant Physiol 119: 839-847). Regeneration and transformation of alfalfa is genotype dependent and therefore a regenerating plant is required. Methods to obtain regenerating plants have been described. For example, these can be selected from the cultivar Rangelander (Agriculture Canada) or any other commercial alfalfa variety as described by Brown DCW and A Atanassov (1985, Plant Cell Tissue Organ Culture 4: 111-112). Alternatively, the RA3 variety (University of Wisconsin) has been selected for use in tissue culture (Walker et al., 1978 Am J Bot 65:654-659). Petiole explants are cocultivated with an overnight culture of *Agrobacterium tumefaciens* C58C1 pMP90 (McKersie et al., 1999 Plant Physiol 119: 839-847) or LBA4404 containing the expression vector. The explants are cocultivated for 3 d in the dark on SH induction medium containing 288 mg/L Pro, 53 mg/L thioproline, 4.35 g/L K₂SO₄, and 100 µm acetosyringone. The explants are washed in half-strength Murashige-Skoog medium (Murashige and Skoog, 1962) and plated on the same SH induction medium without acetosyringone but with a suitable selection agent and suitable antibiotic to inhibit *Agrobacterium* growth. After several weeks, somatic embryos are transferred to BOi2Y development medium containing no growth regulators, no antibiotics, and 50 g/L sucrose. Somatic embryos are subsequently germinated on half-strength Murashige-Skoog medium. Rooted seedlings were transplanted into pots and grown in a greenhouse. T1 seeds are produced from plants that exhibit tolerance to the selection agent and that contain a single copy of the T-DNA insert.

Cotton Transformation

Cotton is transformed using *Agrobacterium tumefaciens* according to the method described in U.S. Pat. No. 5,159,135. Cotton seeds are surface sterilised in 3% sodium hypochlorite solution during 20 minutes and washed in distilled water with 500 µg/ml cefotaxime. The seeds are then transferred to SH-medium with 50 µg/ml benomyl for germination. Hypocotyls of 4 to 6 days old seedlings are removed, cut into 0.5 cm pieces and are placed on 0.8% agar. An *Agrobacterium* suspension (approx. 108 cells per ml, diluted from an overnight culture transformed with the gene of interest and suitable selection markers) is used for inoculation of the hypocotyl explants. After 3 days at room temperature and lighting, the tissues are transferred to a solid medium (1.6 g/l Gelrite) with Murashige and Skoog salts with B5 vitamins (Gamborg et al., Exp. Cell Res. 50:151-158 (1968)), 0.1 mg/l 2,4-D, 0.1 mg/l 6-furfurylaminopurine and 750 µg/ml MgCL₂, and with 50 to 100 µg/ml cefotaxime and 400-500 µg/ml carbenicillin to kill residual bacteria. Individual cell lines are isolated after two to three months (with subcultures every four to six weeks) and are further cultivated on selective medium for tissue amplification (30° C., 16 hr photoperiod). Transformed tissues are subsequently further cultivated on non-selective medium during 2 to 3 months to give rise to somatic embryos. Healthy looking embryos of at least 4 mm length are transferred to tubes with SH medium in fine vermiculite, supplemented with 0.1 mg/l indole acetic acid, 6 furfurylaminopurine and gibberellic acid. The embryos are cultivated at 30° C. with a photoperiod of 16 hrs, and plantlets at the 2 to 3 leaf stage are transferred to pots with vermiculite and nutrients. The plants are hardened and subsequently moved to the greenhouse for further cultivation.

Example 10

Phenotypic Evaluation Procedure

10.1 Evaluation Setup

Approximately 35 independent T0 rice transformants were generated. The primary transformants were transferred from a tissue culture chamber to a greenhouse for growing and harvest of T1 seed. Events, of which the T1 progeny segregated 3:1 for presence/absence of the transgene, were retained. For each of these events, approximately 10 T1 seedlings containing the transgene (hetero- and homo-zygotes) and approximately 10 T1 seedlings lacking the transgene (nullizygotes) were selected by monitoring visual marker expression. The transgenic plants and the corresponding nullizygotes were grown side-by-side at random positions. Greenhouse conditions were of short days (12 hours light), 28° C. in the light and 22° C. in the dark, and a relative humidity of 70%. Plants grown under non-stress conditions were watered at regular intervals to ensure that water and nutrients were not limiting and to satisfy plant needs to complete growth and development.

T1 events were further evaluated in the T2 generation following the same evaluation procedure as for the T1 generation but with more individuals per event. From the stage of sowing until the stage of maturity the plants were passed several times through a digital imaging cabinet. At each time point digital images (2048×1536 pixels, 16 million colours) were taken of each plant from at least 6 different angles.

Drought Screen

Plants from T2 seeds are grown in potting soil under normal conditions until they approached the heading stage. They are then transferred to a "dry" section where irrigation is withheld. Humidity probes are inserted in randomly chosen pots to monitor the soil water content (SWC). When SWC goes below certain thresholds, the plants are automatically re-watered continuously until a normal level is reached again.

The plants are then re-transferred again to normal conditions. The rest of the cultivation (plant maturation, seed harvest) is the same as for plants not grown under abiotic stress conditions. Growth and yield parameters are recorded as detailed for growth under normal conditions.

Nitrogen Use Efficiency Screen

Rice plants from T2 seeds are grown in potting soil under normal conditions except for the nutrient solution. The pots are watered from transplantation to maturation with a specific nutrient solution containing reduced N nitrogen (N) content, usually between 7 to 8 times less. The rest of the cultivation (plant maturation, seed harvest) is the same as for plants not grown under abiotic stress. Growth and yield parameters are recorded as detailed for growth under normal conditions.

Salt Stress Screen

Plants are grown on a substrate made of coco fibers and argex (3 to 1 ratio). A normal nutrient solution is used during the first two weeks after transplanting the plantlets in the greenhouse. After the first two weeks, 25 mM of salt (NaCl) is added to the nutrient solution, until the plants are harvested. Seed-related parameters are then measured.

10.2 Statistical Analysis: F Test

A two factor ANOVA (analysis of variants) was used as a statistical model for the overall evaluation of plant phenotypic characteristics. An F test was carried out on all the parameters measured of all the plants of all the events transformed with the gene of the present invention.

The F test was carried out to check for an effect of the gene over all the transformation events and to verify for an overall effect of the gene, also known as a global gene effect. The threshold for significance for a true global gene effect was set at a 5% probability level for the F test. A significant F test value points to a gene effect, meaning that it is not only the mere presence or position of the gene that is causing the differences in phenotype.

Because two experiments with overlapping events were carried out, a combined analysis was performed. This is useful to check consistency of the effects over the two experiments, and if this is the case, to accumulate evidence from both experiments in order to increase confidence in the conclusion. The method used was a mixed-model approach that takes into account the multilevel structure of the data (i.e. experiment—event—segregants). P values were obtained by comparing likelihood ratio test to chi square distributions.

10.3 Parameters Measured

Biomass-Related Parameter Measurement

From the stage of sowing until the stage of maturity the plants were passed several times through a digital imaging cabinet. At each time point digital images (2048×1536 pixels, 16 million colours) were taken of each plant from at least 6 different angles.

The plant aboveground area (or leafy biomass) was determined by counting the total number of pixels on the digital images from aboveground plant parts discriminated from the background. This value was averaged for the pictures taken on the same time point from the different angles and was converted to a physical surface value expressed in square mm by calibration. Experiments show that the aboveground plant area measured this way correlates with the biomass of plant parts above ground. The above ground area is the area measured at the time point at which the plant had reached its maximal leafy biomass. The early vigour is the plant (seedling) aboveground area three weeks post-germination. Increase in root biomass is expressed as an increase in total root biomass (measured as maximum biomass of roots observed during the lifespan of a plant); or as an increase in the root/shoot index (measured as the ratio between root mass and shoot mass in the period of active growth of root and shoot).

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Early vigour was determined by counting the total number of pixels from aboveground plant parts discriminated from the background. This value was averaged for the pictures taken on the same time point from different angles and was converted to a physical surface value expressed in square mm by calibration. The results described below are for plants three weeks post-germination.

Seed-Related Parameter Measurements

The mature primary panicles were harvested, counted, bagged, barcode-labelled and then dried for three days in an oven at 37° C. The panicles were then threshed and all the seeds were collected and counted. The filled husks were separated from the empty ones using an air-blowing device. The empty husks were discarded and the remaining fraction was counted again. The filled husks were weighed on an analytical balance. The number of filled seeds was determined by counting the number of filled husks that remained after the separation step. The total seed yield was measured by weighing all filled husks harvested from a plant. Total seed number per plant was measured by counting the number of husks harvested from a plant. Thousand Kernel Weight (TKW) is extrapolated from the number of filled seeds counted and their total weight. The Harvest Index (HI) in the present invention is defined as the ratio between the total seed yield and the above ground area (mm²), multiplied by a factor 10⁶. The total number of flowers per panicle as defined in the present invention is the ratio between the total number of seeds and the number of mature primary panicles. The seed fill rate as defined in the present invention is the proportion (expressed as a %) of the number of filled seeds over the total number of seeds (or florets).

Examples 11

Results of the Phenotypic Evaluation of the Transgenic Plants

11.1. Aspartate Amino Transferase (ASPAT)

The results of the evaluation of transgenic rice plants in the T2 generation and expressing a nucleic acid comprising the longest Open Reading Frame in SEQ ID NO: 1 under the control of the rice GOS2 promoter in non-stress conditions are presented below (Table G1). See previous Examples for details on the generations of the transgenic plants. An increase of at least 5% was observed for aboveground biomass (AreaMax), emergence, seed yield (totalwgseeds), number of filled seeds (nrfilledseed), fill rate (fillrate), and plant height (HeightMax) (Table G1).

TABLE G1

Phenotype transgenic plants transformed with pGOS2::ASAPT.	
Parameter	% increase in transgenic plants versus control plants
AreaMax	7.4
totalwgseeds	11.8
nrfilledseed	9.3
fillrate	5.0
HeightMax	5.0

The results of the evaluation of transgenic rice plants in the T1 generation and expressing a nucleic acid comprising the longest Open Reading Frame in SEQ ID NO: 5 under the control of the rice GOS2 promoter in non-stress conditions are presented below (Table G2). See previous Examples for details on the generations of the transgenic plants. An increase of at least 5% was observed for plant height (HeightMax).

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TABLE G2

Phenotype transgenic plants transformed with ExprVect2.	
Parameter	% increase in transgenic plants versus control plants
Plant height	5.2

The results of the evaluation of transgenic rice plants in the T1 generation and expressing a nucleic acid comprising the longest Open Reading Frame in SEQ ID NO: 5 under the control of the rice PR promoter in non-stress conditions are presented below (Table G3). See previous Examples for details on the generations of the transgenic plants. An increase of at least 5% was observed for aboveground biomass (AreaMax), emergence vigour (EmerVigor), seed yield (totalwgseeds), number of filled seeds (nrfilledseed), number of flowers per panicle (flowerperpan), number of first panicle (firstpan), total number of seeds (nrtotalseed) and plant height (HeightMax).

TABLE G3

Phenotype transgenic plants transformed with the expression vector ExprVect3.	
Parameter	% increase in transgenic plants versus control plants
AreaMax	29.3
EmerVigor	49.8
totalwgseeds	31.2
nrfilledseed	32.0
flowerperpan	9.5
firstpan	15.8
nrtotalseed	26.8
HeightMax	11.6

The results of the evaluation of transgenic rice plants in the T2 generation and expressing a nucleic acid comprising the longest Open Reading Frame in SEQ ID NO: 5 under the control of the rice PR promoter in non-stress conditions are presented below (Table G4). See previous Examples for details on the generations of the transgenic plants. An increase of at least 5% was observed for aboveground biomass (AreaMax), emergence vigour (EmerVigor), total seed yield (totalwgseeds), number of filled seeds (nrfilledseed), number of flowers per panicle (flowerperpan), number of first panicle (firstpan), total number of seeds (nrtotalseed) and plant height (HeightMax).

TABLE G4

Phenotype transgenic plants transformed with the expression vector ExprVect3.	
Parameter	% increase in transgenic plants versus control plants
AreaMax	9.7
EmerVigor	17.8
totalwgseeds	24.4
nrfilledseed	23.3
fillrate	8.4
harvestindex	14.7
firstpan	10.8
nrtotalseed	14.9
HeightMax	5.3

The results of the evaluation of transgenic rice plants in the T1 generation and expressing a nucleic acid comprising the longest Open Reading Frame in SEQ ID NO: 3 under the control of the rice PR promoter in non-stress conditions are

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presented below (Table G5). See previous Examples for details on the generations of the transgenic plants. An increase of at least 5% was observed for seed yield (totalwgseeds), number of filled seeds (nrfilledseed), harvest index (harvestindex), and seed filling rate (fillrate).

TABLE G5

Phenotype transgenic plants transformed with the expression vector ExprVect1.	
Parameter	% increase in transgenic plants versus control plants
totalwgseeds	23.0
nrfilledseed	20.1
fillrate	9.9
harvestindex	13.8

The results of the evaluation of transgenic rice plants in the T1 generation and expressing a nucleic acid comprising the longest Open Reading Frame in SEQ ID NO: 9 under the control of the rice GOS2 promoter in non-stress conditions are presented below (Table G6). See previous Examples for details on the generations of the transgenic plants. An increase of at least 5% was observed for filled seeds (nrfilledseed) and harvest index (harvestindex).

TABLE G6

Phenotype transgenic plants transformed with the expression vector ExprVect5.	
Parameter	% increase in transgenic plants versus control plants
fillrate	6.6
harvestindex	6.0

The results of the evaluation of transgenic rice plants under non-stress conditions are presented below. An increase of at least 5% was observed for fill rate and harvest index.

11.2. MYB91 Like Transcription Factor (MYB91)

The results of the evaluation of T1 generation transgenic rice plants expressing the nucleic acid sequence encoding a MYB91 polypeptide as represented by SEQ ID NO: 221, under the control of a constitutive promoter, and grown under normal growth conditions, are presented below.

There was a significant increase in plant height, in harvest index (HI), and in Thousand Kernel Weight (TKW).

TABLE G7

Results of the evaluation of T1 generation transgenic rice plants expressing the nucleic acid sequence encoding a MYB91 polypeptide as represented by SEQ ID NO: 221, under the control of a promoter for constitutive expression.	
Trait	Overall average % increase in 4 events in the T2 generation
Plant height	3%
Harvest index	8%
Thousand kernel weight	6%

11.3. Gibberellic Acid-Stimulated *Arabidopsis* (GASA)

The results of the evaluation of transgenic rice plants expressing the tomato GASA nucleic acid under control of a medium strength constitutive promoter under non-stress conditions are presented below in Table G8.

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TABLE G8

overall increase (%) for yield parameters		
parameter	1 st evaluation	2 nd evaluation
Time to flower	2.1	3.5
Fill rate	10.4	8.3
Flowers per panicle	4.8	14.7

The flowering time was reduced compared to control plants, and there was an increase of more than 5% for fill rate and for the number of flowers per panicle.

The results of the evaluation of transgenic rice plants expressing the poplar GASA nucleic acid under control of a medium strength constitutive promoter under non-stress conditions are presented below in Table G9.

TABLE G9

overall increase (%) for yield parameters		
parameter	1 st evaluation	2 nd evaluation
Total weight of seeds	13.3	13.7
Harvest index	18.8	22.2
Thousand Kernel weight	4.2	2.9

11.4. Auxin/Indoleacetic Acid Genes (AUX/IAA)

The results of the evaluation of transgenic rice plants in the T2 generation and expressing a nucleic acid comprising the longest Open Reading Frame in SEQ ID NO: 431 under non-stress conditions are presented below. See previous Examples for details on the generations of the transgenic plants.

The results of the evaluation of transgenic rice plants under non-stress conditions are presented below (Table G10). An increase of at least 5% was observed for the number of filled seed per plant (nrfilledseed), harvest index (harvestindex) and seed yield (totalwgseeds).

TABLE G10

Yield-related trait	Percentage increase in transgenic plants compared to control plants
totalwgseeds	12.0
harvestindex	8.3
nrfilledseed	11.2

11.5. IAA14 Polypeptides

The results of the evaluation of T2 transgenic rice plants expressing the IAA14-like nucleic acid of SEQ ID NO: 738 under non-stress conditions are presented below (Table G11).

TABLE G11

Overall yield increase (in %) of transgenic plants expressing SEQ ID NO: 738	
Parameter	Overall increase
totalwgseeds	19.2
nrfilledseed	18.6
fillrate	18.8
harvestindex	21.1
HeightMax	5.5
GravityYMax	6.6

An increase was found for total weight of seeds, the number of filled seeds, for the fill rate (number of filled seeds divided by the total number of seeds and multiplied by 100), harvest index, height of the plant and the gravity center (indication of branching of plants). For each of the parameters listed in Table G11, the p-value was p<0.05.

SEQUENCE LISTING

The patent contains a lengthy "Sequence Listing" section. A copy of the "Sequence Listing" is available in electronic form from the USPTO web site (<http://seqdata.uspto.gov/?pageRequest=docDetail&DocID=US09062322B2>). An electronic copy of the "Sequence Listing" will also be available from the USPTO upon request and payment of the fee set forth in 37 CFR 1.19(b)(3).

The invention claimed is:

1. A method for enhancing yield-related traits in a plant relative to a control plant, comprising:
 - a) introducing and expressing in a plant a nucleic acid encoding an ASPAT (Aspartate Aminotransferase) polypeptide, wherein the nucleic acid is operably linked to a PR (Protochlorophyllide reductase) promoter and comprises a polynucleotide selected from the group consisting of:
 - (i) a polynucleotide comprising the nucleotide sequence of SEQ ID NO: 5;
 - (ii) a polynucleotide encoding a polypeptide comprising the amino acid sequence of SEQ ID NO: 6; and
 - (iii) a polynucleotide encoding a polypeptide comprising an amino acid sequence which has at least 95% overall sequence identity to the amino acid sequence of SEQ ID NO: 6;and
 - b) selecting a plant having enhanced yield-related traits relative to a control plant, wherein said enhanced yield-related traits comprise increased biomass and/or increased seed yield relative to a control plant.
2. The method of claim 1, wherein said enhanced yield-related traits are obtained under non-stress conditions.
 3. The method of claim 1, wherein said enhanced yield-related traits are obtained under conditions of drought stress, salt stress or nitrogen deficiency.
 4. The method of claim 1, wherein said PR promoter is a PR promoter from rice.
 5. A construct comprising:
 - (i) a nucleic acid encoding an ASPAT polypeptide;
 - (ii) one or more heterologous control sequences capable of driving expression of the nucleic acid of (i); and optionally
 - (iii) a transcription termination sequence,wherein said nucleic acid comprises a polynucleotide selected from the group consisting of:
 - (a) a polynucleotide comprising the nucleotide sequence of SEQ ID NO: 5;
 - (b) a polynucleotide encoding a polypeptide comprising the amino acid sequence of SEQ ID NO: 6; and
 - (c) a polynucleotide encoding a polypeptide comprising an amino acid sequence which has at least 95% overall sequence identity to the amino acid sequence of SEQ ID NO: 6;and wherein one of said control sequences is a PR promoter which is operably linked to the nucleic acid of (i).
 6. The construct of claim 5, wherein said PR promoter is a PR promoter from rice.
 7. A method for making a plant having increased biomass and/or increased seed yield relative to a control plant, comprising transforming the construct of claim 5 into a plant and selecting for a plant having increased biomass and/or increased seed yield relative to a control plant.
 8. A plant, plant part or plant cell transformed with the construct of claim 5.
 9. A method for the production of a transgenic plant having increased biomass and/or increased seed yield relative to a control plant, comprising:
 - (i) introducing and expressing in a plant a nucleic acid encoding an ASPAT polypeptide;
 - (ii) cultivating the plant under conditions promoting plant growth and development; and
 - (iii) selecting for a plant having increased biomass and/or increased seed yield relative to a control plant,wherein said nucleic acid is operably linked to a PR promoter and comprises a polynucleotide selected from the group consisting of:
 - (a) a polynucleotide comprising the nucleotide sequence of SEQ ID NO: 5;
 - (b) a polynucleotide encoding a polypeptide comprising the amino acid sequence of SEQ ID NO: 6; and
 - (c) a polynucleotide encoding a polypeptide comprising an amino acid sequence which has at least 95% overall sequence identity to the amino acid sequence of SEQ ID NO: 6.
 10. A transgenic plant comprising the construct of claim 5 and having increased biomass and/or increased seed yield relative to a control plant, wherein said increased biomass and/or increased seed yield is resulted from increased expression of the nucleic acid encoding an ASPAT polypeptide comprised in said construct or a transgenic plant cell comprising said construct and derived from said transgenic plant.
 11. The transgenic plant of claim 10, or a transgenic plant cell derived thereof, wherein said plant is a crop plant, a monocot or a cereal.
 12. Harvestable parts of the transgenic plant of claim 10, wherein said harvestable parts comprise a recombinant nucleic acid encoding said ASPAT polypeptide operably linked to a PR promoter, and wherein said harvestable parts are shoot biomass and/or seeds.
 13. Products derived from the transgenic plant of claim 10 and/or from harvestable parts of said transgenic plant, wherein said products comprise a recombinant nucleic acid encoding said ASPAT polypeptide operably linked to a PR promoter.
 14. The method of claim 1, wherein the plant is a crop plant, a monocot plant or a cereal.
 15. The method of claim 1, wherein the plant is rice, maize, wheat, barley, millet, rye, triticale, sorghum, emmer, spelt, *secale*, einkorn, teff, milo, or oats.
 16. The plant, plant part or plant cell of claim 8, wherein the plant is a crop plant, a monocot plant or a cereal, or wherein the plant part or plant cell is from a crop plant, a monocot plant or a cereal.

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17. The plant, plant part or plant cell of claim 8, wherein the plant is rice, maize, wheat, barley, millet, rye, triticale, sorghum, emmer, spelt, *secale*, einkorn, teff, milo, or oats, or wherein the plant part or plant cell is from a rice, maize, wheat, barley, millet, rye, triticale, sorghum, emmer, spelt, *secale*, einkorn, teff, milo, or oats plant.

18. The method of claim 9, wherein the plant is a crop plant, a monocot plant or a cereal.

19. The method of claim 9, wherein the plant is rice, maize, wheat, barley, millet, rye, triticale, sorghum, emmer, spelt, *secale*, einkorn, teff, milo, or oats.

20. The transgenic plant of claim 10, or a transgenic plant cell derived thereof, wherein said plant is rice, maize, wheat, barley, millet, rye, triticale, sorghum, emmer, spelt, *secale*, einkorn, teff, milo, or oats.

21. A method for increasing biomass and/or seed yield in a plant relative to a control plant, comprising:

- a) transforming the construct of claim 5 into a plant, plant cell, or plant part;
- b) selecting for a plant having increased biomass and/or seed yield relative to a control plant under non-stress growth conditions.

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22. The method of claim 21, wherein said plant has at least 5% increase in biomass and/or seed yield as compared to the control plant.

23. The plant, plant part or plant cell of claim 8, wherein the plant is rice, or wherein the plant part or plant cell is from a rice plant.

24. The plant, plant part or plant cell of claim 8, wherein the plant is maize, or wherein the plant part or plant cell is from a maize plant.

25. The plant, plant part or plant cell of claim 8, wherein the plant is wheat, or wherein the plant part or plant cell is from a wheat plant.

26. The transgenic plant of claim 10, or a transgenic plant cell derived thereof, wherein said plant is rice.

27. The transgenic plant of claim 10, or a transgenic plant cell derived thereof, wherein said plant is maize.

28. The transgenic plant of claim 10, or a transgenic plant cell derived thereof, wherein said plant is wheat.

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